

Light-driven catalytic regulation of enzymes at the interface with plasmonic nanomaterials

Original

Light-driven catalytic regulation of enzymes at the interface with plasmonic nanomaterials / RIBEIRO DE BARROS, Heloise; López-Gallego, Fernando; Liz-Marzán, Luis M.. - In: BIOCHEMISTRY. - ISSN 0006-2960. - 60:13(2020), pp. 991-998. [10.1021/acs.biochem.0c00447]

Availability:

This version is available at: 11583/2995213 since: 2024-12-12T09:35:28Z

Publisher:

American Chemical Society

Published

DOI:10.1021/acs.biochem.0c00447

Terms of use:

This article is made available under terms and conditions as specified in the corresponding bibliographic description in the repository

Publisher copyright

ACS postprint/Author's Accepted Manuscript

This document is the Accepted Manuscript version of a Published Work that appeared in final form in BIOCHEMISTRY, copyright © American Chemical Society after peer review and technical editing by the publisher. To access the final edited and published work see <http://dx.doi.org/10.1021/acs.biochem.0c00447>.

(Article begins on next page)

This is the post-print version of the following article: *Ribeiro de Barros, H; Lopez-Gallego, F; Liz-Marzán, LM., [Light-driven catalytic regulation of enzymes at the interface with plasmonic nanomaterials](#), *Biochemistry* 2021, 60, 13, 991–998*

DOI: [10.1021/acs.biochem.0c00447](https://doi.org/10.1021/acs.biochem.0c00447)

This article may be used for non-commercial purposes in accordance with ACS Terms and Conditions for Self-Archiving.

1 Light-driven catalytic regulation of enzymes at 2 the interface with plasmonic nanomaterials

3
4 *Heloise Ribeiro de Barros,^{a,b} Fernando López-Gallego,^{a,c,*} Luis M. Liz-Marzán^{a,c,d,*}*

5 ^a CIC biomaGUNE, Basque Research and Technology Alliance (BRTA), Paseo de
6 Miramón 182, 20014 Donostia – San Sebastián, Spain

7 ^b Institute of Chemistry, University of São Paulo, Av. Prof. Lineu Prestes, 748, Vila
8 Universitária, 05508-000 São Paulo, SP, Brazil.

9 ^c IKERBASQUE, Basque Foundation for Science, 48013 Bilbao, Spain

10 ^d Centro de Investigación Biomédica en Red, Bioingeniería, Biomateriales y
11 Nanomedicina (CIBER-BBN), Paseo de Miramón 182, 20014 Donostia – San Sebastián,
12 Spain

13

14 *Corresponding authors' email: flopez@cicbiomagune.es (F.L.-G.);

15 lizmarzan@cicbiomagune.es (L.M.L.-M.)

16

17 **ABSTRACT**

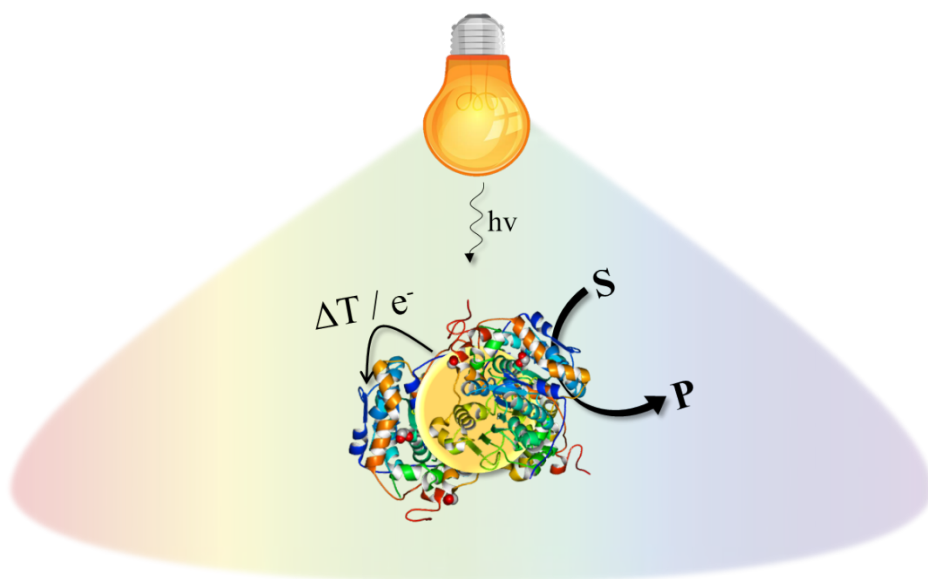
18 Regulation of enzymes is highly relevant toward orchestrating cell-free and stepwise
19 biotransformations, thereby maximizing their overall performance. Plasmonic
20 nanomaterials offer a great opportunity to tune the functionality of enzymes, through their
21 remarkable optical properties. Localized surface plasmon resonances (LSPR) can be used
22 to modify chemical transformations at the nanomaterial's surface, upon light irradiation.
23 Incident light can promote energetic processes, which may be related to an increase of
24 local temperature (photothermal effects), but also to effects triggered by generated hot-
25 spots or hot-electrons (photoelectronic effects). As a consequence, light irradiation of the
26 protein-nanomaterial interface affects enzyme functionality. To harness these effects to
27 finely and remotely regulate enzyme activity, the physicochemical features of the

28 nanomaterial, properties of the incident light and parameters governing molecular
29 interactions must be optimized. In this perspective article, we discuss relevant examples
30 that illustrate the use of plasmonic nanoparticles to control enzyme function through
31 LSPR excitation. Finally, we also highlight the importance of expanding the use of
32 plasmonic nanomaterials to the immobilization of multi-enzyme systems for light-driven
33 regulation of cell-free biosynthetic pathways. Although this concept is living its infancy,
34 we encourage the scientific community to advance in the development of novel light-
35 controlled biocatalytic plasmonic nanoconjugates and explore their application in
36 biosensing, applied biocatalysis, and biomedicine.

