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(Article begins on next page)

1	Organics	Removal	from	Shale	Gas	Wastewater

# <sup>2</sup> by Pre-oxidation Combined with Biologically <sup>3</sup> Active Filtration

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20 **ABSTRACT:** Biological treatment technology is increasingly explored in shale gas 21 wastewater (SGW) treatment owing to its cost effectiveness and requires efforts to 22 improve its efficacy. In this work, ozone and ferrate(VI) oxidation pre-treatment were 23 evaluated to enhance the performance of the subsequent biologically active filtration 24 (BAF) in the removal of organic contaminants. The oxidation improve the SGW 25 biodegradability and organic composition in the presence of high salinity (~20 g/L). 26 Due to the degradation activity of microorganisms, the organics removal efficiency in 27 the BAF system was observed to gradually improve and then reaching stability in long-term continuous-mode operation. The removal rate of dissolved organic carbon 28 29 (DOC) of the ozone-BAF (O<sub>3</sub>-BAF) and the ferrate(VI)-BAF (Fe(VI)-BAF) systems 30 was 83.2% and 82.8%, respectively, higher than that of BAF alone (80.9%). This 31 increase was attributed to higher activity and content of microorganisms in O<sub>3</sub>-BAF 32 and Fe(VI)-BAF systems. The presence of uncultured bacteria of genus 33 Rehaibacterium with high abundance of 7.2-21.0% was significantly correlated with 34 DOC removal. Also, uncultured bacteria of genus *Methyloversatilis* (2.24-22.31%) 35 were significantly correlated with fluorescent organics removal. Results suggest that these two bacterial species have strong ability to degrade organics. More research is 36 37 needed to understand whether the species were new and their specific function. This 38 study provides valuable suggestions for extracting safe water from SGW with an 39 efficient treatment train.



KEYWORDS: Shale gas wastewater; Biologically active filtration; Ozone; Ferrate

41 (VI); Microbial community

#### 43 **1. INTRODUCTION**

With the fast development of the shale gas industry, associated shale gas 44 45 wastewater (SGW) streams caused by hydraulic fracturing increasingly threaten water ecosystem and human health. In 2016, 6 million tons and 131 million tons of SGW 46 were produced in the Sichuan Basin of China and in the United States, respectively.<sup>1,2</sup> 47 SGW contains high concentrations of salt, heavy metals, microorganisms, and 48 refractory organics released from the shale formation or as chemical residue of 49 compounds added to enhance hydraulic fracturing.<sup>3, 4</sup> Effectively managing SGW 50 produced from shale gas extraction has become an urgent environmental and 51 52 engineering issue.

53 Membrane technologies are usually considered as the most effective tertiary treatment for SGW, but they require significant energy and face serious fouling 54 challenges.<sup>4, 5</sup> A limited number of studies has investigated hybrid systems that apply 55 biological treatment to reduce membrane fouling, and found that biological treatment 56 processes can efficiently control fouling in ultrafiltration and nanofiltration.<sup>6</sup> 57 Furthermore, cost-effective biological treatment technology is regarded as a 58 high-potential treatment technology for SGW, because the large amount of organics 59 present in SGW is mostly biodegradable.<sup>7,8</sup> In the past few years, activated sludge,<sup>9</sup> 60 sequencing batch reactor,<sup>10, 11</sup> membrane bioreactor,<sup>12-14</sup> microbial mats,<sup>15, 16</sup> moving 61 bed biofilm reactor,<sup>17</sup> biologically active filtration (BAF)<sup>6, 18-21</sup> and bioelectrochemical 62 system<sup>22-26</sup> have been evaluated for SGW treatment. 63

64	BAF exploits the biofilm attached to filter media to degrade and adsorb organics
65	from the wastewater. <sup>3</sup> Limited research has shown that BAF can remove organics
66	(72-90% COD and 72-92% DOC) from six different SGW generated in basins of USA
67	with varying salinity (10.5-31.2 g/L TDS) and organics content (85-6360 mg/L COD
68	and 36-2170 mg/L DOC). <sup>6, 19</sup> The BAF efficiency under different operating conditions
69	(aeration rate, temperature, empty bed contact time, and type of activated carbon) was
70	systematically studied. <sup>19-21</sup> However, analysis of the microbial community
71	composition and function in BAF is still scarce. Concurrently, the feasibility of BAF
72	in treating SGW needs further evaluation and the mechanism of pollutant migration
73	and transformation in BAF needs more detailed study.

74 At present, the combination of ozonation and BAF (O<sub>3</sub>-BAF) has been widely used in water treatment, because ozone can degrade refractory pollutants and improve 75 the performance of the subsequent biological process.<sup>27, 28</sup> Also, biological processes 76 can effectively remove ozonation by-products. This system has never been tested for 77 78 the treatment of shale gas wastewater and its feasibility is still unclear for this application. Ferrate(VI) or Fe(VI) has been also successfully used in water and 79 wastewater treatment as a new type of green oxidant.<sup>29</sup> The redox potential of Fe(VI) 80 is +0.7-+2.2 V.<sup>29</sup> The redox potential of ozone is slightly higher than that of Fe(VI) in 81 solutions of basic pH.<sup>29</sup> SGW typically contains high concentrations of Cl<sup>-</sup> and Br<sup>-</sup>, 82 which seriously weaken the oxidation ability and the safety of ozonation.<sup>30</sup> On the 83 contrary, Fe(VI) has no known reactivity with halogens,<sup>29</sup> indicating that Fe(VI) might 84

85	have a certain advantage in oxidizing shale gas wastewater. Similar to ozone, Fe(VI)
86	can degrade refractory pollutants and improve their biodegradability. Nevertheless,
87	there are only a few studies on the combination of Fe(VI) and biological
88	processes. <sup>31-33</sup> Ma et al. <sup>32</sup> found that Fe(VI) (1 mg/L) pretreatment significantly
89	increased the removal rate of $\text{COD}_{\text{Mn}}$ , $\text{UV}_{254}$ , $\text{NH}_4$ by BAF in treating river water.
90	Besides, a simple comparison of Fe(VI)-BAF and $O_3\mbox{-}BAF$ showed that the $\mbox{COD}_{Mn}$
91	removal rate of Fe(VI)-BAF was slightly lower than that of $O_3$ -BAF, and the $\mathrm{NH_4^+}$
92	removal rate of Fe(VI)-BAF was much higher than that of O3-BAF. In general,
93	Fe(VI)-BAF has shown interesting potential, but a more systematic and
94	comprehensive assessment of Fe(VI)-BAF is needed, especially in the treatment of
95	SGW.

Therefore, the objectives of this study are to (i) evaluate the effect of Fe(VI) and O<sub>3</sub> pre-oxidation on organics removal; (ii) assess the effect of Fe(VI) and O<sub>3</sub> pre-oxidation on the performance of BAF systems; (iii) analyze the composition and evolution of microbial community in such hybrid systems, and explore the dominant and functional microorganisms; (iv) comprehensively analyze the feasibility of Fe(VI)-BAF in treating SGW compared with that of O<sub>3</sub>-BAF.

