

Review of biochar role as additive in anaerobic digestion processes

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Review of biochar role as additive in anaerobic digestion processes / Chiappero, Marco; Norouzi, Omid; Hu, Mingyu; Demichelis, Francesca; Berruti, Franco; Di Maria, Francesco; Mašek, Ondej; Fiore, Silvia. - In: RENEWABLE & SUSTAINABLE ENERGY REVIEWS. - ISSN 1364-0321. - STAMPA. - 131:(2020), p. 110037.
[10.1016/j.rser.2020.110037]

Availability:

This version is available at: 11583/2840328 since: 2020-07-15T15:51:26Z

Publisher:

Elsevier

Published

DOI:10.1016/j.rser.2020.110037

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<http://dx.doi.org/10.1016/j.rser.2020.110037>

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1 **Review of biochar role as additive in anaerobic digestion processes**

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16

17 **Abstract**

18 Anaerobic digestion (AD) could be considered as a mature technology and nowadays it

19 can still play a pivot role because of the urgent need to provide renewable energy sources

20 and efficiently manage the continuously growing amount of organic waste. Biochar (BC)

21 is an extremely versatile material, which could be produced by carbonization of organic

22 materials, including biomass and wastes, consistently with Circular Economy principles,

23 and “tailor-made” for specific applications. The potential BC role as additive in the

24 control of the many well-known critical issues of AD processes has been increasingly

25 explored over the past few years. However, a clear and comprehensive understanding of
26 the connections between BC and AD is still missing. This review paper analyses and
27 discusses significant references (review articles, research papers and international
28 databases and reports), mostly published in the last 10 years. This review is aimed at
29 addressing three key issues related to the better understanding of the BC role in AD
30 processes: 1. Investigation of the influence of BC properties on AD performances and of
31 their ability to counteract its main challenges; 2. Assessment of the optimal BC
32 production chain (i.e. feedstock-pyrolysis-activation) to achieve the desired features; 3.
33 Evaluation of the economic and environmental advantages connected to BC use in AD
34 processes, compared to conventional solutions applied to address AD challenges.

35

36 **Highlights**

37 - Biochar key properties were specific surface, pores and surface functional groups

38 - Lignocellulosic biomass, slow pyrolysis and physical activation made the best biochar

39 - Biochar addition exhibited economic and environmental advantages

40 - Biochar improved methane production (in average 25 %) and digestate quality

41

42 **Keywords:** anaerobic digestion; biochar; bioenergy; biomass; circular economy; waste

43

44 **Abbreviations**

45 AD: anaerobic digestion; BC: biochar; BET: Brunauer–Emmett–Teller; CE: circular

46 economy; CEC: cationic exchange capacity; CIC: controlling internal circulation; CLSM:

47 confocal laser scanning microscopy; COD: chemical oxygen demand; DIET: direct

48 interspecies electron transfer; EAC: electron accepting capacity; EDC: electron donating

49 capacity; EU: European union; FAN: free ammonia nitrogen; FISH: Fluorescence in situ

50 hybridization; FW: food waste; GAC: granular activated carbon; GHG: green-house gas;
 51 HRT: hydraulic retention time; IET: interspecies electron transfer; LCA: life cycle
 52 analysis; MSW: municipal solid waste; OFMSW: organic fraction of municipal solid
 53 waste; OLR: organic loading rate; PAH: polyaromatic hydrocarbons; PCBs:
 54 polychlorinated biphenyls; S/I: substrate to inoculum ratio; SMP: specific methane
 55 potential; SS: suspended solids; SSA: specific surface area; TAN: total ammonia
 56 nitrogen; TPAD: two phased anaerobic digestion; TSS: total suspended solids; UASB:
 57 up-flow anaerobic sludge blanket; VFA: volatile fatty acids; WAS: waste activated
 58 sludge; WWTP: wastewater treatment plant; WV: working volume; V: volume.

59

60 Contents

| | |
|--|----|
| 61 Abbreviations | 2 |
| 62 1. Introduction | 4 |
| 63 2. Review methodology..... | 7 |
| 64 3. Biochar..... | 10 |
| 65 3.1. Feedstocks | 13 |
| 66 3.2. Thermochemical production..... | 14 |
| 67 3.3. Activation..... | 16 |
| 68 3.4. Physico-chemical characteristics..... | 17 |
| 69 4. Application of biochar as additive in anaerobic digestion..... | 21 |
| 70 4.1. Buffer capacity and alkalinity..... | 21 |
| 71 4.2. Adsorption of inhibitors..... | 25 |
| 72 4.3. Effects on microbial populations..... | 33 |
| 73 4.3.2. Selective colonization of functional microbes..... | 34 |
| 74 4.3.3. Promotion of syntrophic metabolisms..... | 35 |
| 75 4.4. Effect of biochar on digestate quality..... | 53 |
| 76 4.5. Effects on biogas upgrading | 56 |
| 77 4.5.1. In-situ biogas upgrading..... | 56 |
| 78 4.5.2. Ex-situ biogas cleaning and upgrading..... | 57 |
| 79 5. Economic and environmental assessments..... | 61 |
| 80 6. Conclusions | 68 |
| 81 | |

82 1. Introduction

83 The global energy consumption increased from around 8,800 million tonnes of oil
84 equivalents (Mtoe, including coal, gas, oil, electricity, heat and biomass) in 1990 to
85 14,400 Mtoe in 2018 [1]. In 2016 renewable energy sources met this demand for about
86 14 % of primary energy supply, while fossil fuels accounted for 81% of greenhouse gas
87 (GHG) emissions [2]. The need to decrease fossil fuel dependency and GHG emissions
88 and to generate economic benefits is stimulating initiatives to produce energy as well as
89 high-value chemicals and products from sustainable non-food biomass, residues, co-
90 products and wastes.

91 Anaerobic digestion (AD) is a well-established technology for the treatment of
92 wastewater and organic waste, e.g. the organic fraction of the municipal solid waste
93 (OFMSW), waste activated sludge (WAS), animal manure, etc. Biogas can directly
94 replace fossil fuels in heat and power generation or be upgraded to bio-methane as vehicle
95 fuel or injected in the gas grid [3], while digestate can be employed as soil amendment
96 [4,5]. Other benefits of AD, compared to alternative treatment options applied to organic
97 waste, include control of odor and GHG emissions, removal of pathogens and adaptability
98 to many substrates [6]. AD has been extensively implemented, from small-scale
99 applications, particularly in rural areas and developing countries, to large-scale industrial
100 plants [7]. Despite the rapidly growing number of biogas installations in Europe (from
101 about 6,200 to 18,202 between 2009 and 2018) [8], many challenges regarding AD
102 processes still persist. Different options have been explored to address these challenges:
103 optimization of working parameters and bio-reactor configuration, co-digestion and
104 nutrients control [9]; adoption of pre-treatments to enhance the degradability of various
105 substrates [10,11]; use of inorganic and biological additives to support biomass

106 immobilization, supplement nutrients, mitigate inhibitors and improve process stability
107 [12–14].

108 Among the additives, conductive carbonaceous materials, as granular activated carbon
109 (GAC) and biochar (BC), gained interest because of their ability of enhancing methane
110 production [15,16]. BC may present several advantages over other additives; it can be
111 produced with a wide variety of physico-chemical properties by controlling feedstock,
112 pyrolysis operating conditions and the activation process [17,18], resulting “tailor-made”
113 for specific applications. In recent years, several authors confirmed the possibility of
114 increasing methane production through BC addition (Table 1), suggesting different
115 potential mechanisms: (1) increase of the buffering capacity of the AD system; (2)
116 mitigation of inhibition phenomena or agents; (3) support media for biomass
117 immobilization; (4) promotion of syntrophic metabolisms; (5) enhancement of digestate
118 quality; (6) biogas cleaning and upgrading. The most significant and updated scientific
119 literature investigating the BC role within AD processes can be classified as follows
120 (Table 1):

- 121 - Studies considering the advantages of coupling AD with thermal processes
122 (gasification, pyrolysis and hydrothermal carbonization) from a general point of view;
- 123 - Studies exploring the positive influence of BC on AD, based on methane and/or biogas
124 production enhancement and/or improvement of digestate quality;
- 125 - Studies analysing in detail the BC features that may influence one or more of the
126 above-mentioned mechanisms.