37

38

39 **ToC Graphic**



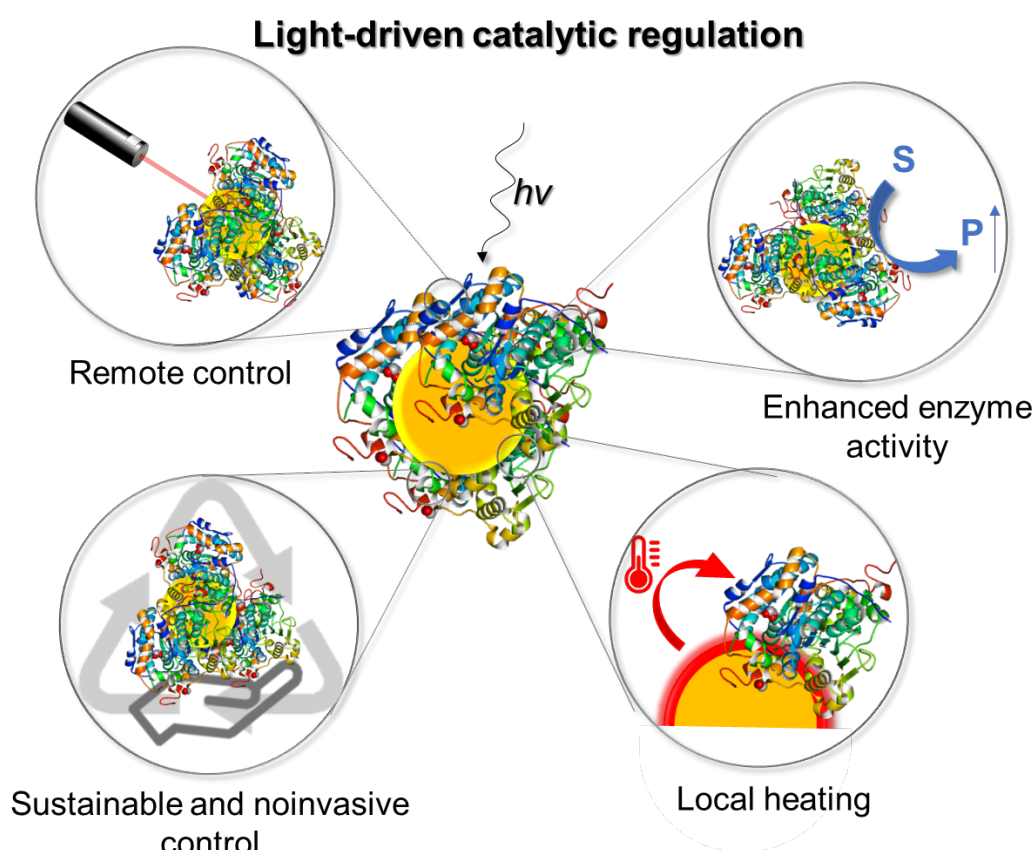
40

41

42 Remote control of extracellular enzyme functionality through external stimuli
43 provides access to more complex and concurrent cell-free biosynthetic routes. Enzyme
44 activity regulation additionally offers the possibility of orchestrating several enzymes to
45 simultaneously work in tuned cascade reactions, which maximizes the efficiency of
46 biosynthetic pathways. Inside the cell, enzymes are tightly regulated, both at genomic and
47 proteomic levels,¹ however such a regulation is lost when they are isolated. Therefore,
48 breakthroughs in this topic are expected to advance toward enzymatic designs with a finer
49 control for fully artificial applications. Light is one of the most efficient stimuli to
50 remotely control enzymatic activity. The bio-conjugation of enzymes to light-responsive
51 molecules and polymers has been the most common approximation hereto.²⁻⁴ Interfacing
52 enzymes onto plasmonic nanomaterials has been recently introduced as a promising
53 alternative tool to design bioconjugates with tailored properties for remote control of
54 biocatalysis. As any immobilization protocol, the conjugation of enzymes onto
55 nanoparticles must be designed to enhance both the activity and the stability of the
56 resulting bioconjugates. Hence, nanoparticle size and shape, as well as the conjugation
57 chemistry, are fundamental aspects to control the orientation, density and binding strength
58 that ultimately influence the functionality of the immobilized enzymes.^{5, 6} Not only the
59 immobilization of enzymes on plasmonic nanomaterials may enhance their functional
60 properties,^{7, 8} but optimization of the interactions between enzymes and plasmonic
61 nanoparticles additionally offers opportunities to develop nanoengineered materials for
62 light-controlled biocatalysis.^{9, 10} For example, combination of the remarkable biological
63 functions of enzymes and the unique optical properties of plasmonic nanomaterials can
64 contribute to applications including cancer therapy,¹¹⁻¹³ biosensing,¹⁴⁻¹⁶ applied
65 biocatalysis and biotransformations,^{17, 18} and intra- or extra-cellular nanosurgery.¹⁹⁻²¹

66 The suitability of plasmonic nanomaterials to tune enzyme activity arises from
67 their tunable localized surface plasmon resonances (LSPR), in the UV-Vis and near-
68 infrared wavelength ranges.²²⁻²⁴ Under incident light, conduction electrons in metal
69 nanoparticles display coherent collective oscillations, thereby generating intense
70 absorption and scattering at specific wavelengths, resulting in LSPR signals.^{25, 26} It is
71 well-known that the LSPR response is largely dictated by the dimensions and morphology
72 of the plasmonic nanoparticles, but it is also highly sensitive to the interaction between
73 biomolecules and the nanoparticles' surface.^{27, 28} Therefore, LSPR excitation in resonance
74 with light irradiation can be used as a potential tool to regulate protein functionality at the
75 interface with the nanomaterial. For example, enzymes immobilized on the surface of

76 nanoparticles can be readily triggered by light irradiation, to enhance their catalytic
 77 activity.^{9, 10, 17, 29} Harnessing LSPR, biocatalysis can be remotely regulated upon light
 78 irradiation to fine tuning artificial reaction biocascades.^{9, 10, 17, 29-31} Hence, the use of light
 79 is a precise, sustainable, noninvasive and remote method to foster innovative solutions in
 80 the biocatalysis field through merging enzymes and plasmonic nanoparticles (Figure 1).³²
 81 ³³ This perspective provides an overview of this emerging research field, in an attempt to
 82 present and discuss the most exciting studies where light controls the enzyme activity at
 83 the interface with plasmonic nanomaterials, and ultimately offering our own outlook.
 84



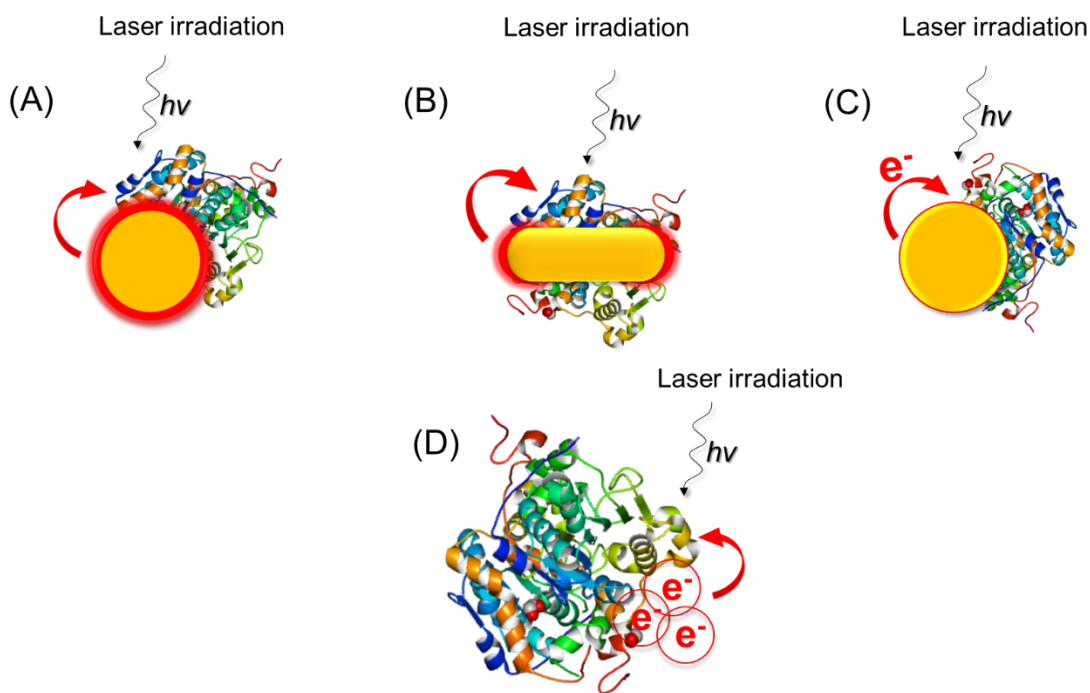
85
 86 **Figure 1.** Schematic representation of some foreseen prospects for light-driven enzymatic
 87 activity regulation onto plasmonic nanoparticles. For clarity purposes, enzymes and
 88 nanoparticles are presented in similar sizes, but this is just a graphic representation, not
 89 necessarily the real scale.