102

## 103 2. EXPERIMENTAL SECTION

104 2.1. Water Samples and Water Quality Analysis. SGW samples were collected
105 from the Changning shale gas play (Sichuan Basin, China). Due to the high turbidity (>

106 1000 NTU) of SGW, SGW was pre-treated with coagulation-sedimentation before the subsequent processing. Therefore, the raw water in the article referred to the SGW 107 108 after coagulation. Aluminum sulfate was chosen as flocculant and the dose was 600 mg/L according to our previous work.<sup>34</sup> The coagulation step was divided into three 109 110 stages: rapid mixing at 200 rpm for 1 minute, then slow mixing at 40 rpm for 20 min, and settling for 30 min.<sup>34</sup> The water quality parameters of SGW were summarized in 111 112 Table S2 of the Supporting Information (SI). The methods for the quantification of 113 dissolved organic carbon (DOC), turbidity, pH, chemical oxygen demands (COD), biochemical oxygen demand (BOD<sub>5</sub>), UV absorbance at 254 nm (UV<sub>254</sub>), total 114 115 dissolved solid (TDS), and fluorescence excitation-emission matrix (EEM) can be found in our previous articles<sup>30, 35</sup> and in Text S1 of the SI. 116

117 **2.2. Experimental Setups and Procedures of Pre-oxidation.** Ferrate (VI) 118 treatment was one of pre-oxidation method. The dosage of Fe(VI) was 40 mg/L. The 119 recrystallization method was used in this study to increase the purity of potassium 120 ferrate ( $K_2$ FeO<sub>4</sub>) to >90%.<sup>36-38</sup> The Fe(VI) pre-oxidation experiment consisted of three 121 stages: rapid mixing at 200 rpm for 2 min, then slow mixing at 40 rpm for 20 min, and 122 settling for 30 min. The supernatant was used as feed water for the subsequent BAF 123 process.

Ozonation was another pre-oxidation method. The dosage of ozone was 80 mg/L, according to our previous work.<sup>30</sup> Treatment of raw water by pre-ozonation was carried out in batch experiment. In each batch experiment, 1 L raw water was added

into a reactor and oxidized by ozone produced form ozone generator (Beijing Tonglin
Co., Ltd., China) at desired flow rate. Before the subsequent BAF treatment, the
residual ozone in water was quenched by water bath heating with 30 min at 50 °C.

130 2.3. Experimental Setup and Protocol of BAF Tests. In the BAF column 131 acclimation process, microorganisms in shale gas wastewater were gradually enriched 132 in activated carbon carrier through sequential batch influent and gradient dilution of 133 raw water, thus forming biofilm on activated carbon. Operation parameters and steps 134 of BAF column acclimation can be found in Table S1 (SI). A carbon source (sodium 135 acetate anhydrous) was added to adjust the C:N ratio to 3.5:1 of the raw water, which 136 was beneficial to the growth of microorganisms. BAF systems were operated in 137 batch-mode at influent flow rate of 0.14 L/h and aerated at a rate of 50 mL/min. The 138 volume of circulation feed tank was 0.55 L, and 0.5 L raw water was changed every 139 two days. The inner diameter and height of the BAF reactor were 1.4 cm and 80 cm, 140 respectively. The filling height of activated carbon (CPG LH 12×40, Calgon Carbon 141 Co., Ltd., USA) was 45 cm, and the filling mass was about 30 g. The corresponding 142 data and analysis during the acclimation stage are summarized in Figure S2 and 143 Figure S3 (SI).

In the continuous-mode BAF systems, the raw water pre-oxidized by ozone or Fe(VI) was used as BAF influent to investigate the treatment effect of combined processes of ozone-BAF ( $O_3$ -BAF) and Fe(VI)-BAF. The raw water was also used as BAF influent for comparison. Three BAF systems were thus operated in

continuous-mode at influent flow rate of 0.014 L/h and aerated at a rate of 10 mL/min.
The backwashing frequency, backwashing flow rate, and backwashing time were 20
days, 0.14 L/h, and 10 min, respectively.

151 2.4. Analysis of Biofilm on GAC. At the end of BAF operation, a certain 152 amount of granular activated carbon (GAC) samples were collected from each BAF 153 column to evaluate the activity, relative concentration and growth of the biofilm on 154 GAC through measurement of the oxygen uptake rate (OUR), extracellular polymeric 155 substances (EPS), GAC static adsorption, and using scanning electron microscopy 156 (SEM) combined with energy dispersive X-ray spectroscopy (EDS) analysis. GAC 157 samples were collected from ~15 cm bed-depth (from the top).

A mass equivalent to 0.3 g GAC was placed into a 150 mL conical flask on a magnetic stirrer. The conical flask was then filled with raw water (about 8 mg/L dissolved oxygen). A rubber plug with a dissolved oxygen probe was installed on the conical flask to ensure that there were no bubbles. Then, the dissolved oxygen was measured as a function of time under stirring (400 rpm) at 20 °C. The final result was expressed in mg of dissolved oxygen consumed per g of activated carbon per hour  $(mg_{02}/(g_{GAC}\cdot h))$ .

Extraction and detection of EPS, which was defined as the sum of polysaccharides and proteins, were conducted using the standard methods, while the specific detection of polysaccharides and proteins was undertaken using the anthrone/ $H_2SO_4$  and Bradford methods, respectively.

169 Static adsorption experiments were performed on new GAC, used GAC, and 170 used GAC after sterilization to distinguish the adsorption and microbial degradation in 171 each BAF systems. A high pressure steam sterilizer was used to sterilize the used 172 GAC. The operating temperature was 125 °C and the sterilization time was 20 min. In 173 the static adsorption experiment at 20 °C, the dosage of GAC was 2 g/L, the stirring 174 speed was 400 rpm and the running time was 72 h. Reaction mixtures were withdrawn 175 at specific time intervals to measure the variation of DOC and UV<sub>254</sub>.

GAC samples were analyzed using SEM (FE-SEM, Regulug-8230, Hitachi, Japan) and EDS (X-MAX Extreme, Oxford-Instruments, UK) to detect physical and chemical changes on the GAC surface and observe the morphology of biofilm. GAC was prepared for SEM imaging by fixation with 2% glutaraldehyde, dehydration in 20–100% ethanol, and drying in a freeze vacuum dryer. EDS was applied in tandem with SEM to map and evaluate the deposition of elemental content on the GAC surface throughout BAF treatment.

**2.5. Microbial diversity analysis**. Through the microbial diversity sequencing of the raw water, the GAC at the end of BAF column acclimation, and the GAC at different times of continuous-mode BAF systems, the temporal and spatial variation of the microbial community and the dominant functional microorganisms were analyzed. Details about microbial diversity sequencing and analysis are presented in Text S2 of the SI and in our previous study.<sup>35, 39</sup> Note that the same amount of GAC was filled into the reactor after sampling. 190