127 Considering the digestate, BC has been reported to enhance its quality in terms of nutrient
128 retention, increase of the carbon-to-nitrogen ratio and reduction of nutrient leaching [19].

129 Consequently, BC doesn't need to be separated from digestate at the end of its utilization
130 in AD.

131

132 Table 1. Overview of literature studies exploring the connections between BC and AD

| Studies considering the general advantages of coupling AD with thermal processes | Studies exploring the positive influence of BC on AD, based on a general analysis of methane and/or biogas production enhancement | Studies exploring the positive influence of BC on AD, based on a general analysis of the improvement of digestate quality | Studies analysing in details the biochar features that may influence one or more of the six above-mentioned mechanisms |
|---|--|---|--|
| Pecchi and Baratieri, 2019 [20] Fabbri and Torri, 2016 [21] Hübner and Mumme, 2015 [22] Salman et al., 2017 [23] | Linville et al., 2017 [24] Torri and Fabbri, 2014 [25] Sunyoto et al., 2016 [26] Jang et al., 2018 [27] Sun et al., 2019 [28] Zhao et al., 2015 [29] Inthapanya et al., 2012 [30] Inthapanya and Preston, 2013 [31] | Bruun et al., 2011 [32] Shen et al., 2016 [33] Shen et al., 2017 [34] | Fagbohunge et al., 2017 [19] Ye et al., 2018 [13] González et al., 2018 [15] Zhang et al., 2018 [16] Codignole Luz et al., 2018 [35] Masebinu et al., 2019 [36] Pelaez-Samaniego et al., 2018 [37] G. Wang et al., 2018b [38] D. Wang et al., 2017 [39] Fagbohunge et al., 2016 [40] Mumme et al., 2014 [41] Shanmugam et al., 2018 [42] Su et al., 2019 [43] Lü et al., 2016 [44] Cruz Viggi et al., 2017 [45] Li et al., 2018 [46] Martínez et al., 2018 [47] Luo et al., 2015 [48] Wang et al., 2019 [49] C. Wang et al., 2018 [50] Zhao et al., 2016 [51] Yu et al., 1999 [52] Cooney et al., 2016 [53] Dang et al., 2017 [54] Sasaki et al., 2010 [55] Martins et al., 2018 [56] S. Chen et al., 2014 [57] Pan et al., 2019b [58] Qiu et al., 2019 [59] |

133

134

135 To the best of our knowledge, despite the growing number of studies investigating the
136 connections between BC and AD, three main issues still need to be explored: a clear
137 comprehension of the above-mentioned six mechanisms; a correlation between BC
138 features and their effects on AD processes; environmental and economic outcomes related
139 to BC use. The present review is therefore aimed at addressing the following issues: 1)
140 investigation of the influence of BC properties on AD performance and of their ability to
141 counteract its main challenges; 2) assessment of the optimal BC production chain (i.e.
142 feedstock-pyrolysis-activation) to achieve the desired features; 3) evaluation of the
143 economic and environmental advantages connected to BC use in AD processes, compared
144 to conventional solutions applied to address AD challenges.

145 **2. Review methodology**

146 The selection of scientific literature was made considering the following criteria:

147 a) *Relevant international databases and information sources.* Bibliometric sources as
148 Web of Science, Science Direct and Scopus were used to retrieve articles, book-chapters
149 and proceedings of indexed conferences. Official international databases (Eurostat,
150 International Energy Agency, European Biogas Association, etc.) were searched for the
151 selection of technical reports and statistics;

152 b) *Chronological order.* The majority of the references in this review are from 2015 to
153 2019 (67 %) and from 2009 to 2014 (26 %), with the balance before 2009 (7 %);

154 c) *Relevant keywords for the topics of interest.* A first survey adopted the following
155 keywords in different combinations: “biochar, anaerobic digestion, methane, pyrolysis,
156 economic assessment, environmental assessment, life cycle analysis, LCA”. Based on

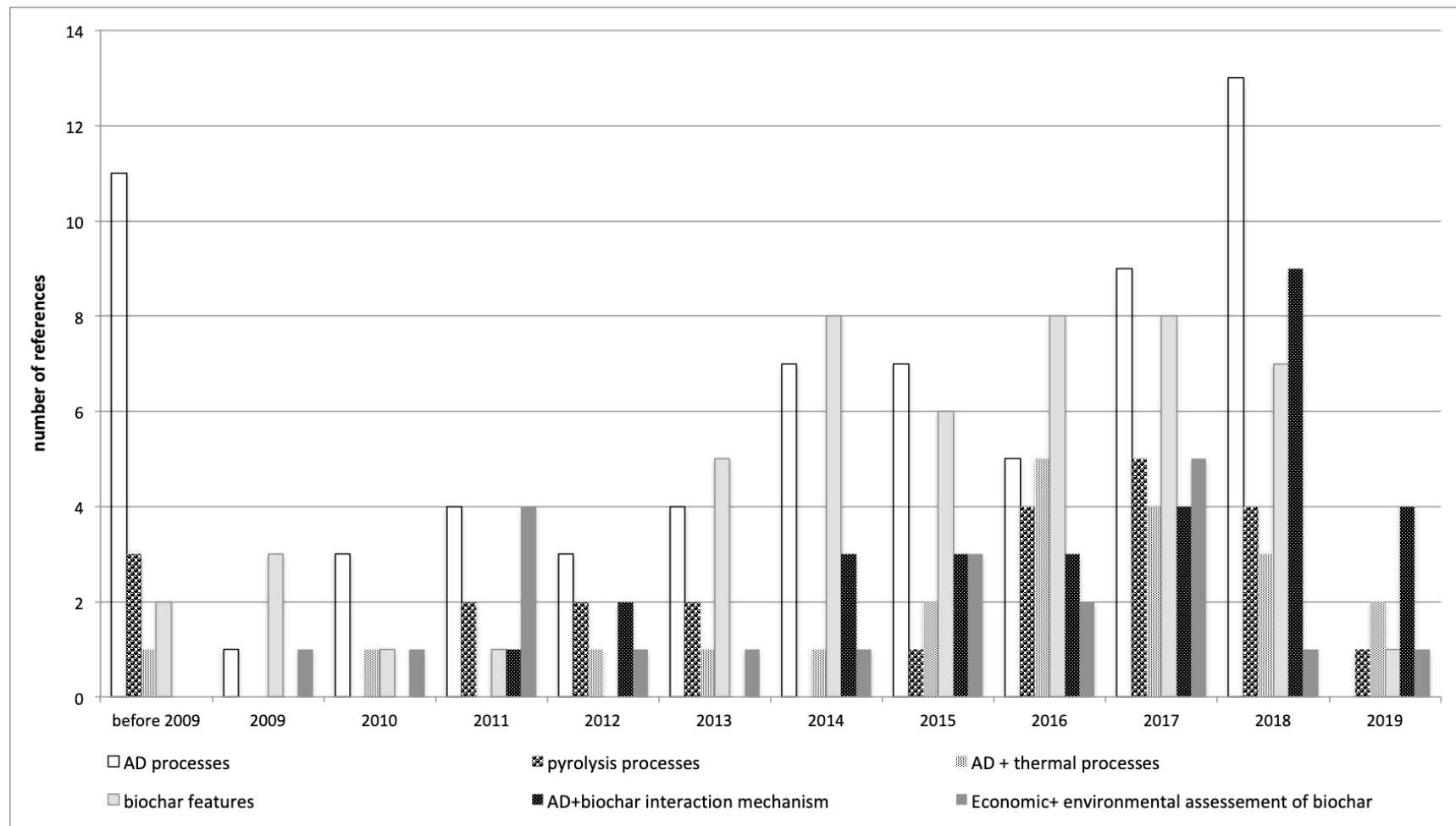
157 this preliminary selection, the review questions have been identified. Thereby, additional
158 keywords have been adopted to face specific issues, as: “biochar, buffer capacity,
159 anaerobic digestion, volatile fatty acids”, “biochar, ammonia inhibition, ammonia
160 removal, anaerobic digestion”, “syntrophic metabolisms, biomass immobilization,
161 biochar, anaerobic digestion, DIET, electron transfer”.