90
 91

92 LIGHT AS THE MAJOR PLAYER IN THE FIELD: MECHANISMS BEHIND LIGHT-
 93 DRIVEN BIOCATALYSIS INTERFACING PLASMONIC NANOPARTICLES

94 Light irradiation can introduce enhanced energetic processes through different
95 mechanisms into plasmonic nanoparticles, depending on the nanoparticle features, such
96 as their size and morphology. For example, anisotropic nanoparticles can concentrate the
97 generated energy at sharp corners or tips, with a significant enhancement of several orders
98 of magnitude.³⁴ Plasmonic effects are thus expected to influence the functional properties
99 of enzymes (activity and stability) in their neighborhood. Unfortunately, we are still
100 lacking fundamental knowledge and understanding to properly explain the mechanisms
101 underlying this phenomenon. It is well-established that chemical reactions triggered by
102 plasmonic effects in nanoparticles are mainly driven by one or more of the following
103 physical effects: (i) temperature increase promoted by incident light, also termed
104 thermoplasmonics (Figure 2A); (ii) enhancement of the electromagnetic near-field at the
105 nanoparticles surface, creating “hotspots” of concentrated energy per unit volume (Figure
106 2B); and (iii) light-induced hot electrons, where light promotes the transfer of an electron
107 from the plasmonic nanoparticle to an adjacent molecule on their surface (Figure 2C).³⁴
108 On the other hand, in photo-biocatalysis, enzyme activity is enhanced by directly shining
109 light onto the enzymes, usually oxidoreductases, which harbor a photoactive cofactor.³⁵
110 ³⁶ The mechanisms driving such photobiocatalytic reactions are mainly related to the
111 photoexcitation of such photoactive cofactors, which shuttle electrons between the
112 substrates and the products (Figure 2D).³⁷⁻⁴⁰ Hence, on the basis of this fundamental
113 knowledge, some attempts have been made to address the study of the mechanisms
114 governing light-driven biocatalysis of enzymes interfacing with plasmonic nanoparticles,
115 which we briefly review in this section.

116



117

118 **Figure 2.** Schematic representation of potential mechanisms of enzymes interfacing with
 119 plasmonic nanomaterials, upon laser irradiation. (A-C) Concepts from nanoplasmonics
 120 through temperature increase (A), “hot-spots” generated from the enhancement of the
 121 optical near-field (B), transfer of hot electrons to the enzyme structure (C). (D) Concepts
 122 from photo-biocatalysis with photoinduction of electron transfer to the enzyme. For
 123 clarity purposes, enzymes and nanoparticles are presented in similar sizes, but this is just
 124 a graphic representation, not necessarily the real scale.

125

126 The photo-induced increase of local temperature on plasmonic nanoparticles is
 127 arguably the most intensively explored effect to explain the mechanisms involved in
 128 enhanced enzymatic activity.^{9, 10, 12, 13, 17, 29, 30, 41} Gold, the most common plasmonic
 129 nanomaterial, features inherent properties such as low specific heat capacity associated
 130 to high thermal conductivity, which render it an ideal candidate for thermally enhanced
 131 processes.⁸ In principle, the activity of enzymes immobilized on the surface of plasmonic
 132 nanoparticles should rise along with the local temperature increase induced by light,¹²
 133 according to Arrhenius equation. However, a local high temperature can also promote a
 134 partial or even complete deactivation of the enzymes, due to conformational changes.
 135 This dichotomy makes it extremely important to precisely tune the heating ability of the
 136 involved plasmonic nanoparticles,⁴² so that the enzyme surroundings can reach an optimal
 137 temperature ensuring maximum activity while avoiding deactivation.

138 On the other hand, hardly any studies are available that explore electron transfer
 139 between excited plasmonic nanoparticles and enzyme structures. One of the few, if not

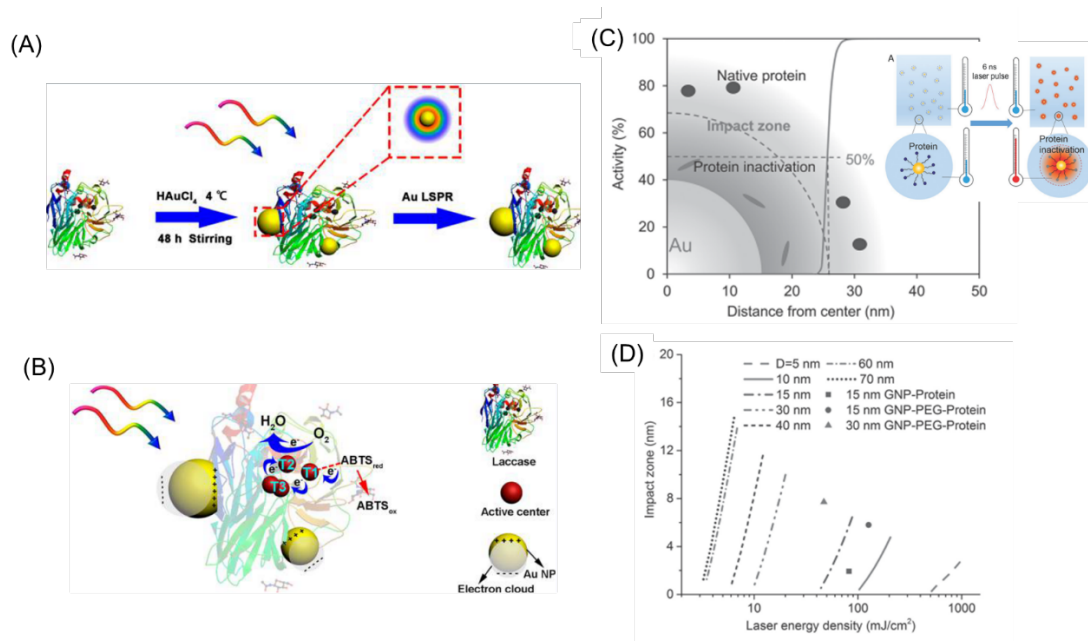
140 the only example that addresses these light-driven electron transfer mechanisms uses a
141 laccase immobilized onto gold nanoparticles (AuNPs) under green light.³⁰ Laccase is a
142 multicopper oxidase that transfers electrons from the substrate to the molecular oxygen
143 towards two copper clusters found at the enzyme's active site. More specifically, laccases
144 work through a copper atom (T1 site) that takes electrons from a reducing substrate, to
145 subsequently transfer those electrons to a trinuclear copper center (T2/T3 sites) that
146 ultimately shuttles the electrons to the molecular oxygen as the final electron acceptor to
147 produce water. It has been observed that green light irradiation increases the local
148 temperature at the surface of AuNP biohybrids, promoting conformational changes in the
149 immobilized laccase that likely alter electron transfer within its active site. Therefore, the
150 local temperature increase at AuNPs surfaces seems to unpair the electron transfer
151 between T1 and T2/T3 sites, which consequently diminishes the laccase activity (Figure
152 3A and B).