191

#### **3. RESULTS AND DISCUSSION**

192 3.1. Effect of Pre-oxidation on Water Quality. Pre-oxidation had a negligible 193 effect on the DOC parameter, shown in Figure 1a. The DOC change upon Fe(VI) 194 treatment was -2.1%, while that upon O<sub>3</sub> treatment was +4.1%, caused by the 195 competing effect of organic matter mineralization and solubilization. More explicitly, 196 pre-oxidation only partly mineralizes the organic content, which would translate into a decrease of DOC values. However, this reaction simultaneously increases the 197 solubility of suspended organic matter and promotes the release of intracellular 198 organic substances from sterilized bacteria, with the effect of increasing the DOC.<sup>30</sup> 199 200 The effect of oxidation was instead directly associated with the reduction of other 201 parameters related to organic composition. The  $UV_{254}$  removal rates by  $O_3$  and Fe(VI)were 13% and 23%, respectively (Figure 1b). As shown in Figure 1c, O<sub>3</sub> and Fe(VI) 202 removed part of the COD (9.2%-11.8%) and, most importantly, increased the 203 concentration of BOD<sub>5</sub> (33.2%-22.7%), as well as the value of BOD<sub>5</sub>/COD 204 (47.2%-39.6%), 205 suggesting that pre-oxidation significantly improved the biodegradability of SGW. The composition and relative content of fluorescent organic 206 207 matters in SGW were obtained through fluorescence EEM spectra and FRI analysis method (Figure S4 and Figure 1d).<sup>40</sup> The soluble microbial by-product-like matters 208 209 (region IV) and humic acid-like matters (region V) were the dominant fluorescent 210 organic components in SGW. O<sub>3</sub> had excellent removal effect on all kids of

211 fluorescent organic matters (74.5%), while Fe(VI) only slightly removed fluorescent 212 organic matters (9.4%), mainly acting on soluble microbial by-product-like matters 213 and humic acid-like matters. In summary, the mineralization of organic matters by  $O_3$ 214 or Fe(VI) was limited, while pre-oxidation mainly changed the properties of organic 215 matters.

216 Some interesting phenomena were found in GAC static adsorption experiments 217 of pre-oxidized SGW. In the first 24 h of adsorption, the adsorption rate of DOC in 218 pre-oxidized SGW was practically the same of that measured in raw SGW, as shown 219 in Figure 1f. The same trend was observed for  $UV_{254}$  before and after ozonation 220 (Figure 1g). On the contrary, the adsorption rate and equilibrium adsorption capacity 221 of GAC for UV<sub>254</sub> in SGW treated by Fe(VI) was lower than that assessed in raw SGW. The value of  $UV_{254}$  mainly represents the content of low molecular weight 222 aromatic compounds.<sup>41, 42</sup> This result might indicate that, as this fraction of organic 223 224 matter was removed more efficiently by Fe(VI) than O<sub>3</sub> (Figure 1b), the affinity or the 225 kinetics of adsorption for other fractions was higher upon oxidation with ferrate. In 226 general, these tests suggested a complex effect of pre-oxidation on organic content 227 and composition. Also, the data evidently indicate that pre-oxidation would not 228 translate into better water quality if followed by simple GAC adsorption in the 229 absence of microorganisms. However, the analysis of organic matter biodegradability upon oxidation suggests that the performance of BAF systems may be improved 230 231 compared to a raw water not subjected to this pre-treatment step, and this effect is

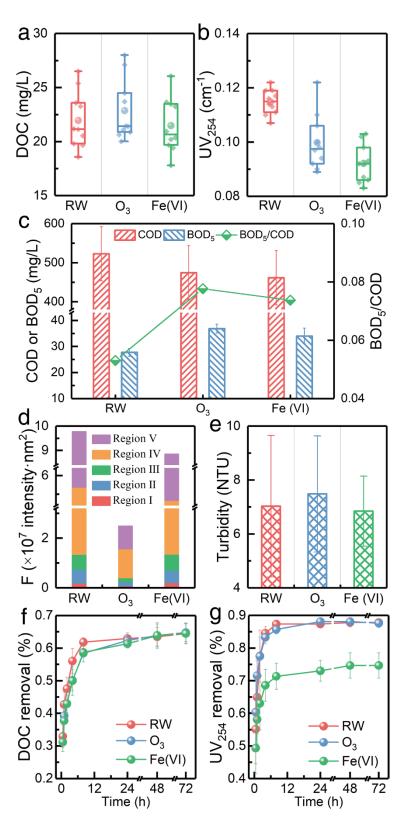


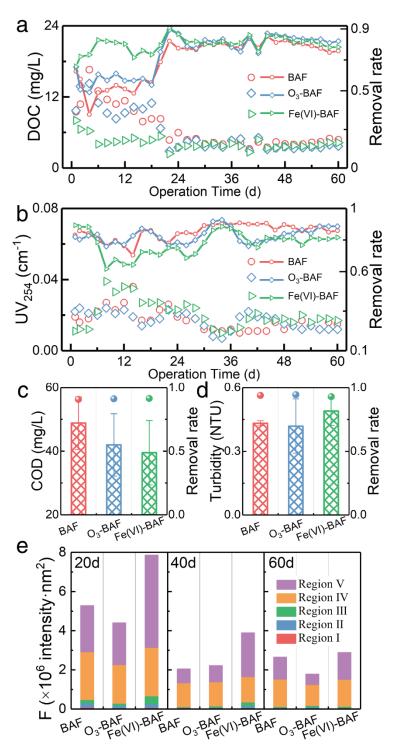
Figure 1. Effect of preoxidation on (a-e) water quality, (f) DOC adsorption on GAC,

and (g)  $UV_{254}$  adsorption on GAC. The dosages of  $O_3$  and Fe(VI) were 80 mg/L and

236 40 mg/L, respectively. The GAC dosage was 2 g/L.

237 3.2. Performance of BAF, O<sub>3</sub>-BAF, and Fe(VI)-BAF. The variation of DOC 238 and UV<sub>254</sub> in three BAF systems effluent measured during 60 days of continuous 239 operation is shown in Figure 2a and Figure 2b. In the first 18 days of continuous 240 operation, the DOC removal rate of the three BAF systems was somewhat erratic with 241 the water pre-treated by Fe(VI) oxidation showing the highest value. With the 242 operation and related strengthening of microbial activity, the removal rate of DOC in 243 all three systems increased and reached stability at values around 80%, specifically with rates that decreased in the order  $O_3$ -BAF > Fe(VI)-BAF > BAF. Similarly, the 244 245 removal rate of UV<sub>254</sub> by three BAF systems was more inconsistent during the initial 246 stage of BAF experiments and reached stability toward the end. Generally, the  $UV_{254}$ 247 removal rate of BAF was the highest, while that of Fe(VI)-BAF was the lowest, 248 consistent with what reported in Figure 1g for static adsorption tests. However, the efficiency of UV<sub>254</sub> removal by Fe (VI)-BAF was gradually improved during 249 250 operation, which may be attributed to the continuous enrichment of microorganisms. Overall, the BAF systems had high COD removal rates (90.7%-91.4%) (Figure 251 252 2c). In particular, the COD content in effluent from BAF, O<sub>3</sub>-BAF, and Fe(VI)-BAF 253 was 48.8, 42.0, and 39.5 mg/L, respectively. The turbidity (0.42-0.49 NTU) of the 254 effluents was also low (Figure 2d). The composition and relative content of fluorescent organic compounds were measured in the effluents of the three BAF 255 256 systems on the 20th day, 40th day, and 60th day of operation, and the results are

shown in Figure 2e and Figure S4. BAF systems efficiently removed fluorescent organic compounds, and the removal rate increased gradually with the operation. The soluble microbial by-product-like matters (region IV) and humic acid-like matters (region V) were the dominant fluorescent organic components in the effluents.



**Figure 2.** Quality of the effluents from the BAF systems. (**a-b**) DOC and UV254 parameters as a function of time during operation: here the larger data points refer to the concentrations (left axis) and the small data points connected by lines to the removal rate (right axis). (**c-d**) COD and turbidity values of the effluent: here, the bars refer to the value of each parameter (left axis) and the circles to the average removal rate (right axis). (**e**) Fluorescent organic components at three moments of the BAF operation.