162 d) *Selection of the references based on content analysis.* After a first screening
163 considering the abstracts of all identified reference sources, 259 relevant references
164 were analyzed as full content and included in this review. The relevant scientific
165 content was sourced from scientific articles (240 research papers and reviews – 93 %),
166 book chapters and indexed conference proceedings (11 documents – 4 %), and
167 international databases and reports (8 documents – 3 %). Considering only the
168 scientific research papers and the reviews, they were related to (Figure 1): AD (31 %)
169 and pyrolysis (10 %); coupling AD and thermal processes (10 %); BC features (25 %);
170 BC and AD interaction mechanisms (14 %); economic and environmental aspects
171 related to BC use (10 %). The distribution of the relative contributes of the scientific
172 literature to the different topics concerning the investigation of BC role in AD in 2009
173 – 2019 exhibits a clear increasing trend due to a growing interest, particularly in the
174 last five years, for the topics covered by the present work and consequently strengthens
175 the need to address the above mentioned review questions.

176 This review is structured according to the contents outline. At the end of each section, the
177 last paragraph summarizes the main findings and research gaps of the specific topic
178 explored, which are finally condensed in the conclusions.

179

180 Figure 1. Relative contributions of the scientific literature selected for this work



183 **3. Biochar**

184 BC is the porous carbonaceous solid residue of the complex physical and chemical
185 phenomena simultaneously occurring during the thermochemical treatment of raw
186 biomass [60–63]. While AD is particularly suitable to produce biogas and digestate from
187 wet organic waste (Figure 2), pyrolysis is a thermochemical process able to convert solid
188 dry biomass in an oxygen-limited environment into valuable liquid chemicals (bio-oil),
189 as well as gaseous biofuels and charcoal (biochar, BC) [64–67]. Although pyrolysis was
190 conventionally mostly focusing on the production of liquid biofuels from woods and
191 purpose-grown energy crops, the most recent studies found that it is not cost effective
192 unless the feedstock is inexpensive, dry and readily available, and there are valuable
193 chemical co-products (liquid bio-oil and solid BC) or favorable government policies [68].
194 Both AD and pyrolysis can contribute to fulfill Circular Economy targets, but research
195 on biological and thermochemical processes traditionally progressed along parallel and
196 separate pathways, often in competition.

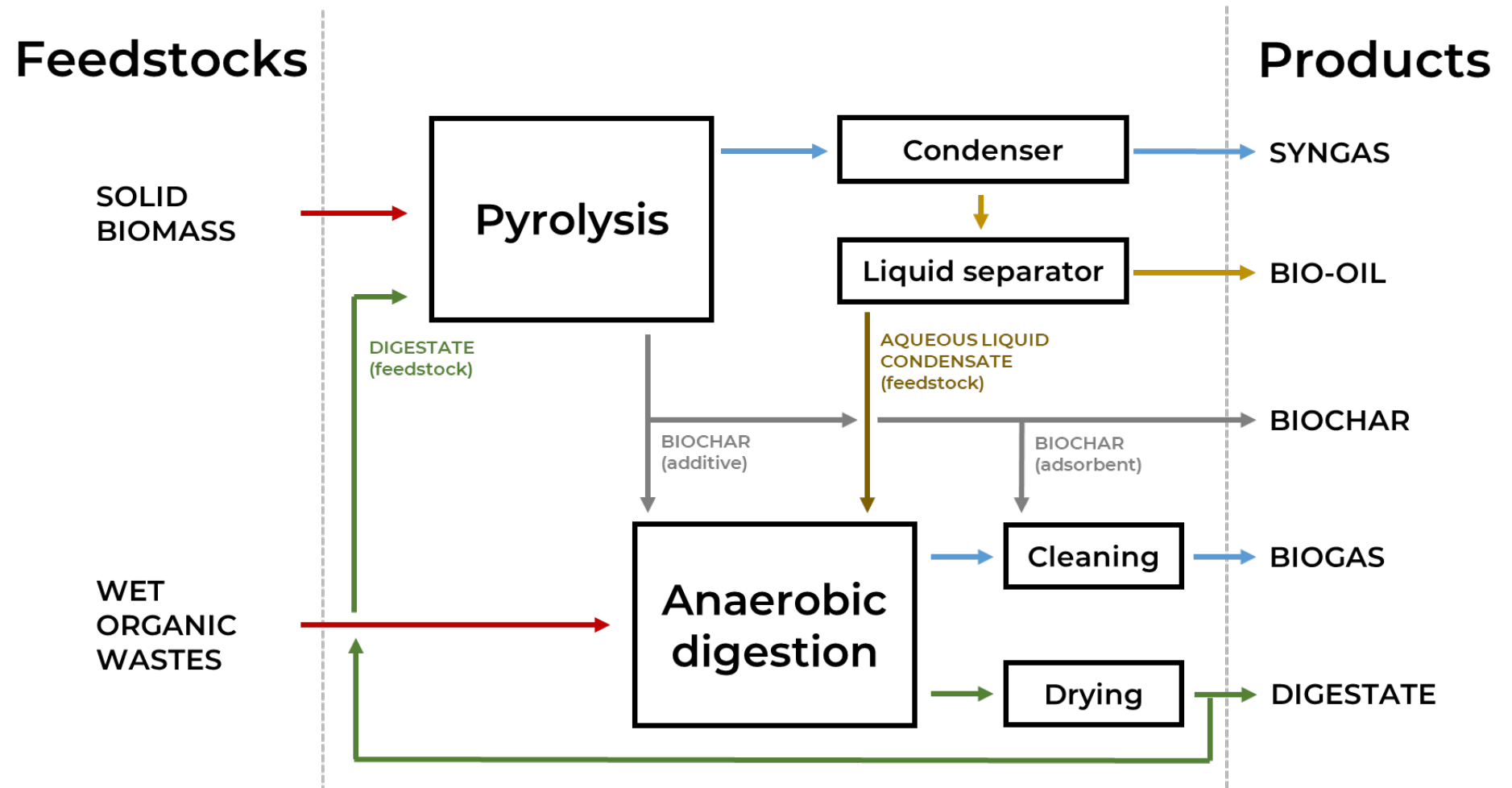
197 In recent years, improved understanding of the characteristics of bio-oil and BC has
198 helped to develop key quality requirements to expand their potential value [69]. High
199 value liquid bio-oil products can be obtained from the pyrolysis of biomass residues, such
200 as phenolic substitutes for adhesives or antioxidants [70,71]. However, there are
201 limitations: the most valuable chemicals are usually mixed with impurities that impair
202 their properties. Similarly, there are many promising applications for BC, such as soil
203 improver [18], composite materials [72], activated carbon [73], electrodes for batteries or
204 electrolysis cells [74], metallurgical coke substitute [75], catalyst for tars [76], and could
205 be eligible for carbon credits [77]. The efficiency of BC in most applications significantly

206 depends on its physico-chemical properties and elemental composition, which are subject
207 to variations depending on feedstock, pyrolysis conditions, pre- and post-processing
208 treatments.