153 Mechanistic studies concerning electron transfer upon LSPR excitation are
154 usually carried out using redox enzymes, because their mechanisms intrinsically involve
155 the transfer of electrons. In an illustrative example, Wang *et al* identified hot charge
156 carriers generated on AuNPs under visible light excitation (450, 532, 652, and 808 nm)
157 in a plasmon-accelerated electrochemical reaction (PAER).⁴³ The experiments confirmed
158 the presence of hot charge carriers, responsible for the enhanced electrocatalytic oxidation
159 of glucose, as a model system. Although enzymes were not explored in this study, it
160 highlights opportunities to exploit PAER with enzymes to regulate biocatalysis and
161 elucidate the mechanisms involving electron transfer. For example, in the oxidation of
162 D-glucose catalyzed by glucose oxidase (GOx), the mechanism involves molecular
163 oxygen as an electron acceptor and a flavin cofactor as an electron shuttle.⁴⁴ The same
164 concept could be expanded to other flavin oxidases that oxidize a diversity of substrates,
165 such as alcohols, amino acids, and cholesterol, also using oxygen as the final electron
166 acceptor.

167 In this context, it is clear that different events can concurrently occur upon light-
168 driven biocatalysis through LSPR excitation, but some of them (i.e. temperature increase)
169 are easier to identify than others (electronic effects). An interplay of different mechanisms
170 can also result in one mechanism triggering another. Despite the few mechanistic studies
171 involving plasmonic regulation of enzyme properties, we consider that the interactions
172 between enzymes and nanomaterials are essential to harness the plasmonic effects upon
173 light irradiation, as it happens in electrobiocatalysis, where electrons go back and forth

174 between the enzyme active sites and the electrodes.⁸ In these cases, the distance, the
175 number of attachments, the orientation and the density of the immobilized enzyme have
176 been proven to play a central role in the interplay between biocatalysts and materials, for
177 a variety of applications ranging from biosensing to biotransformations.⁴⁵ As an example,
178 Kang *et al.* observed that the photothermal inactivation of α -chymotrypsin immobilized
179 on AuNPs (irradiated at 532 nm) was strongly dependent on its distance to the AuNP
180 surface.¹³ Because of the rapid local temperature gradients generated in the region termed
181 impact zone, enzyme deactivation occurred immediately under nanosecond laser pulses
182 (Figure 3C). The authors additionally observed that the size of AuNPs (5-70 nm) was also
183 a determining factor behind the temperature gradient responsible for protein inactivation:
184 the larger particles induce larger impact zones at low laser power (Figure 3D). In this
185 scenario, small AuNPs (5 nm) did not cause enzyme inactivation, while larger AuNPs
186 (>50 nm) readily inactivated the enzymes upon laser irradiation. The AuNPs surface-to-
187 volume ratio and the laser energy, related to the efficiency of heat dissipation to the
188 surroundings, were argued to explain enzyme inactivation under irradiation conditions.
189 Since the local temperature in the enzyme surroundings strongly depends on the distance
190 between enzyme and nanoparticle surface, the heating and dissipation capacity of the
191 nanoparticles, together with the intrinsic enzyme thermal stability, should determine the
192 optimal immobilization distance in light-controlled biocatalysis. We suggest that further
193 studies are needed to address the impact of protein orientation and the separation between
194 enzymes and nanomaterials on photothermal effects. We foresee that protein engineering
195 and chemical biology will facilitate these studies, *e.g.* by creating new enzyme variants
196 with engineered surfaces, so as to precisely control their interaction with plasmonic
197 nanomaterials. We therefore stress that, using plasmonic nanoparticles to manipulate
198 enzyme activity is still highly challenging and demands a clear understanding of the
199 mechanisms responsible for plasmon-enhanced biocatalysis. We envision plenty of
200 opportunities to elucidate and dissect the mechanisms underlying LSPR effects on
201 enzyme functionality. For instance, light-driven plasmonic effects may control the
202 intermolecular interactions or bond breaking to timely and remotely activate or deactivate
203 enzymes of interest.

204



205

206 **Figure 3.** Light effect on enzymes interfacing with plasmonic nanomaterials.
 207 Temperature effect (A) and electron transfer effect (B) in laccase-AuNPs bioconjugates,
 208 under green light irradiation. Reproduced with permission from ref. 30. Copyright 2015
 209 American Chemical Society. (C) Impact zone on the nanoparticle surface for α -
 210 chymotrypsin inactivation and (D) effect of AuNP size and laser energy on impact zone.
 211 Adapted with permission from ref. 13. Copyright 2017 John Wiley and Sons. For clarity
 212 purposes, enzymes and nanoparticles are presented in similar sizes, but this is not
 213 necessarily the real scale.

214

215

216 PLASMONIC NANOMATERIALS AS AN OPPORTUNITY TO INTRODUCE
 217 REMOTE CONTROL IN ENZYME ACTIVITY

218

219

220

221

222

223

224

225

226

227

228

The response of plasmonic materials under irradiation conditions, and thus their effect on enzyme functionality, can be regulated by tuning parameters of the incident light such as wavelength, power, and illumination time. Some examples illustrate how light decreases the activity of enzymes immobilized on plasmonic nanoparticles. Guo *et al.* observed³⁰ a decrease in the activity of laccase-AuNP bionanohybrids by 8.1% under visible light ($\lambda > 420$ nm) illumination for 3 minutes, with recovery upon turning the light off. This effect was wavelength-dependent, since green light ($\lambda = 500 - 560$ nm) induced the lowest enzyme activity, likely due to resonance with the LSPR of spherical AuNPs ($\lambda \sim 520$ nm). Interestingly, the intensity of green light - ranging 0 - 60 $\text{mW} \cdot \text{cm}^{-2}$ - followed a negative correlation with the activity of the immobilized laccase. Similar results were obtained for horseradish peroxidase (HRP) immobilized onto spherical AuNPs upon laser

229 irradiation at $\lambda = 532 \text{ nm}$,⁴¹ suggesting that enzyme inactivation is related to released heat
230 under resonance conditions. Another interesting example of enzyme inactivation by heat
231 generation was provided by Thompson *et al.*¹² By using gold nanorods (AuNRs) (λ at
232 $\sim 785 \text{ nm}$) in resonance with incident near-infrared (NIR) laser irradiation ($\lambda = 785 \text{ nm}$),
233 the authors showed selective control of a cascade reaction induced by glucose oxidase
234 (GOx) immobilized on AuNRs and free horseradish peroxidase (HRP), through selective
235 inactivation of GOx (Figure 4A). As the bi-enzyme system is controlled through the
236 activity of one of the enzymes – GOx – this is a one-time (shot) control because once
237 GOx is inactivated upon light irradiation, the bi-enzyme cascade cannot be subsequently
238 recovered.