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270 3.3. Biofilm Morphology and Microbial Activity. The results in Figure 3a and Figure S6 indicate the presence of a large amount of microorganisms and of some 271 272 microbial micelleons on the GAC surface from the three BAF systems. The 273 microorganisms were mainly bacilli, cocci, and filamentous bacteria. Clearly, the 274 microorganisms observed in samples from O<sub>3</sub>-BAF and Fe(VI)-BAF reactors were far more than those observed in the BAF reactor. As shown in Figure 3b and Figure 3c, 275 276 the OUR of microorganisms and the concentration of EPS decreased in the order Fe(VI)-BAF > O<sub>3</sub>-BAF > BAF, corroborating that the activity of the microorganism in 277 BAF systems was higher upon pre-oxidation.<sup>43, 44</sup> 278

The adsorption performance of GAC after long-term operation in BAF systems was always lower than that of the new GAC, as presented in Figure 3d. This result is rationalized with the larger density of available sites for adsorption on pristine GAC. In real operation, the lower adsorption of biologically-enhanced GAC would be

283	compensated by the concurrent degradation of organic matter, which is the main target
284	of this treatment. Interestingly, sterilization of the GAC used in the BAF system
285	without pre-oxidation did not change its DOC removal performance compared to the
286	material analyzed after use and without sterilization. On the contrary, both the
287	adsorption rate and the equilibrium adsorption capacity of sterilized used GAC for
288	DOC were significantly lower than that of used GAC from O <sub>3</sub> -BAF and Fe(VI)-BAF
289	systems. This analysis, combined with the observations from Figure 3a-d, suggests
290	that microorganisms in O <sub>3</sub> -BAF and Fe(VI)-BAF systems were highly functional and
291	played an important role in the removal of organic matter. This result is attributed to
292	the better biodegradability of pre-oxidized SGW, which helped sustaining a healthier
293	and more active microbial community in the BAF systems.

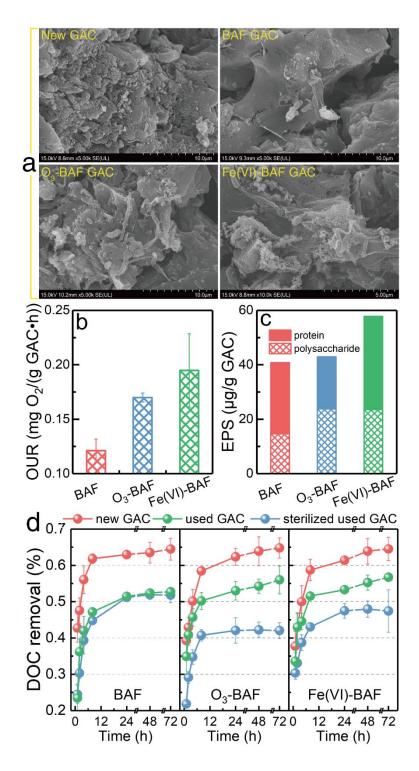




Figure 3. (a) Surface micrographs of new GAC and used GAC from three BAF systems. (b) OUR of microorganisms and (c) EPS concentration in the used GAC biofilm. (d) Variation of DOC removal in static adsorption experiment with new GAC, used GAC, and sterilized used GAC as adsorbents. The used GAC was collected on

the 60th day of continuous operation of the three BAF systems. The magnification of surface micrographs is  $5k \times \text{ or } 10k \times$ . In static adsorption experiment, the GAC dosage was 2 g/L.

302

303 3.4. Microbial Community Analysis. The number of effective sequences, OUTs, alpha diversity indexes, and rarefaction curves for microbial communities in raw 304 water and three BAF systems at different operation times are presented in Table S4 305 306 and Figure S8. The richness and diversity of microbial communities in raw water were higher than those in the three BAF reactors. The coverage values and rarefaction 307 curves suggested the sequencing depth were sufficient.<sup>45, 46</sup> Through principal 308 309 component analysis (PCA) at OUT level (Figure S9), the affinity relationships of 310 microbial community between raw water and three BAF reactors, as well as among 311 the three BAF reactors are revealed. The microbial community in the raw water was 312 vastly different from that in the BAF reactor at 0 day after acclimation process, 313 indicating new dominant microorganisms had been formed in BAF reactors. Also, the microbial communities in the same reactor at different times were similar, and O<sub>3</sub> 314 315 seemed to have an effect in affecting more pronounced changes of the microbial 316 community compared to ferrate pre-treatment.

Figure 4a and Figure 4b show in details the microbial community composition at the phylum and genus level, respectively. *Proteobacteria* (30.4%), *Actinobacteriota* (18.1%), *Bacteroidota* (14.7%), *Firmicutes* (12.3%), *Desulfobacterota* (9.5%), and 320 Synergistota (7.0%) were the major phyla and constituted 92% of bacteria in raw water. Through acclimation, Proteobacteria (98.1%) became the absolute dominant 321 322 microbial phylum in BAF reactors. Similarly, the major genera in raw water, which detected in SGW from shale gas wells,<sup>46-48</sup> 323 were widely also were norank f Coriobacteriaceae (18.0%), Marinobacterium (8.7%), Lentimicrobium 324 (6.0%), Roseovarius (5.0%), and Desulfovibrio (4.6%). Family Coriobacteriaceae is 325 within phylum Actinobacteriota.<sup>23</sup> 326 bacteria an anaerobic fermentative 327 Marinobacterium is a strict aerobe microorganism capable of utilizing a wide range of carbon sources.<sup>49, 50</sup> Lentimicrobium is a strictly anaerobic bacterium with the function 328 of hydrolyzing organics.<sup>51, 52</sup> Almaraz et al.<sup>18</sup> reported *Roseovarius* as an iodine 329 330 oxidation bacterium, which can promote the formation of large amounts of iodinated organic compounds that would cause serious negative implications to the water 331 332 environment. Desulfovibrio as a sulfate-reducing bacterium is widely detected in 333 shale gas wastewater, and is associated with the risk of corrosion to shale gas production facilities.<sup>47, 53, 54</sup> 334

A large amount of relatively low abundance microorganisms were enriched upon acclimation, with the major genus components being *Methyloversatilis* (39.1%), *Rhizobium* (20.0%), *Rehaibacterium* (10.4%), *Acinetobacter* (6.3%), *Pseudomonas* (4.5%), and *Acidovorax* (2.9%). During the BAF tests, the communities adapted differently based on the presence and type of oxidation pre-treatment. Consistent with PCA analysis, the microbial communities in the same BAF reactor at different 341 operation times were similar. Although there were some differences in the microbial community structure of the three BAF systems, the core microorganisms were similar. 342 343 Methyloversatilis, These were Rehaibacterium, Pseudomonas, Rhizobium, 344 Porphyrobacter, Acinetobacter, Bosea, Roseovarius, Acidovorax and Xanthobacter. 345 Methyloversatilis is a salinity tolerant bacterium with the ability of denitrification and organics degradation.<sup>55, 56</sup> Rhizobium are typical denitrifying bacteria, which widely 346 exists in activated sludge, soil, and wastewater.<sup>35, 57, 58</sup> Genus Acinetobacter is related 347 348 rich functions, such as degradation of organics, denitrification, phosphorus removal, and oxidation of heavy metals.<sup>59-62</sup> Members of *Pseudomonas* can degrade organics 349 like toluene and chloroform.<sup>63</sup> Some research shows that members of Acidovorax 350 could conduct heterotrophic denitrification.<sup>64, 65</sup> In general, SGW contains a large 351 number of microorganisms with ability of degrading organic matter, removing 352 nitrogen, and oxidizing heavy metals. Efficiently taking advantage of these 353 microorganisms for biological treatment has great prospects in SGW management. 354 355 Furthermore, a large number of anaerobes were eliminated during BAF acclimation process, and new dominant microorganisms were formed with the variation of 356 environmental factors (aeration, variation of TDS, and the addition of sodium 357 358 acetate).