209 The characteristics of AD and pyrolysis processes and products may offer interesting
210 opportunities for the integration of such technologies (Figure 2), with the objective of
211 contributing to the Circular Economy. For example, pyrolysis could be utilized to convert
212 dewatered digestate into BC, which could then be used for a variety of applications
213 [17,78,79], including its potential use to improve the quality of biogas and digestate [37],
214 or to reduce instabilities and inhibition in digestors [16,24]. On the other hand, the acetic
215 acid-rich aqueous pyrolysis condensate could be utilized as a feedstock for AD, possibly
216 with the addition of BC for the selective adsorption of toxic inhibitors [21–23,25,80–88].
217 In conclusion, BC represents the key link between pyrolysis and AD and, consequently,
218 in this section we wish to review its characteristics, including suitable feedstocks,
219 production technologies, activation processes and characterization techniques.

220

221 Figure 2. Outline of potential integrations between anaerobic digestion and pyrolysis (derived from [20,21]).



222

223

224 3.1. Feedstocks

225 During thermal processing under oxygen free or oxygen-limited conditions (as pyrolysis),
226 as the temperature increases, the polymeric components of lignocellulosic biomass crack
227 and liquid intermediate phases are formed with different visco-elastic behaviors [89,90];
228 under slow progressive heating, cellulose remains hard and elastic up to 400 °C, whereas
229 hemicellulose softens within a relatively narrow temperature range (230 to 280 °C) and
230 remains viscous, while lignin softens and bubbles over a broader temperature range (150
231 to 350 °C) and remains mainly elastic. The overall process cannot be simply designed
232 based on the linear combination of the behaviors of the individual components, but it is
233 the result of their interactions at different temperatures and it is catalyzed by the mineral
234 matter present in the ashes. As the temperature increases over these ranges, the
235 depolymerized liquid undergoes two competitive processes, oligomer evaporation and
236 crosslinking reactions, with the former favored at high heating rates, and the latter under
237 slow pyrolysis. These findings explain why, under slow pyrolysis, the char yield is
238 greater, and the particles globally keep the same macro-structure of the original biomass,
239 although forming an intermediate soft material. Montoya et al. (2017) [91] confirmed the
240 presence of molten liquid intermediates formed from cellulose, xylan and lignin and
241 showed that the presence of minerals prevents the complete liquefaction of the xylan,
242 therefore contributing to the preservation of the general shape and structure of the solid
243 material during pyrolysis.

244 Therefore, BC can be produced from a wide range of lignocellulosic materials, containing
245 cellulose, hemicellulose (xylan), lignin, small quantities of other organic compounds
246 (phenols, phytosterols and fats) and inorganics (P, N, S, Si, alkali and alkaline earth
247 metals and traces of other minerals). Such materials include wood, agricultural and forest

248 residues, food waste, sugars, industrial organics, sewage sludge and manure. BC can be
249 mesoporous or microporous depending on the operating conditions employed for its
250 production and on the feedstock.

251 3.2. *Thermochemical production*

252 Recent reviews illustrated the different technologies and processes used experimentally
253 and industrially to produce BC [17,92]. These include torrefaction, pyrolysis,
254 gasification, combustion and hydrothermal carbonization [93]. Scale and potential
255 mobility are important considerations in relation to the feedstock supply, logistics,
256 seasonality, further refining, of products quantities, characteristics and value, and
257 potential markets [17,60].

258 *Torrefaction* involves biomass treatment at atmospheric pressure and within a
259 temperature range of 200–300 °C, without oxygen or with limited oxygen supplies [94].
260 As discussed earlier, partial depolymerization and liquefaction of the biomass
261 components take place reducing the biomass tenacity and improving grindability.
262 Torrefaction produces char with less moisture, higher energy density, lower weight, lower
263 O/C and H/C ratio, increased hydrophobic nature and resistance to biological degradation
264 with respect to the original feedstock. Typical yields of torrefied biomass range between
265 50 and 80 % [95].

266 *Gasification* is a thermochemical process carried out at temperatures higher than 750 °C
267 in the presence of a gasifying agent (air, oxygen, or steam) at atmospheric or at high
268 pressures. Under these conditions, biochar yields are not sufficiently significant to
269 consider gasification an appropriate BC production process. Similarly, combustion is not
270 a suitable BC production process, since, under good combustion conditions, BC yield
271 should be negligible.

272 The various modes of *pyrolysis* include slow, intermediate, fast, flash, and ultra-pyrolysis,
273 carried out under vacuum, atmospheric pressure or under pressure [65,69]. As a result of
274 the balance between primary, secondary cracking and recombination reactions, bio-oil
275 yields are typically maximized (up to 70 %) at intermediate temperatures (450-550 °C),
276 faster heating rates (100-500 °C/s) and short vapor residence times (< 1~2 s), which are
277 characteristic of fast and flash pyrolysis. Under these conditions, BC yields are typically
278 of the order of 15~20 %. On the other hand, higher BC yields (25~40 %) are achieved at
279 moderate temperatures (300-450 °C), slower heating rates (~ 1 °C/s) and longer vapor
280 residence times (> 5~10 s), representative of slow or intermediate pyrolysis, when bio-oil
281 yields vary between 40 and 50 % with the balance being gas.

282 *Hydrothermal carbonization (HTC)* is performed on wet biomass (moisture > 10 %) as
283 feedstock, making the process highly energy-intensive [96]. HTC happens under water in
284 a sealed confined system and heated at the temperature range of 175–300 °C for up to 16
285 h under saturated pressure under subcritical conditions producing tar-free BC (hydrochar)
286 with large number of functional groups [97–99]. Hydrochars are usually obtained at
287 yields of 20-25 % [96], and they contain mainly aliphatic compounds and more oxygen
288 functional groups and higher cation exchange capacity than conventional BC. On the
289 other hand, they have lower surface area, microporosity and carbon stability [96,100].

290 Among the various thermal technologies, pyrolysis has been the most investigated
291 technique and it is considered the best technology for the production of BC [101] from
292 relatively dry feedstock (moisture content below 10 %). However, also HTC is gaining
293 interest to produce hydrochar, particularly from wet biomass [96]. In a comparison
294 between the chars, it should be noticed that BC yields are higher, as well as porous
295 structure and aromatics and ash contents, while hydrochar exhibits a non-porous

296 structure, lower fixed and total carbon contents and higher amounts of alkyl functional
297 groups [96].

298 3.3. *Activation*

299 BC with different properties can be produced from a wide range of feedstock and by
300 adjusting processing conditions. Further modification of BC chemical and physical
301 properties can be achieved through a process defined “activation”, aimed at increasing
302 BC porosity and modifying its pore size distribution (Figure 3), as well as to some extent
303 surface chemistry. Activation can be carried out in a number of ways, depending on type
304 of activation agents (e.g. physical and chemical activation) or mode of operation into
305 single-stage or multi-stage activation.

306 *Chemical activation* utilizes chemical agents, e.g., H_3PO_4 , HNO_3 , KOH , NaOH , H_2SO_4 ,
307 and ZnCl_2 [102,103]. It typically involves two steps; in the first the feedstock is
308 impregnated with a selected chemical agent, and then thermally treated in the second step.
309 Another option is the activation of already produced BC by soaking it in a chemical agent,
310 followed by a thermal treatment. Depending on the agent selected and thermal treatment
311 conditions used, different degrees of activation can be achieved. The activated BC needs
312 to be thoroughly washed with deionized water to neutralize its pH and to remove any
313 remaining chemicals [104], and this procedure can contribute to a negative environmental
314 impact of the technology. Oxidative activation that uses acidic or alkaline agents is among
315 the most common activation methods. Besides enhancing porosity and surface area, it
316 also creates oxygen-containing functional groups on the surface of BC (e.g. carboxyl,
317 hydroxyl, lactone, phenol, carbonyl, and peroxide groups). These functional groups play
318 an essential role in different applications of BC, such as for contaminant removal or
319 nutrient sorption.