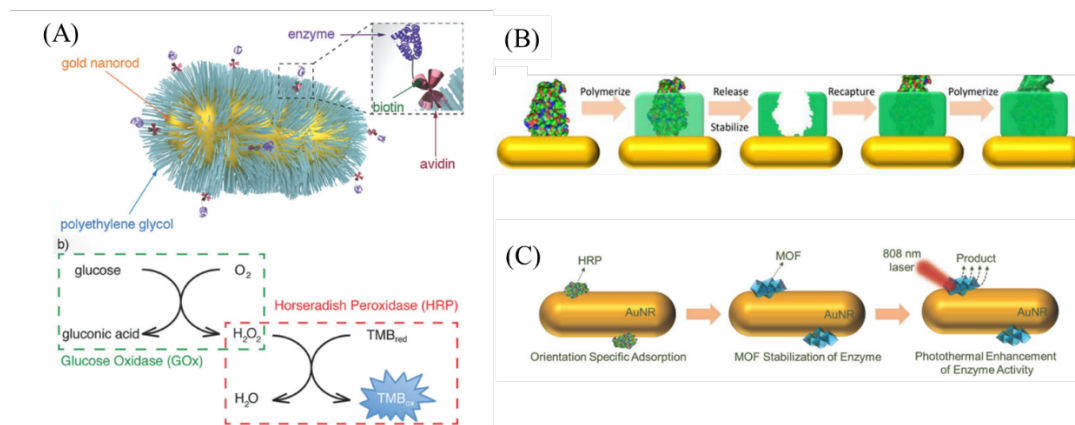
239 Light-driven biocatalysis on plasmonic nanoparticles becomes more attractive
240 when using thermophilic enzymes, since they are highly active and stable at high
241 temperature ($>70 \text{ }^\circ\text{C}$). The local heating triggered by the LSPR effect may be thus
242 transferred to thermophilic enzymes immobilized on plasmonic NPs, thereby enhancing
243 their catalytic activity. This concept has been demonstrated for several classes of
244 temperature-resistant enzymes. Li *et al.* compared the enzymatic performance of three
245 enzymes with different thermal stability, conjugated to AuNRs and irradiated with a NIR
246 laser (808 nm).¹⁷ The results were compared for a mesophilic enzyme pig pancreatic
247 lipase (PPL), a thermophilic protease from *Sulfolobus tokodaii* strain T7 (ST0779), and a
248 hyperthermophilic protease from *Aeropyrum pernix* strain K1 (APE1547), with optimal
249 temperatures of 37, 70 and 90 $^\circ\text{C}$, respectively. Under laser irradiation, the activity of the
250 enzymes immobilized onto AuNRs increased by 75.3%, 118.4% and 218.4%,
251 respectively, compared to their free enzyme counterparts. It was interesting to confirm
252 that more thermophilic enzymes underwent a higher activity enhancement by the
253 photothermal effect as a consequence of LSPR illumination. To further concentrate the
254 heating generated by the plasmonic effect in the enzyme surroundings, Blankschien *et al.*
255 trapped AuNRs conjugated with a thermophilic glucokinase from *Aeropyrum pernix* into
256 alginate macrobeads.²⁹ Under 800 nm laser irradiation, the enzymatic activity was found
257 to increase by 60%, compared to alginate-trapped enzyme, whereas a negligible effect
258 was observed by illuminating the same biohybrid in dispersion (outside the beads). This
259 study shows how a surrounding matrix can preclude fast heat dissipation, thereby
260 preserving the generated heat to enhance enzymatic activity. However, entrapment of the
261 biohybrids in macroporous hydrogels brings along a 10-fold decrease in enzymatic

262 activity, likely due to mass transport restrictions for the substrate to reach the enzyme
263 active sites. To overcome mass transport limitations, other solutions have been proposed
264 to confine the heat generated by photothermal effects at the nanoscale. Tadepalli *et al.*
265 explored two strategies to encapsulate bionanohybrids through either *in situ*
266 polymerization or by means of metal-organic frameworks (MOF) (Figure 4B and C).^{9, 10}
267 The enzymatic activity of HRP immobilized on AuNRs and protected by either a polymer
268 layer or MOF architecture, was stimulated by approximately 110% and 80% under laser
269 irradiation (808 nm, 400 mW·cm⁻²), respectively, compared to the dark reaction. These
270 strategies showed that local heating may be preserved at the enzymes surroundings,
271 thereby stimulating their activity at the NP interface. Hence, nanoencapsulation seems to
272 be a suitable strategy to promote photothermal effects in enzymes of different nature and
273 origin (besides thermophilic) without jeopardizing their stability under the resulting high
274 local temperatures induced by LSPR illumination.

275 From the studies reported during the past decade, we conclude that the activity of
276 enzymes immobilized on plasmonic nanomaterials upon light irradiation depends on
277 various parameters related to enzyme nature and type, nanomaterial properties
278 (morphology, size, LSPR signal) and laser features (power, wavelength, illumination
279 time). It is important to mention that nanoparticle concentration in the dispersion,
280 regarding the density of the immobilized enzyme, is decisive to tune the heat output
281 transfer in these systems, and therefore controlling bionanohybrid activity under
282 irradiation.^{46, 47} Finally, we identify scenarios where the enzyme activity either increases
283 or decreases upon light irradiation. However, the vast majority of studies are focused on
284 quantifying the effect of light once the enzyme is immobilized without accounting for the
285 enzyme activity lost upon conjugation. To evaluate the effect of light on the enhancement
286 of enzyme functionality, we encourage reporting the specific activity of the enzyme before
287 and after immobilization, under irradiation and non-irradiation conditions.

288

289



290

291 **Figure 4.** Examples of enzyme inactivation or activation onto plasmonic nanoparticles
 292 upon laser irradiation. (A) Strategy to selective targeting of GOx or HRP immobilized
 293 onto PEG-AuNRs, upon NIR laser irradiation. Reproduced with permission from ref. 12.
 294 Copyright 2017 John Wiley and Sons. (B,C) Schematic representation of strategies to
 295 confine the heat generated at AuNRs to enhance HRP enzymatic activity by *in situ*
 296 polymerization (B) and by using MOFs (C). Reproduced with permission from ref. 9.
 297 Copyright 2017 American Chemical Society. Reproduced with permission from ref. 10.
 298 Copyright 2018 John Wiley and Sons.

299

300 CELL-FREE PLASMONIC REGULATION OF BIOSYNTHETIC PATHWAYS

301 Metabolic networks within living cells have evolved for billions of years to
 302 maximize fitness and ensure optimal cell survival and reproducibility within specific
 303 environments. Hence, the cell metabolism relies on a multitude of chemical reactions
 304 catalyzed by highly chemo-, regio- and stereo-selective enzymes. These biological
 305 catalysts are perfectly orchestrated to work simultaneously within the cell milieu, under
 306 similar reaction conditions. Although some enzymatic reactions are compartmentalized,
 307 the vast majority of them occur inside the cytosol. For this reason, the regulation of
 308 enzyme activity is extremely important to efficiently route biosynthetic pathways. Within
 309 the cells, such metabolic pathways are mainly regulated at the genomic level by
 310 enhancing or suppressing the expression of the genes encoding the enzymes involved in
 311 the target pathway, and at the protein level through allosterism and post-translational
 312 modifications. In the former case, gene regulation pathways respond to both metabolite
 313 accumulation and environmental changes, by either increasing or decreasing the
 314 intracellular enzyme concentrations. In contrast, the latter case promotes conformational
 315 changes in enzymes, thereby modulating their activity through partial proteolysis or
 316 chemical modification.

317 Human beings exploit metabolic networks for their survival but also for
318 technological purposes. Since the beginning of biotechnology in the 7th century B.C,
319 ancient societies made use of enzymes for food processing (wine, beer, bread...).
320 Nowadays, enzymes are routed to catalyze non-natural biosynthetic pathways demanded
321 by modern biotechnological applications in chemical manufacturing, biosensing and
322 biomedicine.³¹ These applications often require the enzyme to work far from
323 physiological conditions. Therefore, driving the enzymes out of their natural environment
324 makes us face technological challenges related to their functionality toward non-natural
325 substrates and their responsiveness to external stimuli. Protein engineering has
326 successfully adapted the functionality of a myriad natural enzymes to artificial synthetic
327 reactions,^{48, 49} yet the regulation of such artificial cascades must be engineered.