In order to determine the microorganisms with significant abundance differences between the three BAF systems, we performed biomarker analysis using the linear discriminant analysis effect size (LEfSe) method. As shown in Figure S10, 9 bacterial

362 clades presented statistically significant differences with an LDA threshold of 3.4. 363 Each reactor had its own characteristic microorganisms whose abundance was higher 364 than that of other reactors. Specifically, norank\_o\_Bacteroideres\_VC2\_1\_Bac22 and 365 norank\_f\_Vermiphilaceae were enriched in the BAF reactor without pre-oxidation. Norank\_f\_Rhodospirillaceae, Gemmobacter, and Rhizobium were enriched in 366 367 O<sub>3</sub>-BAF reactor. Instead, Dietzia, Roseovarius, norank\_o\_ Gammaproteobacteria Incertae Sedis, norank\_f\_Rhodobacteraceae 368 and were 369 enriched in the Fe(VI)-BAF reactor.

370 The correlation analysis between microbial community at genus level (top 50) 371 and environmental variables (organic matter removal rate) is shown in Figure 4c. It 372 was found that Rehaibacterium with the high abundance of 3.0-21.1% was significantly correlated with DOC removal rate (P < 0.01). In a previous study, 373 374 Rehaibacterium terrae, a thermotolerant and strictly aerobic bacterium was found in geothermally heated soil of Rehai National Park, China.<sup>66</sup> Rehaibacterium terrae can 375 survive under the conditions of 0-30 g/L NaCl solution and 30-55 °C and degrade 376 some organics.<sup>66</sup> One species in the genus *Rehaibacterium* was detected in our study 377 but could not be defined: the base pair fragments were different from those of 378 379 Rehaibacterium terrae. This result might indicate that a new species of genus 380 Rehaibacterium was present with the strong ability of degrading DOC. Of course, 381 more research is needed to study this hypothesis and to understand the new functions 382 of this putative species. *Mesorhizobium* (0.01-0.20%) were correlated with  $UV_{254}$ 

removal rate (P < 0.05). Research studies reported that *Mesorhizobium* members are halotolerant potential denitrifying bacteria and organics degrading bacteria.<sup>67, 68</sup> In addition, *Labrenzia* (0.01-1.29%), *Magnetospira* (0.01-3.04%), and *SM1A02* (0.01-2.42%) were correlated with COD removal rate (P < 0.05). *Bosea* (0.75-3.58%) and *unclassified\_f\_Rhodobacteraceae* (0.02-1.25%) were correlated with EEM removal rate (P < 0.05). *Methyloversatilis* (2.24-22.31%) was significantly correlated

389 with EEM removal rate (P < 0.01).

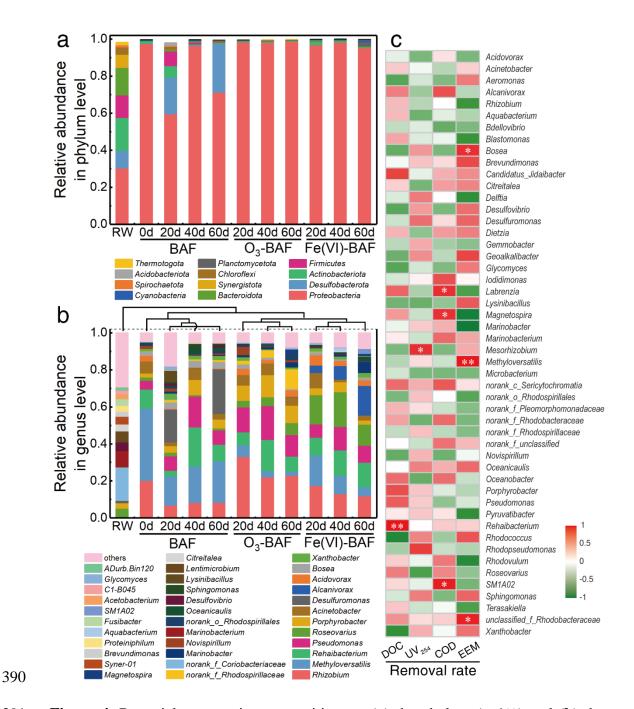


Figure 4. Bacterial community compositions at (a) the phylum (> 1%) and (b) the genus level (> 1.5%) in raw water and the three BAF systems at different operation times. (c) Correlation analysis between microbial community at genus level (top 50) and environmental variables (organic matter removal rate). Here, "\*" represents a value of p < 0.05 and "\*\*" represents a value of p < 0.01.

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397 Implications. O<sub>3</sub> and Fe(VI) pre-oxidation can effectively improve the removal 398 efficiency of organics in BAF, which is attributed to higher activity and content of 399 microorganisms in O<sub>3</sub>-BAF and Fe(VI)-BAF systems compared with BAF. In our 400 experiments, the removal rate of organic matters by BAF systems gradually increased 401 and stabilized, owing to the enhancement of the microbial degradation function, with 402 the enrichment of a large number of microorganisms with specific functions, such as organic matter degradation, nitrogen removal, heavy metals oxidation. For example, 403 404 *Rehaibacterium* is significantly correlated with DOC removal rate (P < 0.01). Besides, Methyloversatilis is significantly correlated with fluorescent organics removal (P < 405 0.01). 406

407 The oxidation behavior of  $O_3$  is different from that of Fe(VI), but both processes can effectively improve the biodegradability of wastewater. However, the 408 409 mineralization rate and the improvement of organic quality in the effluent of systems upon pre-oxidation with O<sub>3</sub> and Fe(VI) were still limited. Combination with other 410 411 oxidants (such as  $H_2O_2$ ) or with electrooxidation may further improve the oxidation 412 efficiency and the efficiency of the BAF process. Ozonation has feasibility and 413 application value in the treatment of SGW, already at this stage of its technological 414 development. Compared with pre-ozonation, Fe(VI) pre-oxidation has the potential 415 advantages of easy operation and maintenance, but its application is still under development and should be optimized.<sup>32</sup> The in-situ Fe(VI) synthesis in wastewater 416 417 treatment plant through wet chemical or electrochemical method is expected to further

418	reduce the chemical cost. <sup>69</sup> It should be noted that the results presented here were
419	obtained at the lab scale, while further studies are needed to evaluate the relevant

420 systems at the pilot and full scales in long-term operation.

421

#### 422 ASSOCIATED CONTENT

#### 423 Supporting Information

424 The Supporting Information is available free of charge on the ACS Publications

425 website.

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#### 441 **REFERENCES**

Zou, C.; Ni, Y.; Li, J.; Kondash, A.; Coyte, R.; Lauer, N.; Cui, H.; Liao, F.;
Vengosh, A., The water footprint of hydraulic fracturing in Sichuan Basin, China. *Sci. Total Environ.* 2018, *630*, 349-356.