320 *Physical activation* does not utilize chemicals, thus avoiding some of the negative
321 environmental impacts that can be associated with chemical activation. Instead, BC is
322 exposed to physical oxidizing agents, such as steam, CO₂, ozone or air/O₂, typically at
323 temperatures above 700 °C. These gaseous agents penetrate the internal structure of BC
324 gasifying the carbon atoms, which results in opening and widening of pores [105].
325 Similarly to chemical activation, physical activation not only enhances BC porosity, but
326 also modifies its surface functional groups, increasing the abundance of oxygen
327 functional groups [105].

328 In conclusion, both chemical and physical agents can be used effectively to activate BC
329 and obtain high porosity, surface areas over 1000 m²/g, and modified surface functional
330 groups, tuned to specific applications. The key challenges of BC activation are without
331 any doubt related to their potential negative environmental impacts, mainly for chemical
332 activations, and high-energy intensity need.

333 3.4. *Physico-chemical characteristics*

334 The variability of BC physico-chemical properties resulting from the proper selection of
335 feedstock, pyrolysis and activation can be adapted to a wide range of applications.

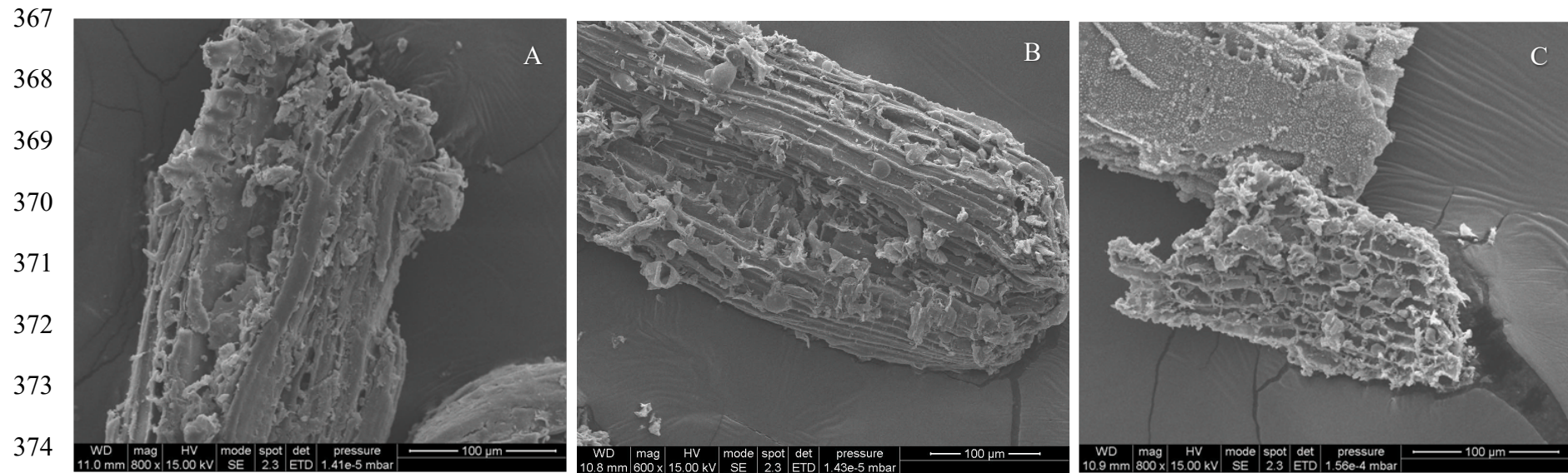
336 *Specific surface area* (SSA), expressed as m²g⁻¹, is the ratio between the total surface area
337 and the total particle mass of BC [106], commonly determined by Brunauer–Emmett–
338 Teller (BET) analysis. The reported SSA values of BC can vary significantly, in the range
339 10⁰ - 10² m²g⁻¹ [107,108] depending on pyrolysis temperature and feedstock, and up to
340 over 10³ m²g⁻¹ after activation. SSA is a key parameter enhancing the adsorption capacity
341 of BC, even if the chemical composition of its surface is determinant in the interactions
342 with other species [109]. SSA is related to BC pore size distribution [18].

343 *Porosity* can be described by the pore volume (m^3g^{-1}) and by the pore size distribution,
344 including micro-, meso-, and macro-pores (respectively $< 2\text{nm}$, $2\text{-}50\text{ nm}$, $>50\text{ nm}$) [110],
345 based on their internal diameters. The relative abundance of these pores produces
346 different BC structures (Figure 3), since micro-pores have the major contribute to BC
347 surface area, micro- and meso-pores are essential for its adsorption capacity, while macro-
348 pores can provide an appropriate habitat for microorganisms [18,111]. Yin et al. (2017)
349 [112] stated that within a proper range, a high temperature results in a large surface area
350 and pore formations. Trigo et al. (2016) [113] found that SSA increased between 350°C
351 and 700°C for different hardwood biochars, while Chen et al. (2014) [114] revealed an
352 enhanced porous structure of BC from sewer sludge varying the temperature from 500°C
353 to 900°C .

354 Specifically considering the effect of the feedstock on BC characteristics [101], yields
355 and aromatic functional groups are much higher from lignin (48.8 %) than from
356 hemicellulose (21.1 %), while no differences were found in the elemental composition of
357 resulting BCs. High lignin and mineral content in the feedstock result in higher yields of
358 BC [115,116]. Woody and grassy biomasses generate a more carbon-rich BC when
359 compared to sewage sludge and manure [32,117,118]. Specifically considering the effect
360 of pyrolysis process on BC characteristics [119], increasing pyrolysis temperature
361 decreased concentrations of aliphatic carbons, oxygenated functional groups, nitrogen,
362 oxygen and hydrogen. However, carbon mass fraction, micropore volume, ash content,
363 fixed carbon and carbon stability increase with higher pyrolysis temperatures.

364

365 Figure 3. SEM micrographs of (A) raw biomass (eucalyptus) and of derived (B) biochar and (C) physically activated biochar (Courtesy of
366 the authors. The samples were produced at ICFAR and analysed at Politecnico di Torino).



375

376

377 BC *composition* is mainly of C, H, O, N, and S, along with mineral trace components,
378 such as K, Ca and Mg, varying according to pyrolysis conditions and feedstock [110].
379 The mineral content can be enhanced by increasing the temperature of pyrolysis [107].
380 The *cationic exchange capacity* (CEC) of BC can be defined as its ability of absorbing
381 and attracting cations (nutrients and heavy metals) decreasing with higher pyrolysis
382 temperature [18]. A high CEC value is due to a negative surface charge of BC. The
383 presence of *surface functional groups* such as hydroxyl, carboxylic, and amino, expressed
384 respectively as atomic ratio of H/C, O/C, and N/C (generally studied by Fourier transform
385 infrared (FTIR) spectroscopy), mostly depends on the feedstock and it is reduced with a
386 higher temperature of pyrolysis [107]. Further, the H/C ratio is considered as a measure
387 of the degree of aromatization of BC and the O/C ratio as an indication of more
388 oxygenated functional groups, contributing to high CEC values and hydrophilicity of BC
389 [120,121]. An increase in pyrolysis temperature has been reported to negatively affect the
390 amount of acidic functional groups and the CEC [112,122].

391 *pH values* are generally basic and increase as the pyrolysis temperature grows, due to the
392 volatilization of acidic functional groups and to the higher ash content [18]. Moreover,
393 BC can present redox characteristics, being able to donate and accept electrons, and these
394 tendencies are quantified respectively by the *electron donating* and *accepting capacities*
395 [110]. Phenolic groups may be the electron donating species, while quinones and
396 polycondensed aromatic structures the electron accepting compounds [110,123]. In
397 addition, the *electrical conductivity*, expressed as S m^{-1} , decrease as consequence of the
398 increase of the pyrolysis temperature [17].