328 In recent years, many approaches have been developed to artificially regulate the
329 catalytic activity of isolated enzymes. With the revolution of cell-free synthetic biology,
330 more complex biosynthetic pathways are assembled to work under non-physiological
331 conditions. Defining synthetic biology as the discipline that integrates the engineering
332 principles behind biological processes, materials chemistry and engineering can
333 contribute to expand this concept beyond DNA modifications. Through the incorporation
334 of biological machinery (i.e multi-enzyme systems) at the interfaces between stimuli-
335 responsive materials (i.e. plasmonic nanoparticles), biological processes can gain novel
336 functionalities. This revolution is still seeking radical solutions to control the activity of
337 enzymes, so that their performance can be tuned in the context of cell-free biosynthetic
338 pathway.

339 Enzyme cascades based on NADH-dependent alcohol dehydrogenases have been
340 controlled using light as an external stimulus, through different approaches. The
341 combination of enzymes and materials with photochromic functionality is usually
342 exploited for this aim. The most common approach comprises mixing soluble enzymes
343 and photoactive materials in one-pot, which couples a photocatalytic step with a
344 biocatalytic one.⁵⁰ Whereas the enzymes consume the redox cofactor to perform the
345 reaction of interest, the light-irradiated material replenishes the pool of the cofactor
346 demanded by the enzyme. A similar strategy has been recently adopted for the
347 photocatalytic *in situ* production of H₂O₂ employed by peroxygenases.⁵¹
348 Events where light remotely controls the activity of more than one enzyme attached to
349 the surface of light-responsive or plasmonic nanomaterials are however scarce. The
350 assembly of two NADH-dehydrogenases in DNA origami scaffolds is one of the few

351 examples where light can directly control the activity of two enzymes.⁵² This system
352 relies on DNA strands functionalized with azo groups that control the relative position of
353 each enzyme, bridging them together and activating the cascade under visible light, but
354 separating and inhibiting them under UV irradiation. More complex systems where more
355 than two enzymes are simultaneously controlled by light have not been reported so far.
356 We can definitely see here an opportunity for the use of plasmonic effects (photothermal
357 or electron transfer) interfacing with multi-enzyme systems. In fact, one can envision in
358 the long term a cohort of bionanoplasmonic hybrids whose catalytic activities are
359 activated under different irradiation conditions (lasers with different wavelengths). In this
360 scenario, different enzymes could be remotely activated through either simultaneous (two
361 lasers at a time) or sequential (one laser after another) light stimuli to orchestrate a given
362 enzymatic cascade reaction on demand. An enzyme cascade composed of thermophilic
363 enzymes and thermolabile cofactors transforming temperature-sensitive substrates would
364 be an excellent proof of this concept. While the enzymes interfacing the nanoplasmonic
365 materials operate at their optimal temperature under irradiation conditions due to local
366 heating, the bulk remains at room temperature avoiding the deterioration of both
367 substrates and cofactors.

368

369 CHALLENGES AND PERSPECTIVES

370 Light-regulation at the interface between enzymes and nanomaterials is an
371 outstanding concept to control enzymatic activity through remote stimuli. Regulation of
372 enzymes is particularly relevant for sequential enzymatic reactions, since pairing the
373 activities of all involved enzymes is paramount to optimally drive the chemical flux
374 toward the target product. When enzyme cascades simultaneously work in one pot, their
375 activities must be orchestrated to avoid accumulation of inhibiting intermediates and toxic
376 by-products. Within living cells, such biosynthetic orchestration is met through
377 sophisticated genomic and proteomic regulation pathways that control the expression
378 level and post-translational modifications of the enzyme working in the cell milieu. When
379 using enzymes for *ex vivo* applications, enzyme regulation becomes an arduous task that
380 requires artificial actuators to remotely exert control over the enzymatic activities. These
381 challenges open up the opportunity of using plasmonic nanomaterials, which harness the
382 incident light to tune enzyme functionalities at the protein-nanomaterial interface.

383 Heat generated at the surface of plasmonic nanoparticles is attractive to
384 manipulate the catalytic properties of enzymes, but effective strategies to protect enzyme

385 activity are still challenging. Even though progress has been made for thermophilic
386 enzymes, little has been developed for mesophilic enzymes in terms of enzymatic activity
387 enhancement. Strategies for remote-control of mesophilic enzymes are highly relevant
388 because these enzymes represent the majority of applications in biocatalysis and
389 biotransformations, overall when compared to thermophilic ones. Hence, plasmonic
390 nanoparticles can introduce numerous advantages for remote-control of biocatalysis using
391 mesophilic enzymes if we can find the optimal trade-off between activity and stability
392 under light-induced local heating. Besides increasing the local temperature, LPSR effects
393 can also generate hot-electrons at the surface of plasmonic NPS, which may induce
394 electronic effects on enzymatic mechanisms. Unfortunately such light-driven plasmonic
395 effects have been rarely exploited to control enzyme activity. Here, we identify a
396 scientific niche to remotely regulate the activity of redox enzymes with catalytic
397 mechanisms relying on electron transfer. New discoveries in this direction are yet to
398 come.

399 Controlling more than one enzyme simultaneously is an unmet challenge using
400 plasmonic nanomaterials. Even using phenomena other than plasmonics, few examples
401 have been reported where light itself can control a multi-enzyme process. Therefore, the
402 creativity window stays open toward the design of novel biohybrids, where the interface
403 between materials and enzymes comes to play in the presence of an external stimulus
404 such as light. Hence, enzymatic material interfaces are needed to allow efficient light
405 control over enzyme functionality. Protein engineering is a promising route toward
406 gaining control over orientation, number of attachments and distance between the enzyme
407 and the plasmonic nanomaterial.

408 The revolution of synthetic biology has arrived to cell-free systems, where many
409 isolated enzymes work simultaneously in the same pot and under the same conditions. It
410 is thus the right time to introduce regulators that exert control over cell-free enzymatic
411 systems to up- or down-regulate enzymes without altering the reaction medium. To that
412 aim, such remote regulators must respond to external stimuli as plasmonic nanomaterials
413 do to light. Hence, we envision enzymes immobilized on plasmonic nanomaterials as new
414 tools to endow cell-free synthetic biology with spatio-temporal control. Additionally, the
415 immobilization of enzymes on light-responsive materials, such as plasmonic
416 nanoparticles, is likely to pave the way to more futuristic applications like the chemical
417 communication between compartmentalized cell-free systems, or even living cells. More

418 rational interfaces between enzymes and nanomaterials must therefore be created and new
419 theoretical knowledge must be acquired to better understand the mechanisms governing
420 the enzyme functionality at the interface with plasmonic nanomaterials.

421

422 ACKNOWLEDGEMENTS

423 HRB thanks the São Paulo Research Foundation FAPESP for fellowship (FAPESP
424 2019/09668-0). Financial support was provided by the European Research Council (ERC-
425 AdG-2017 #787510 and ERC-CoG-2018 #818089). Funding from IKERBASQUE to
426 L.L.M and F.L.G is also acknowledged. This work was performed under the Maria de
427 Maeztu Units of Excellence Program from the Spanish State Research Agency – Grant
428 No. MDM-2017-0720.