445 2. Kondash, A. J.; Lauer, N. E.; Vengosh, A., The intensification of the water 446 footprint of hydraulic fracturing. *Sci. Adv.* **2018**, *4*, (8), eaar5982.

Acharya, S. M.; Chakraborty, R.; Tringe, S. G., Emerging Trends in Biological
Treatment of Wastewater From Unconventional Oil and Gas Extraction. *Front. Microbiol.* 2020, *11*, (2203).

4. Chang, H.; Li, T.; Liu, B.; Vidic, R. D.; Elimelech, M.; Crittenden, J. C., Potential
and implemented membrane-based technologies for the treatment and reuse of
flowback and produced water from shale gas and oil plays: A review. *Desalination* **2019**, *455*, 34-57.

Tong, T.; Carlson, K. H.; Robbins, C. A.; Zhang, Z.; Du, X., Membrane-based
treatment of shale oil and gas wastewater: The current state of knowledge. *Front. Env. Sci. Eng.* 2019, *13*, (4), 63.

457 6. Riley, S. M.; Oliveira, J. M. S.; Regnery, J.; Cath, T. Y., Hybrid membrane
458 bio-systems for sustainable treatment of oil and gas produced water and fracturing
459 flowback water. *Sep. Purif. Technol.* 2016, *171*, 297-311.

460 7. Butkovskyi, A.; Bruning, H.; Kools, S. A. E.; Rijnaarts, H. H. M.; Van Wezel, A.
461 P., Organic pollutants in shale gas flowback and produced waters: Identification,
462 potential ecological impact, and implications for treatment strategies. *Environ. Sci.*463 *Technol.* 2017, *51*, (9), 4740-4754.

464 8. Camarillo, M. K.; Domen, J. K.; Stringfellow, W. T., Physical-chemical
465 evaluation of hydraulic fracturing chemicals in the context of produced water
466 treatment. *J. Environ. Manag.* 2016, *183*, 164-174.

467 9. Zhang, X.; Chen, A.; Zhang, D.; Kou, S.; Lu, P., The treatment of flowback water
468 in a sequencing batch reactor with aerobic granular sludge: Performance and
469 microbial community structure. *Chemosphere* 2018, *211*, 1065-1072.

470 10. Frank, V. B.; Regnery, J.; Chan, K. E.; Ramey, D. F.; Spear, J. R.; Cath, T. Y.,
471 Co-treatment of residential and oil and gas production wastewater with a hybrid
472 sequencing batch reactor-membrane bioreactor process. *J. Water Process Eng.* 2017,
473 17, 82-94.

474 11. Sitterley, K. A.; Silverstein, J.; Rosenblum, J.; Linden, K. G., Aerobic biological
475 degradation of organic matter and fracturing fluid additives in high salinity hydraulic
476 fracturing wastewaters. *Sci. Total Environ.* 2020, 143622.

477 12. Kose Mutlu, B.; Ozgun, H.; Ersahin, M. E.; Kaya, R.; Eliduzgun, S.; Altinbas, M.;

478 Kinaci, C.; Koyuncu, I., Impact of salinity on the population dynamics of

479 microorganisms in a membrane bioreactor treating produced water. *Sci. Total Environ.*480 **2019**, *646*, 1080-1089.

481 13. Sharghi, E. A.; Bonakdarpour, B.; Roustazade, P.; Amoozegar, M. A.; Rabbani, A.

482 R., The biological treatment of high salinity synthetic oilfield produced water in a

- 483 submerged membrane bioreactor using a halophilic bacterial consortium. J. Chem.
  484 *Technol. Biot.* 2013, 88, (11), 2016-2026.
- 485 14. Abass, O. K.; Zhang, K., Nano-Fe mediated treatment of real hydraulic fracturing
  486 flowback and its practical implication on membrane fouling in tandem anaerobic-oxic
  487 membrane bioreactor. *J. Hazard. Mater.* 2020, *395*, 122666.
- 488 15. Akyon, B.; Stachler, E.; Wei, N.; Bibby, K., Microbial Mats as a Biological
  489 Treatment Approach for Saline Wastewaters: The Case of Produced Water from
  490 Hydraulic Fracturing. *Environ. Sci. Technol.* 2015, 49, (10), 6172-6180.
- 491 16. Akyon, B.; McLaughlin, M.; Hernández, F.; Blotevogel, J.; Bibby, K.,
  492 Characterization and biological removal of organic compounds from hydraulic
  493 fracturing produced water. *Environ. Sci. Proc. Imp.* 2019, *21*, (2), 279-290.
- 494 17. Zhuang, Y.; Zhang, Z.; Zhou, Z.; Chen, M.; Li, J.; Chen, S., Co-treatment of
  495 shale-gas produced water and municipal wastewater: Removal of nitrogen in a
  496 moving-bed biofilm reactor. *Process Saf. Environ. Prot.* 2019, *126*, 269-277.
- 497 18. Almaraz, N.; Regnery, J.; Vanzin, G. F.; Riley, S. M.; Ahoor, D. C.; Cath, T. Y.,
  498 Emergence and fate of volatile iodinated organic compounds during biological
  499 treatment of oil and gas produced water. *Sci. Total Environ.* 2020, 699, 134202.
- 500 19. Freedman, D. E.; Riley, S. M.; Jones, Z. L.; Rosenblum, J. S.; Sharp, J. O.; Spear, 501 J. R.; Cath, T. Y., Biologically active filtration for fracturing flowback and produced
- 502 water treatment. J. Water Process Eng. 2017, 18, 29-40.
- 20. Riley, S. M.; Ahoor, D. C.; Cath, T. Y., Enhanced biofiltration of O&G produced
  water comparing granular activated carbon and nutrients. *Sci. Total Environ.* 2018,
  640-641, 419-428.
- Siley, S. M.; Ahoor, D. C.; Regnery, J.; Cath, T. Y., Tracking oil and gas
  wastewater-derived organic matter in a hybrid biofilter membrane treatment system: A
  multi-analytical approach. *Sci. Total Environ.* 2018, *613-614*, 208-217.
- 509 22. Zhang, X.; Zhang, D.; Huang, Y.; Wu, S.; Lu, P., The anodic potential shaped a 510 cryptic sulfur cycling with forming thiosulfate in a microbial fuel cell treating 511 hydraulic fracturing flowback water. *Water Res.* **2020**, *185*, 116270.
- 512 23. Zhang, X.; Zhang, D.; Huang, Y.; Zhang, K.; Lu, P., Simultaneous removal of 513 organic matter and iron from hydraulic fracturing flowback water through sulfur 514 cycling in a microbial fuel cell. *Water Res.* **2018**, *147*, 461-471.
- 515 24. Forrestal, C.; Haeger, A.; Dankovich Iv, L.; Cath, T. Y.; Ren, Z. J., A liter-scale
- 516 microbial capacitive deionization system for the treatment of shale gas wastewater.
  517 *Environ. Sci. Water Res. Technol.* 2016, 2, (2), 353-361.
- 518 25. Forrestal, C.; Stoll, Z.; Xu, P.; Ren, Z. J., Microbial capacitive desalination for 519 integrated organic matter and salt removal and energy production from 520 unconventional natural gas produced water. *Environ. Sci. Water Res. Technol.* **2015**, *1*, 521 (1), 47-55.
- 522 26. Shrestha, N.; Chilkoor, G.; Wilder, J.; Ren, Z. J.; Gadhamshetty, V., Comparative
- 523 performances of microbial capacitive deionization cell and microbial fuel cell fed with
- 524 produced water from the Bakken shale. *Bioelectrochemistry* **2018**, *121*, 56-64.
- 525 27. Gomes, J.; Costa, R.; Quinta-Ferreira, R. M.; Martins, R. C., Application of