399 A full characterization of BC involves several analytical methods [124]. Scanning
400 electron microscopy (SEM) is employed to investigate the superficial structure of BC, as

401 well as its chemical composition if coupled with energy dispersive X-ray spectroscopy
402 (SEM-EDX); transmission electron microscopy (TEM) allows to observe BC bulk
403 structure; X-ray diffraction (XRD) analysis detects the crystalline phases; nuclear
404 magnetic resonance (NMR) spectroscopy investigates BC composition, specifically the
405 aromatic structure; X-ray photoelectron spectroscopy (XPS) quantitatively analyses the
406 superficial composition.

407 **4. Application of biochar as additive in anaerobic digestion**

408 *4.1. Buffer capacity and alkalinity*

409 The volatile fatty acids (VFAs) produced as intermediates during AD tend to lower the
410 pH [125]. This effect is generally counteracted by syntrophic acetogens and methanogens
411 microorganisms, which convert VFAs to methane and carbon dioxide [126]. However, in
412 case of high organic loads of easily biodegradable wastes (i.e. when the VFAs production
413 rate exceeds the consumption rate), the accumulation of VFAs can occur, resulting in pH
414 drop and even in the failure of AD [126,127]. The buffer capacity in a digester is linked
415 to the ability to neutralize VFAs, and it is considered a rate-limiting step in AD processes
416 [38,39,128]. The buffer capacity is determined by the alkalinity of the AD system, mainly
417 in the form of carbon dioxide and bicarbonate [6,125]. So it is technologically challenging
418 to build a bioreactor aimed at digesting easily biodegradable feedstock with high pH
419 buffer capacity [16]. Even if several methods have been proposed (Table 2) so far, the
420 most diffused solution to improve buffer capacity implemented both at lab [129] and full
421 scale [130] is co-digestion. In particular, manure or sewage sludge showed high buffer
422 capacity, mainly due to the quite high ammonia content, which is able to counteract the

pH drop due to VFAs production. Therefore, manure or sewage sludge are often co-digested with OFMSW and/or agricultural waste biomass.

425

Table 2. Advantages and disadvantages of conventional methods for creating buffering capacity in bioreactors

| Methods | Advantages | Disadvantages |
|--|---|---|
| Alkaline pretreatment [131] | Cellulose more available to the enzymes | Expensive and considered as the technological bottleneck |
| Commercial alkaline materials (e.g. NaOH and CaCO ₃) [132,133] | Easy pH adjustment | pH continually should be analyzed Temporary Lower biogas output at higher organic loads |
| Co-digestion [16] | Increasing methane yield Low cost and green | Hard to achieve an appropriate mixture of feedstocks to limit VFAs production |
| Geopolymer (Composite and monolith) [132,133] | Controllable alkali leaching Prevent the need for continuous pH adjustment | 25 g L ⁻¹ costs around 0.10 \$ |

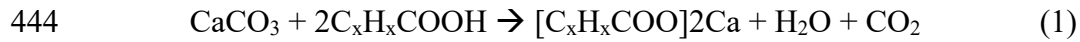
428

However, it still remains a great challenge to develop simple, permanent and cost-effective methods to improve buffer capacity of AD systems. Nowadays, BC is considered as an attractive alternative to the aforementioned methods for two important reasons. Firstly, it can be produced via cost-effective and environmentally friendly approaches. Secondly, its physical and chemical properties can be matched with the operational conditions [19,40,41].

BC's buffer capacity mainly depends on two factors:

I. *Functional groups*: Rapid accumulation of VFAs during AD process results in a medium with low pH value in which some functional groups of BC like amine adsorbs H⁺ and accepts electron. This phenomenon could mitigate the sudden pH drop.

440 II. *Inorganic materials*: Ash portion of biochar contains inorganic materials such as Ca,
441 K, Mg, Na, Al, Fe, Si and S. Among them, alkali and alkaline earth metals (AAEMs)
442 are responsible for alkalinity of biochar via reaction 1 (Ca and C_xH_xCOOH are
443 selected as representative of AAEMs and VFAs, respectively) [16]:



445 Table 3 shows the literature data related to BC buffering capacity and its effects on bio-
446 methane production, showing total alkalinity, pH values and the increase in CH₄
447 production (%), ranging from 8 % up to about 26 %. Some studies suggest that the
448 alkalinity of BC due to AAEMs in ash fraction could effectively contribute to the
449 buffering capacity of AD against VFAs inhibition. Jang et al. (2018) [27] investigated the
450 effects of BC from dairy manure on AD of dry dairy manure at 20°C, 35°C, and 55°C. In
451 each case they observed a lower total VFAs concentration and increased methane
452 production. They suggested the role of high nutrients (9.1% Ca, 3.6% Mg, 1.3% N, 0.14%
453 P) and alkalinity potential of BC in enhancing methane production. Wang et al. (2017)
454 [39] investigated the role of BC from vermicompost on the buffering capacity of AD of
455 high organic loads of kitchen waste and chicken manure. The authors proved the high
456 buffer capacity of vermicompost BC to different short-chain VFAs (700-3800 mg L⁻¹),
457 probably due to the presence of AAEMs and superficial functional groups. Linville et al.
458 (2017) [24] studied the effects of BC from walnut shell during the AD of FW in
459 mesophilic and thermophilic conditions. They observed that BC improved process
460 stability by enhancing the total alkalinity from 2800 to 4800-6800 mg L⁻¹ CaCO₃ and pH
461 (initial pH >8) in BC amended digesters. Wei et al. (2020) [134] found enhanced methane
462 production and solids removal by adding BC from corn stover (rich of alkaline earth
463 metals) to AD of primary sludge. The authors observed higher total alkalinity (3500-4700

464 mg L⁻¹ CaCO₃) and pH in BC amended reactors, suggesting that BC provided a strong
465 buffering capacity. Ambaye et al. (2020) [135] investigated the role of BC from sewage
466 sludge on the AD of fruit waste. They observed that the BC addition enhanced methane
467 production and VFAs degradation. Further, the deficiency of some nutrients or trace
468 elements in the substrate may cause an increase of VFAs inducing inhibition of microbial
469 activity during AD [33,135]. Extra doses of trace metals may be required for an effective
470 activity of methanogens without VFAs accumulation [136]. Thereby, it could be
471 speculated that BC could provide adequate concentrations of trace metals for the stability
472 of the AD system [135].

473 In summary, BC addition can counteract VFAs inhibition in case of high loads of easy
474 degradable wastes such as FW, OFMSW, primary sludge. The alkaline nature of BC
475 determining its pH buffering capacity may contribute to prevent VFAs inhibition. The
476 ash fraction of BC contains AAEMs possibly contributing to its acid-buffering capacity
477 and important trace elements for microorganisms. However, further investigations are
478 required to confirm the effectiveness of these mechanisms and to identify the optimal
479 dose of BC related to these properties. Higher doses can be toxic for AD [24,134], perhaps
480 due to inhibitory concentrations of alkali metals [137]. Besides, other mechanisms could
481 be responsible of BC role in alleviating VFAs inhibition. Porous BC could provide
482 support for biofilm growth and protection to selectively enriched functional microbes
483 closely attached to it under acid stress [48]. Secondly, BC could promote the activity of
484 microbial partners enhancing the syntrophic VFAs degradation and methane production
485 under high organic loads [38,138]. The latter microbial mechanisms will be fully addressed
486 in section 4.3.