429

430 REFERENCES

- 431 [1] Ideker, T., Thorsson, V., Ranish, J. A., Christmas, R., Buhler, J., Eng, J. K., Bumgarner, R.,
432 Goodlett, D. R., Aebersold, R., and Hood, L. (2001) Integrated genomic and proteomic
433 analyses of a systematically perturbed metabolic network, *Science* 292, 929-934.
- 434 [2] Kneuttinger, A. C., Winter, M., Simeth, N. A., Heyn, K., Merkl, R., Konig, B., and Sterner, R.
435 (2018) Artificial light regulation of an allosteric bienzyme complex by a photosensitive
436 ligand, *ChemBioChem* 19, 1750-1757.
- 437 [3] Bautista-Barrufet, A., Lopez-Gallego, F., Rojas-Cervellera, V., Rovira, C., Pericas, M. A., Guisan,
438 J. M., and Gorostiza, P. (2014) Optical control of enzyme enantioselectivity in solid
439 phase, *ACS Catal.* 4, 1004-1009.
- 440 [4] Shimoboji, T., Larenas, E., Fowler, T., Kulkarni, S., Hoffman, A. S., and Stayton, P. S. (2002)
441 Photoresponsive polymer-enzyme switches, *Procs. Nat. Acad. Sci. U. S. A.* 99, 16592-
442 16596.
- 443 [5] Algar, W. R., Prasuhn, D. E., Stewart, M. H., Jennings, T. L., Blanco-Canosa, J. B., Dawson, P.
444 E., and Medintz, I. L. (2011) The controlled display of biomolecules on nanoparticles: A
445 challenge suited to bioorthogonal chemistry, *Bioconjugate Chem.* 22, 825-858.
- 446 [6] Sapsford, K. E., Algar, W. R., Berti, L., Gemmill, K. B., Casey, B. J., Oh, E., Stewart, M. H., and
447 Medintz, I. L. (2013) Functionalizing nanoparticles with biological molecules: Developing
448 chemistries that facilitate nanotechnology, *Chem. Rev.* 113, 1904-2074.
- 449 [7] Ansari, S. A., and Husain, Q. (2012) Potential applications of enzymes immobilized on/in nano
450 materials: A review, *Biotechnol. Adv.* 30, 512-523.
- 451 [8] Kamada, K. (2019) Photo-manipulation of activity of enzymes bound to inorganic
452 nanomaterials, *J. Solid State Chem.* 280, 210-214.
- 453 [9] Tadepalli, S., Yim, J., Madireddi, K., Luang, J. Y., Naik, R. R., and Singamaneni, S. (2017) Gold
454 nanorod-mediated photothermal enhancement of the niocatalytic sctivity of a polymer-
455 encapsulated enzyme, *Chem. Mater.* 29, 6308-6314.
- 456 [10] Tadepalli, S., Yim, J., Cao, S. S., Wang, Z. Y., Naik, R. R., and Singamaneni, S. (2018) Metal-
457 organic framework encapsulation for the preservation and photothermal enhancement
458 of enzyme activity, *Small* 14, 1702382.
- 459 [11] Maji, S. K. (2019) Plasmon-enhanced electrochemical biosensing of hydrogen peroxide from
460 cancer cells by gold nanorods, *ACS Appl. Nano Mater.* 2, 7162-7169.
- 461 [12] Thompson, S. A., Paterson, S., Azab, M. M. M., Wark, A. W., and de la Rica, R. (2017) Light-
462 Triggered inactivation of enzymes with photothermal nanoheaters, *Small* 13, 1603195.

- 463 [13] Kang, P. Y., Chen, Z., Nielsen, S. O., Hoyt, K., D'Arcy, S., Gassensmith, J. J., and Qin, Z. P.
464 (2017) Molecular hyperthermia: Spatiotemporal protein unfolding and inactivation by
465 nanosecond plasmonic heating, *Small* **13**, 1700841.
- 466 [14] Cao, G. X., Sun, D. X., Gu, T. T., Dong, Y. M., and Wang, G. L. (2019) Photoswitching enzymatic
467 activity of horseradish peroxidase by graphene oxide for colorimetric immunoassay,
468 *Biosens. Bioelectron.* **145**, 111707.
- 469 [15] Lin, Y., Zhao, M. M., Guo, Y. J., Ma, X. M., Luo, F., Guo, L. H., Qiu, B., Chen, G. N., and Lin, Z.
470 Y. (2016) Multicolor colorimetric biosensor for the determination of glucose based on
471 the etching of gold nanorods, *Sci. Rep.* **6**, 37879.
- 472 [16] Yan, J., Wang, L. D., Tang, L. H., Lin, L., Liu, Y., and Li, J. H. (2015) Enzyme-guided plasmonic
473 biosensor based on dual-functional nanohybrid for sensitive detection of thrombin,
474 *Biosens. Bioelectron.* **70**, 404-410.
- 475 [17] Li, W., Liu, D. N., Geng, X., Li, Z. Q., and Gao, R. J. (2019) Real-time regulation of catalysis by
476 remote-controlled enzyme-conjugated gold nanorod composites for aldol reaction-
477 based applications, *Catal. Sci. Technol.* **9**, 2221-2230.
- 478 [18] Ding, Y. C., Bertram, J. R., Eckert, C., Bommareddy, R. R., Patel, R., Conradie, A., Bryan, S.,
479 and Nagpal, P. (2019) Nanorg microbial factories: Light-driven renewable biochemical
480 synthesis using quantum dot-bacteria nanobiohybrids, *J. Am. Chem. Soc.* **141**, 10272-
481 10282.
- 482 [19] Schroeder, A., Goldberg, M. S., Kastrop, C., Wang, Y. X., Jiang, S., Joseph, B. J., Levins, C. G.,
483 Kannan, S. T., Langer, R., and Anderson, D. G. (2012) Remotely activated protein-
484 producing nanoparticles, *Nano Lett.* **12**, 2685-2689.
- 485 [20] Lee, S. E., Liu, G. L., Kim, F., and Lee, L. P. (2009) Remote optical switch for localized and
486 selective control of gene interference, *Nano Lett.* **9**, 562-570.
- 487 [21] Knecht, L. D., Ali, N., Wei, Y. N., Hilt, J. Z., and Daunert, S. (2012) Nanoparticle-mediated
488 remote control of enzymatic activity, *ACS Nano* **6**, 9079-9086.
- 489 [22] Mosquera, J., Zhao, Y., Jang, H. J., Xie, N. L., Xu, C. L., Kotov, N. A., and Liz-Marzán, L. M.
490 (2020) Plasmonic nanoparticles with supramolecular recognition, *Adv. Funct. Mater.* **30**,
491 1902082.
- 492 [23] Litti, L., Reguera, J., de Abajo, F. J. G., Meneghetti, M., and Liz-Marzán, L. M. (2020)
493 Manipulating chemistry through nanoparticle morphology, *Nanoscale Horiz.* **5**, 102-108.
- 494 [24] Bodelon, G., Costas, C., Perez-Juste, J., Pastoriza-Santos, I., and Liz-Marzán, L. M. (2017)
495 Gold nanoparticles for regulation of cell function and behavior, *Nano Today* **13**, 40-60.
- 496 [25] Liz-Marzán, L. M., Murphy, C. J., and Wang, J. F. (2014) Nanoplasmonics, *Chem. Soc. Rev.*
497 **43**, 3820-3822.
- 498 [26] Liz-Marzán, L. M. (2004) Nanometals formation and color, *Mater. Today* **7**, 26-31.
- 499 [27] Atale, S. S., Dyawanapelly, S., Jagtap, D. D., Jain, R., and Dandekar, P. (2019) Understanding
500 the nano-bio interactions using real-time surface plasmon resonance tool, *Int. J. Biol.*
501 *Macromolecules* **123**, 97-107.
- 502 [28] Wang, P. Y., Wang, X., Wang, L. M., Hou, X. Y., Liu, W., and Chen, C. Y. (2015) Interaction of
503 gold nanoparticles with proteins and cells, *Sci. Technol. Adv. Mater.* **16**, 034610.
- 504 [29] Blankschien, M. D., Pretzer, L. A., Huschka, R., Halas, N. J., Gonzalez, R., and Wong, M. S.
505 (2013) Light-triggered biocatalysis using thermophilic enzyme-gold nanoparticle
506 complexes, *ACS Nano* **7**, 654-663.
- 507 [30] Guo, S. J., Li, H., Liu, J., Yang, Y. M., Kong, W. Q., Qiao, S., Huang, H., Liu, Y., and Kang, Z. H.
508 (2015) Visible-light-induced effects of Au nanoparticle on laccase catalytic activity, *ACS*
509 *Appl. Mater. Interfaces* **7**, 20937-20944.
- 510 [31] Schrittwieser, J. H., Velikogne, S., Hall, M., and Kroutil, W. (2018) Artificial biocatalytic linear
511 cascades for preparation of organic molecules, *Chem. Rev.* **118**, 270-348.
- 512 [32] Wang, F. M., Ju, E. G., Guan, Y. J., Ren, J. S., and Qu, X. G. (2017) Light-mediated reversible
513 modulation of ROS level in living cells by using an activity-controllable nanozyme, *Small*
514 **13**, 1603051.