- 526 ozonation for pharmaceuticals and personal care products removal from water. *Sci.*527 *Total Environ.* 2017, 586, 265-283.
- 528 28. Wang, W.-L.; Cai, Y.-Z.; Hu, H.-Y.; Chen, J.; Wang, J.; Xue, G.; Wu, Q.-Y.,
- 529 Advanced treatment of bio-treated dyeing and finishing wastewater using
- ozone-biological activated carbon: A study on the synergistic effects. *Chem. Eng. J.* **2019**, *359*, 168-175.
- 532 29. Sharma, V. K.; Zboril, R.; Varma, R. S., Ferrates: Greener Oxidants with 533 Multimodal Action in Water Treatment Technologies. *Acc. Chem. Res.* **2015**. *48*. (2).
- Multimodal Action in Water Treatment Technologies. Acc. Chem. Res. 2015, 48, (2),
  182-191.
- 535 30. Tang, P.; Liu, B.; Zhang, Y.; Chang, H.; Zhou, P.; Feng, M.; Sharma, V. K.,
- Sustainable reuse of shale gas wastewater by pre-ozonation with ultrafiltration-reverse
  osmosis. *Chem. Eng. J.* 2020, *392*, 123743.
- 538 31. Zhu, J.-H.; Yan, X.-L.; Liu, Y.; Zhang, B., Improving alachlor biodegradability by 539 ferrate oxidation. *J. Hazard. Mater.* **2006**, *135*, (1), 94-99.
- 540 32. Ma, J.; Li, C.; Zhang, Y.; Ju, R., Combined Process of Ferrate Preoxidation and
- 541 Biological Activated Carbon Filtration for Upgrading Water Quality. In *Ferrates*,
  542 American Chemical Society: 2008; Vol. 985, pp 446-455.
- 543 33. Li, M.; Liang, B.; Shang, J.; Li, J.; Zhang, H., Treatment of Polysilicon
  544 Production Wastewater by Ferrate(VI) Microcapsule Oxidation and Biological
  545 Aerated Biofilter. *Water Air Soil Pollut.* 2019, 230, (11), 254.
- 546 34. Shang, W.; Tiraferri, A.; He, Q.; Li, N.; Chang, H.; Liu, C.; Liu, B., Reuse of 547 shale gas flowback and produced water: Effects of coagulation and adsorption on 548 ultrafiltration, reverse osmosis combined process. *Sci. Total Environ.* **2019**, *689*, 549 47-56.
- 550 35. Tang, P.; Li, J.; Li, T.; Tian, L.; Sun, Y.; Xie, W.; He, Q.; Chang, H.; Tiraferri, A.;
- 551 Liu, B., Efficient integrated module of gravity driven membrane filtration, solar
- aeration and GAC adsorption for pretreatment of shale gas wastewater. J. Hazard. *Mater.* 2020, 124166.
- 554 36. Schreyer, J. M.; Thompson, G. W.; Ockerman, L. T., Ferrate Oxidimetry. *Anal.*555 *Chem.* **1950**, *22*, (5), 691-692.
- 37. Thompson, G. W.; Ockerman, L. T.; Schreyer, J. M., Preparation and Purification
  of Potassium Ferrate. VI. *J. Am. Chem. Soc.* 1951, *73*, (3), 1379-1381.
- 558 38. Li, C.; Li, X. Z.; Graham, N., A study of the preparation and reactivity of 559 potassium ferrate. *Chemosphere* **2005**, *61*, (4), 537-543.
- 560 39. Chang, H.; Liu, B.; Wang, H.; Zhang, S. Y.; Chen, S.; Tiraferri, A.; Tang, Y. Q.,
- 561 Evaluating the performance of gravity-driven membrane filtration as desalination 562 pretreatment of shale gas flowback and produced water. *J. Membr. Sci.* **2019**, *587*, 563 117187.
- 40. Chen, W.; Westerhoff, P.; Leenheer, J. A.; Booksh, K., Fluorescence excitation-emission matrix regional integration to quantify spectra for dissolved organic matter. *Environ. Sci. Technol.* **2003**, *37*, (24), 5701-5710.
- 41. Acero, J. L.; Benitez, F. J.; Real, F. J.; Teva, F., Micropollutants removal from
   retentates generated in ultrafiltration and nanofiltration treatments of municipal
   30

- secondary effluents by means of coagulation, oxidation, and adsorption processes. *Chem. Eng. J.* 2016, 289, 48-58.
- 42. Wang, X.; Xia, J.; Ding, S.; Zhang, S.; Li, M.; Shang, Z.; Lu, J.; Ding, J.,
  Removing organic matters from reverse osmosis concentrate using advanced
  oxidation-biological activated carbon process combined with Fe3+/humus-reducing
  bacteria. *Ecotox. Environ. Safe.* 2020, 203, 110945.
- 43. Guwy, A. J.; Buckland, H.; Hawkes, F. R.; Hawkes, D. L., Active biomass in
  activated sludge: Comparison of respirometry with catalase activity measured using
  an on-line monitor. *Water Res.* 1998, *32*, (12), 3705-3709.
- 578 44. Shi, Y.; Huang, J.; Zeng, G.; Gu, Y.; Chen, Y.; Hu, Y.; Tang, B.; Zhou, J.; Yang, Y.;
- Shi, L., Exploiting extracellular polymeric substances (EPS) controlling strategies for
  performance enhancement of biological wastewater treatments: An overview. *Chemosphere* 2017, *180*, 396-411.
- 45. Fang, D.; Zhao, G.; Xu, X.; Zhang, Q.; Shen, Q.; Fang, Z.; Huang, L.; Ji, F.,
  Microbial community structures and functions of wastewater treatment systems in
  plateau and cold regions. *Bioresour. Technol.* 2018, 249, 684-693.
- 46. Wang, H.; Lu, L.; Chen, X.; Bian, Y.; Ren, Z. J., Geochemical and microbial characterizations of flowback and produced water in three shale oil and gas plays in the central and western United States. *Water Res.* **2019**, *164*, 114942.
- 588 47. Zhang, Y.; Yu, Z.; Zhang, H.; Thompson, I. P., Microbial distribution and 589 variation in produced water from separators to storage tanks of shale gas wells in 590 Sichuan Basin, China. *Environ. Sci. Water Res. Technol.* **2017**, *3*, (2), 340-351.
- 48. Cluff, M. A.; Hartsock, A.; MacRae, J. D.; Carter, K.; Mouser, P. J., Temporal
  Changes in Microbial Ecology and Geochemistry in Produced Water from
  Hydraulically Fractured Marcellus Shale Gas Wells. *Environ. Sci. Technol.* 2014, 48,
  (11), 6508-6517.
- 49. Murali Mohan, A.; Hartsock, A.; Hammack, R. W.; Vidic, R. D.; Gregory, K. B. J.
  F. m. e., Microbial communities in flowback water impoundments from hydraulic
- 597 fracturing for recovery of shale gas. *FEMS Microbiol. Ecol.* **2013**, *86*, (3), 567-580.
- 598 50. GONZÁLEZ, J. M.; MAYER, F.; MORAN, M. A.; HODSON, R. E.; WHITMAN,
- W. B., Microbulbifer hydrolyticus gen. nov., sp. nov., and Marinobacterium
  georgiense gen. nov., sp. nov., Two Marine Bacteria from a Lignin-Rich Pulp Mill
  Waste Enrichment Community. *Int. J. Syst. Evol. Micr.* 1997, 47, (2), 369-376.
- 51. Wang, H.; Chen, N.; Feng, C.; Deng, Y.; Gao, Y., Research on efficient
  denitrification system based on banana peel waste in sequencing batch reactors:
  Performance, microbial behavior and dissolved organic matter evolution. *Chemosphere* 2020, 253, 126693.
- 52. Sun, L.; Toyonaga, M.; Ohashi, A.; Tourlousse, D. M.; Matsuura, N.; Meng, X.-Y.;
- 607 Tamaki, H.; Hanada, S.; Cruz, R.; Yamaguchi, T.; Sekiguchi, Y., Lentimicrobium
- 608 saccharophilum gen. nov., sp. nov., a strictly anaerobic bacterium representing a new
- family in the phylum Bacteroidetes, and proposal of Lentimicrobiaceae fam. nov. Int.
- 610 J. Syst. Evol. Micr. 2016, 66, (7), 2635-2642.
- 611 53. Suri, N.; Gassara, F.; Stanislav, P.; Voordouw, G., Microbially Enhanced Oil