487

488 Table 3. Results of pH drop and CH₄ production/yield of AD processes in the presence
489 of various types of BCs

| Pyrolysis Operational condition | | | | AD Operational conditions | | | | Results of buffering effect | | | | Ref |
|---------------------------------|------------|----------------|-------------------------|---------------------------|---------------------|-----------------------|------------------|---|---------|---------|---------------------------------|-----------------------------|
| Temp (C°) | Time (min) | Reactor | Feedstock | Temp (C°) | HRT or duration (d) | Substrate | inoculum | Total alkalinity (g L ⁻¹ CaCO ₃) | pH | | Increase in CH ₄ (%) | |
| | | | | | | | | Control | BC | Control | BC | |
| 350 | 180 | Muffle furnace | dairy manure | 20 | 35 | dairy manure | Anaerobic sludge | 1.8-4.8 | 5.3-7.3 | 7.18 | 7.71 | 26.47 ^a [27] |
| 350 | 180 | Muffle furnace | dairy manure | 35 | 35 | dairy manure | Anaerobic sludge | 1.8-4.8 | 5.3-7.3 | 7.09 | 7.68 | 24.90 ^a [27] |
| 350 | 180 | Muffle furnace | dairy manure | 55 | 35 | dairy manure | Anaerobic sludge | 1.8-4.8 | 5.3-7.3 | 7.12 | 7.55 | 24.69 ^a [27] |
| 500 | 120 | Tube furnace | vermi-compost | 35 | 50 | chicken manure | Anaerobic sludge | 6.0 | 7.3 | 7.0-7.1 | 7.8-8.0 | N.P. [39] |
| 500 | 120 | Tube furnace | vermi-compost | 35 | 50 | kitchen waste | Anaerobic sludge | 0.0 | 2.3 | 3.5-3.7 | 4.7-4.9 | N.P. [39] |
| 600 | 120 | Kiln | corn stover | 55 | 15 | primary sludge | Anaerobic sludge | 2.8 | 3.5-4.7 | 7.2 | 7.6-8.0 | 8.6-17.8 ^a [134] |
| 350 | 15 | | sewage sludge digestate | 37 | | fruit waste | Anerobic sludge | N.P. | N.P. | 4.5-5 | 7-8 | 13-27 ^a [135] |
| 550 | 15 | | sewage sludge digestate | 37 | | fruit waste | Anerobic sludge | N.P. | N.P. | 4.5-5.5 | 7-8.6 | 12-22 ^a [135] |
| 500 | 120 | Slow pyrolysis | rice straw | 35 | 25 | Municipal solid waste | Anaerobic sludge | | | 7.8 | 7.5 | 11.69 ^a [139] |

a) CH₄ Yield; N.P: Not provided.

490

491

492 4.2. Adsorption of inhibitors

493 Inhibition is often defined as the predominant cause of reduction of bio-methane yields
494 and instability of AD processes. A substance can be identified as “inhibitor” when it
495 determines an adverse shift in the microbial population or arrests of the bacterial growth
496 [137]. There are direct inhibitors, such as metals (Cu²⁺, Zn²⁺, Cr³⁺, Cd, Ni, Pb⁴⁺, Hg²⁺,
497 Na⁺, K⁺, Mg²⁺, Ca²⁺, Al³⁺), organic compounds (chlorophenols, halogenated aliphatics,
498 pesticides, antibiotics, lignocellulose hydrolysate); and indirect inhibitors, such as VFAs,
499 long-chain fatty acids, hydrogen, ammonium and sulphides [19]. Among the inhibitors,
500 ammonia (NH₃ or free ammonia nitrogen, FAN) and ammonium (NH₄⁺) concentrations
501 are considered crucial (ammonia and ammonium are together considered as total
502 ammonia nitrogen, TAN). The optimal control of ammonia and ammonium may ensure
503 stable bacterial growth and significantly contribute to the buffer capacity of the AD

504 system. However, excess FAN/TAN concentrations are major causes of AD failure
505 [140,141]. Chen et al. (2008) [137] reported a wide range of TAN concentrations
506 (between 1.7 and 14 g L⁻¹) causing a 50 % reduction in methane production. FAN
507 concentration is considered the main cause of inhibitions for methanogens [140,142] and
508 it was reported to have a toxic effect to anaerobes ranging from 150 to 1200 mg L⁻¹
509 [143,144]. Various mitigation strategies have been investigated to effectively counteract
510 TAN/FAN inhibitory effects on AD. Some were based on the removal and
511 immobilization of the inhibitor, as: dilution, co-digestion and microbial adaptation [145–
512 147]; struvite precipitation [148]; use of a microbial desalinisation cell [149]; ammonia
513 stripping [150]. An alternative approach is based on the use of adsorbents, inorganic as
514 zeolites [142] and organic as GAC [143] and BC [41].

515 Based on several studies focusing on BC impacts on AD and ammonia inhibition (Table
516 4), it seems that BC could effectively mitigate ammonia inhibition, resulting in reduced
517 lag phase and enhanced methane production respect to control reactors. Some authors
518 [33,34,151] promoted BC use in AD of sewer sludge. Mumme et al. (2014) [41] revealed
519 that BC from pyrolysis of paper sludge and wheat husks could limit mild ammonia
520 inhibition (2.1 g TAN kg⁻¹). Su et al. (2019) [43] stated that the addition of BC may
521 alleviate inhibition in case of up to 1500 mg L⁻¹ ammonia-N in food waste AD [152]. Lü
522 et al. (2016) [44] revealed that BC can support AD under high ammonium stress (up to 7
523 g-N L⁻¹). The above-mentioned studies suggest a positive effect of BC towards ammonia
524 inhibition, however, there isn't full agreement on the mitigation mechanisms
525 hypothesized: cation exchange capacity [34,151]; chemical and/or physical adsorption
526 capacity and surface functional groups [33,34,151]; promotion of direct interspecies
527 electron transfer (DIET) [44,153]; immobilization of microorganisms [43,44].

528 Therefore, BC may contribute to ammonia mitigation by direct (cation exchange capacity,
529 adsorption, surface functionality) and/or indirect factors (DIET and immobilization of
530 microorganisms), depending on the characteristics of both BC and digested substrate,
531 along with the operating conditions of the AD process (e.g. pH and temperature).
532 Focusing on direct factors, a better understanding of the interactions between BC and
533 FAN/TAN is a key step to identify its physico-chemical properties able to maximise the
534 ammonia removal. The mechanisms for ammonia adsorption on BC from wastewater and
535 digestate have been studied in literature (Table 5), resulting in promising values of
536 ammonium adsorption capacity (up to hundreds of $\text{mg NH}_4\text{-N g}^{-1}$ BC). Yin et al. (2017)
537 [112] reported that physical sorption could be supported by high SSA and large porous
538 structure. However, various studies suggested that porosity and SSA may not be the
539 predominant factors in ammonium adsorption [154,155]. For instance, ion exchange may
540 occur between acidic functional groups on BC surface and ammonium [112,156,157],
541 and CEC could have a major role in enhancing the ammonium adsorption capacity of BC
542 [157]. Zhang et al. (2014) [158] found that BC derived from pyrolysis of corn cob at 400
543 °C exhibited higher ammonium adsorption capacity than those produced at 600 °C due to
544 the presence of functional groups with higher acidity. Thus, the definition of the proper
545 pyrolysis temperature and the other control parameters is crucial for enhancing the
546 adsorption capacity of BC.

547 In conclusion, various studies reported the potential of BC in alleviating ammonia
548 inhibition (Tables 4 and 5) through different mechanisms. The direct adsorption of
549 ammonia on BC may contribute to the mitigation of ammonia inhibition. However,
550 further studies are required to clarify the effective role of direct mechanisms in complex
551 AD systems in presence of other phenomena, i.e. the close bond between ammonia and

552 VFAs inhibition. In some cases the direct removal of ammonium may not be the main
553 contribution to the mitigation of ammonia inhibition by BC [44,82,153]. Instead, other
554 indirect mechanisms (immobilization and faster acclimation of biomass, DIET) have been
555 suggested in literature and will be discussed in the next sections.