- 515 [33] Li, J. Y., Liu, J., and Chen, C. Y. (2017) Remote control and modulation of cellular events by
516 plasmonic gold nanoparticles: Implications and opportunities for biomedical
517 applications, *ACS Nano* **11**, 2403-2409.
- 518 [34] Baffou, G., and Quidant, R. (2014) Nanoplasmonics for chemistry, *Chem. Soc. Rev.* **43**, 3898-
519 3907.
- 520 [35] Zhang, W. Y., Ma, M., Huijbers, M. M. E., Filonenko, G. A., Pidko, E. A., van Schie, M., de
521 Boer, S., Burek, B. O., Bloh, J. Z., van Berkel, W. J. H., Smith, W. A., and Hollmann, F.
522 (2019) Hydrocarbon synthesis via photoenzymatic decarboxylation of carboxylic acids,
523 *J. Am. Chem. Soc.* **141**, 3116-3120.
- 524 [36] Wuyuan Zhang, J.-H. L., Sabry H. H. Younes, Fabio Tonin, Peter-Leon Hagedoorn, Harald
525 Pichler, Yoonjin Baeg, Jin-Byung Park, Robert Kourist, Frank Hollmann. (2020)
526 Photobiocatalytic synthesis of chiral secondary fatty alcohols from renewable
527 unsaturated fatty acids, *Nat. Comm.* **11**, 2258.
- 528 [37] Schmermund, L., Jurkas, V., Ozgen, F. F., Barone, G. D., Buchsenschutz, H. C., Winkler, C. K.,
529 Schmidt, S., Kourist, R., and Kroutil, W. (2019) Photo-biocatalysis: Biotransformations in
530 the presence of light, *ACS Catal.* **9**, 4115-4144.
- 531 [38] Lee, S. H., Choi, D. S., Kuk, S. K., and Park, C. B. (2018) Photobiocatalysis: Activating redox
532 enzymes by direct or indirect transfer of photoinduced electrons, *Angew. Chem. Int. Ed.*
533 **57**, 7958-7985.
- 534 [39] Macia-Agullo, J. A., Corma, A., and Garcia, H. (2015) Photobiocatalysis: The power of
535 combining photocatalysis and enzymes, *Chem. Eur. J.* **21**, 10940-10959.
- 536 [40] Kim, J., and Park, C. B. (2019) Shedding light on biocatalysis: photoelectrochemical platforms
537 for solar-driven biotransformation, *Curr. Op. Chem. Biol.* **49**, 122-129.
- 538 [41] Bretschneider, J. C., Reismann, M., von Plessen, G., and Simon, U. (2009) Photothermal
539 control of the activity of HRP-functionalized gold nanoparticles, *Small* **5**, 2549-2553.
- 540 [42] Baffou, G., and Quidant, R. (2013) Thermo-plasmonics: using metallic nanostructures as
541 nano-sources of heat, *Laser Photon. Rev.* **7**, 171-187.
- 542 [43] Wang, C., Nie, X. G., Shi, Y., Zhou, Y., Xu, J. J., Xia, X. H., and Chen, H. Y. (2017) Direct
543 plasmon-accelerated electrochemical reaction on gold nanoparticles, *ACS Nano* **11**,
544 5897-5905.
- 545 [44] Golikova, E. P., Lakina, N. V., Grebennikova, O. V., Matveeva, V. G., and Sulman, E. M. (2017)
546 A study of biocatalysts based on glucose oxidase, *Faraday Discuss.* **202**, 303-314.
- 547 [45] Wasserberg, D., Cabanas-Danes, J., Prangmsma, J., O'Mahony, S., Cazade, P. A., Tromp, E.,
548 Blum, C., Thompson, D., Huskens, J., Subramaniam, V., and Jonkheijm, P. (2017)
549 Controlling protein surface orientation by strategic placement of oligo-histidine tags,
550 *ACS Nano* **11**, 9068-9083.
- 551 [46] Richardson, H. H., Carlson, M. T., Tandler, P. J., Hernandez, P., and Govorov, A. O. (2009)
552 Experimental and theoretical studies of light-to-heat conversion and collective heating
553 effects in metal nanoparticle solutions, *Nano Lett.* **9**, 1139-1146.
- 554 [47] Kuttner, C., Holler, R. P. M., Quintanilla, M., Schnepf, M. J., Dulle, M., Fery, A., and Liz-
555 Marzán, L. M. (2019) SERS and plasmonic heating efficiency from anisotropic
556 core/satellite superstructures, *Nanoscale* **11**, 17655-17663.
- 557 [48] Chen, Z., and Zeng, A. P. (2016) Protein engineering approaches to chemical biotechnology,
558 *Curr. Op. Biotechnol.* **42**, 198-205.
- 559 [49] Li, Y., and Cirino, P. C. (2014) Recent advances in engineering proteins for biocatalysis,
560 *Biotechnol. Bioeng.* **111**, 1273-1287.
- 561 [50] Son, G., Kim, J., and Park, C. B. (2020) Interference of solvatochromic twist in amyloid
562 nanostructure for light-driven biocatalysis, *ACS Appl. Ener. Mater.* **3**, 1215-1221.
- 563 [51] Zhang, W. Y., Fernandez-Fueyo, E., Ni, Y., van Schie, M., Gacs, J., Renirie, R., Wever, R., Mutti,
564 F. G., Rother, D., Alcalde, M., and Hollmann, F. (2018) Selective aerobic oxidation
565 reactions using a combination of photocatalytic water oxidation and enzymatic
566 oxyfunctionalizations, *Nat. Catal.* **1**, 55-62.

567 [52] Chen, Y. H., Ke, G. L., Ma, Y. L., Zhu, Z., Liu, M. H., Liu, Y., Yan, H., and Yang, C. J. (2018) A
568 synthetic light-driven substrate channeling system for precise regulation of enzyme
569 cascade activity based on DNA origami, *J. Am. Chem. Soc.* *140*, 8990-8996.
570