- 612 Recovery by Alkylbenzene-Oxidizing Nitrate-Reducing Bacteria. Front. Microbiol.
- 2019, 10, (1243). 613
- 54. Struchtemeyer, C. G.; Davis, J. P.; Elshahed, M. S., Influence of the Drilling Mud 614
- 615 Formulation Process on the Bacterial Communities in Thermogenic Natural Gas Wells of the Barnett Shale. Appl. Environ. Microb. 2011, 77, (14), 4744.
- 616
- 55. Lu, Z.; Sun, W.; Li, C.; Ao, X.; Yang, C.; Li, S., Bioremoval of non-steroidal 617 anti-inflammatory drugs by Pseudoxanthomonas sp. DIN-3 isolated from biological 618 619 activated carbon process. Water Res. 2019, 161, 459-472.
- 620 56. Xu, L.; Graham, N. J. D.; Wei, C.; Zhang, L.; Yu, W., Abatement of the 621 biofouling: Performance of integrated membrane an in-situ 622 bioelectrochemical-ultrafiltration system. Water Res. 2020, 179, 115892.
- 57. Kinh, C. T.; Suenaga, T.; Hori, T.; Riya, S.; Hosomi, M.; Smets, B. F.; Terada, A., 623
- Counter-diffusion biofilms have lower N2O emissions than co-diffusion biofilms 624 625 during simultaneous nitrification and denitrification: Insights from depth-profile
- 626 analysis. Water Res. 2017, 124, 363-371.
- 58. Nikolova, C.; Gutierrez, T., Use of Microorganisms in the Recovery of Oil From 627 628 Recalcitrant Oil Reservoirs: Current State of Knowledge, Technological Advances and 629 Future Perspectives. Front. Microbiol. 2020, 10, (2996).
- 630 59. Oberoi, A. S.; Jia, Y.; Zhang, H.; Khanal, S. K.; Lu, H., Insights into the Fate and 631 Removal of Antibiotics in Engineered Biological Treatment Systems: A Critical 632 Review. Environ. Sci. Technol. 2019, 53, (13), 7234-7264.
- 633 60. Ren, B.; Li, C.; Zhang, X.; Zhang, Z., Fe(II)-dosed ceramic membrane bioreactor for wastewater treatment: Nutrient removal, microbial community and membrane 634 635 fouling analysis. Sci. Total Environ. 2019, 664, 116-126.
- 61. Momba, M. N. B.; Cloete, T. E., The relationship of biomass to phosphate uptake 636
- 637 by Acinetobacter junii in activated sludge mixed liquor. Water Res. 1996, 30, (2), 364-370. 638
- 639 62. Tang, X.; Xie, B.; Chen, R.; Wang, J.; Huang, K.; Zhu, X.; Li, G.; Liang, H., 640 Gravity-driven membrane filtration treating manganese-contaminated surface water:
- 641 Flux stabilization and removal performance. Chem. Eng. J. 2020, 397, 125248.
- 642 63. Liden, T.; Santos, I. C.; Hildenbrand, Z. L.; Schug, K. A., Treatment modalities 643 for the reuse of produced waste from oil and gas development. Sci. Total Environ. 644 **2018**, *643*, 107-118.
- 645 64. Willems, A.; Falsen, E.; Pot, B.; Jantzen, E.; Hoste, B.; Vandamme, P.; Gillis, M.;
- Kersters, K.; Deley, J., Acidovorax, A New Genus For Pseudomonas-Facilis, 646
- 647 Pseudomonas-Delafieldii, E-Falsen (Ef) Group 13, Ef Group 16, And Several Clinical
- Isolates, With The Species Acidovorax-Facilis Comb-Nov, Acidovorax-Delafieldii 648
- 649 Comb-Nov, And Acidovorax-Temperans Sp-Nov. Int. J. Syst. Bacteriol. 1990, 40, (4),
- 650 384-398.
- 651 65. Yang, Y.; Chen, T.; Zhang, X.; Qing, C.; Wang, J.; Yue, Z.; Liu, H.; Yang, Z.,
- 652 Simultaneous removal of nitrate and phosphate from wastewater by siderite based
- 653 autotrophic denitrification. Chemosphere 2018, 199, 130-137.
- 654 66. Yu, T.-T.; Yao, J.-C.; Yin, Y.-R.; Dong, L.; Liu, R.-F.; Ming, H.; Zhou, E.-M.; Li,

- W.-J., Rehaibacterium terrae gen. nova, sp nov isolated from a geothermally heated soil sample. *Int. J. Syst. Evol. Micr.* **2013**, *63*, 4058-4063.
- 657 67. Luján-Facundo, M. J.; Fernández-Navarro, J.; Alonso-Molina, J. L.;
- 658 Amorós-Muñoz, I.; Moreno, Y.; Mendoza-Roca, J. A.; Pastor-Alcañiz, L., The role of
- salinity on the changes of the biomass characteristics and on the performance of an
- 660 OMBR treating tannery wastewater. *Water Res.* **2018**, *142*, 129-137.
- 661 68. Xu, J.; Gao, W.; Zhao, B.; Chen, M.; Ma, L.; Jia, Z.; Zhang, J., Bacterial 662 community composition and assembly along a natural sodicity/salinity gradient in 663 surface and subsurface soils. *Appl. Soil Ecol.* **2021**, *157*, 103731.
- 664 69. Ni, B.-J.; Yan, X.; Dai, X.; Liu, Z.; Wei, W.; Wu, S.-L.; Xu, Q.; Sun, J., Ferrate
- 665 effectively removes antibiotic resistance genes from wastewater through combined
- effect of microbial DNA damage and coagulation. *Water Res.* **2020**, *185*, 116273.