556

557 Table 4. A summary of selected works focusing on ammonia/ammonium inhibition mitigation by BC in AD processes (FAN: free ammonia
558 nitrogen; TAN: total ammonia nitrogen).

| Biochar | | | | Anaerobic digestion | | | | | | | | Reference |
|---|---------------------------|--|--|--|--|------------------|--------------------------------------|--|--|---|--|-----------|
| Feedstock | Process: temperature [°C] | Dose of biochar | Substrate | Inoculum | Inoculum to substrate ratio (VS basis) | Temperature [°C] | pH | Critical TAN conc. or as specified [mg L ⁻¹] | Critical FAN conc. or as specified [mg L ⁻¹] | Ammonia removal by biochar | Results and comments | |
| Fruitwood | Pyrolysis: 800-900°C | 10 g L ⁻¹ | Glucose solution (6 g L ⁻¹) | Granular sludge from AD of paper mill wastewater | 1:6 | 35 | 7 | 260 3500 7000 | - - - | | <ul style="list-style-type: none"> • ↑ Specified CH₄ prod. rates increased by 18.6%, 10.1% and 23.5% respectively • ↓ Lag phase by 30.4%, 12.7% and 23.8% respectively | [44] |
| 1:2 (v/v) mix: - paper sludge - wheat husks | Pyrolysis: 500°C | 6.67% w/w (2 g BC per 30 g inoculum) | - | Meso. AD (TS = 4.05%) + Ammonium carbonate | Only 30 g of inoculum | 42 | 7.66 7.73 7.93 7.84 8.07 | 1626 2126 3126 4126 6626 | - - - - - | 2.4 mg TAN g ⁻¹ BC 2.0 mg TAN g ⁻¹ BC 4.2 mg TAN g ⁻¹ BC 4.5 mg TAN g ⁻¹ BC 6.8 mg TAN g ⁻¹ BC | <ul style="list-style-type: none"> • No clear effect on biogas production was observed | [41] |
| Macadamia nut shells | Pyrolysis: 350°C | 33.3 g L ⁻¹ | Food waste (+ water + NaHCO ₃) | AD from UASB (SS = 34 g | - | Room temperature | | 1500 | - | | <ul style="list-style-type: none"> • ↑ COD removal from 78% without BC to 90% with BC | [43] |
| Corn stover | Gasification | 1.82, 2.55, 3.06, 3.64 g BC g ⁻¹ TS sludge | WWTP sludge | Thermo WWTP AD | 2:1 | 55 | 7.5 | 1100 (total N) | 600 (NH ₃ -N) | | <ul style="list-style-type: none"> • CH₄ yield, biomethanation rate constant and ↑ max CH₄ prod. rate up to 7.0%, 8.1% and 27.6% respectively • ↑ NH₃-N concentration by 41.5% after AD without BC, while by 0.2-18.1% with BC | [151] |
| Pine pellets White oak pellets | Gasification | 2.49, 4.97 g BC g ⁻¹ TS sludge 2.20, 4.40 g BC g ⁻¹ TS sludge | WWTP primary sludge (inoc +subs) | Meso. and thermo. WWTP AD (inoc +subs) | 2:1 | 37, 55 | 7.2-7.3 | 750 (total N) | 400-450 (NH ₃ -N) | | <ul style="list-style-type: none"> • ↑ CH₄ yield by 3.9-9.5% • ↑ NH₃-N concentration by 67.0% after AD without BC, while by -7.2 to 4.7% with BC • ↑ CH₄ yield by 5.7-9.6% • ↑ NH₃-N concentration by 67.0% after AD without BC, while by -7.2 to 4.7% with BC | [33] |

| | | | | | | | | | | | | |
|---|--|--|-----------------------------------|---|-----------------------------------|----|-----------------------------------|---------------------|------------------------------|------------------------|---|------|
| Corn stover Pine | Gasification: 850°C | 0.25, 0.375, 0.5, 1.0 g d ⁻¹ | Primary sludge + WAS (TS = 7.01%) | WWTP AD from TPAD: 1 stage (TS = 6.14%) 2 stage (TS = 3.77%) | 2:1 | 55 | 7.4 (stage I); 5.3-6.0 (stage II) | - | - | 6.2-13.2% 3.6-11.2% | <ul style="list-style-type: none"> • ↑ CH₄ content by 13.7-25.3% • ↑ CH₄ prod. rate by 5.5-36.9% • ↑ CH₄ content by 0.7-9.1% • ↑ CH₄ prod. rate by -2.3-16.6% | [34] |
| Wheat bran pellets Coppiced woodlands Orchard pruning | Pyrolysis: 800°C Pyrolysis: 500°C Pyrolysis: 500°C | 25 g L ⁻¹ | Food waste fermentate | Anaerobic methanogenic culture (from WAS AD) | - | 20 | 7.5 | | 200–250 (NH ₃ -N) | | <ul style="list-style-type: none"> • After acclimation: ↑ rate of VFAs degradation and ↓ lag-phase; • No adsorption of ammonia by BC | [45] |
| Wheat straw | Pyrolysis: 350, 450, 550°C | 5% w/w (chicken manure) | Chicken manure (+ water) | Chicken manure meso. AD | Inoculum: 20% of WV of AD reactor | 35 | Initial: 6.7-7.6 | 3450-3540 (average) | 472-600 (max) | up to 25% TAN | <ul style="list-style-type: none"> • ↑ Cum. CH₄ yield • ↓ TAN compared to control | [82] |
| Fruit wood | Pyrolysis: 350, 450, 550°C | | | | | | Initial: 6.7-7.0 | 3440-3650 (average) | 421-634 (max) | | <ul style="list-style-type: none"> • ↑ Cum. CH₄ yield up to 69% • ↓ TAN compared to control • ↓ FAN up to 58% compared to control | |
| Air-dried chicken manure | Pyrolysis: 350, 450, 550°C | | | | | | Initial: 6.9-7.3 | 3880-3960 (average) | 496-701 (max) | | <ul style="list-style-type: none"> • ↑ Cum. CH₄ yield • ↓ TAN compared to control | |

559

560

561 Table 5. Ammonium adsorption capacity of different BCs (NA: not available)

| Biochar production | | | Biochar properties | | | | Adsorption | | | | Reference |
|--------------------|-------------------------------|---|--------------------|--|--|---|--|--|-------|--|-----------|
| Feedstock | Pyrolysis temperature [°C] | BET-SA [m ² g ⁻¹] | pH | CEC [cmol _c kg ⁻¹] | Total pore volume [mL g ⁻¹] | Source of NH ₄ ⁺ -N | Initial NH ₄ ⁺ -N [mg L ⁻¹] | NH ₄ ⁺ adsorption capacity [mg NH ₄ ⁺ -N g ⁻¹ biochar] | | | |
| Oak wood | 400-450 | NA | 9.9 | 59.4 ± 8.1 | NA | Ammonium solution | 1000 | 100.9 ± 3.4 | | | |
| Oak wood | - | NA | 9.7 | 105.8 ± 12.1 | NA | | - | 129.4 ± 34.8 | | | |
| Greenhouse waste | - | NA | 10.6 | 109.5 ± 21.8 | NA | - | - | 118.2 ± 26.9 | | | |
| Municipal waste | - | NA | 9.5 | 65.7 ± 16.2 | NA | - | - | 137.3 ± 0.6 | | | |
| Presscake from AD | - | NA | 10.3 | 51.0 ± 5.5 | NA | - | - | 105.8 ± 11.5 | [154] | | |
| Oak wood | 600-650 | NA | 10.3 | 76.6 ± 0.7 | NA | - | - | 114.4 ± 3.4 | | | |
| Oak wood | - | NA | 8.6 | 65.2 ± 20.2 | NA | - | - | 123.5 ± 28.7 | | | |
| Greenhouse waste | - | NA | 11.0 | 146.2 ± 32.3 | NA | - | - | 99.3 ± 28.5 | | | |
| Municipal waste | - | NA | 10.2 | 67.9 ± 12.5 | NA | - | - | 128.3 ± 6.7 | | | |
| Presscake from AD | - | NA | 10.1 | 52.6 ± 11.5 | NA | - | - | 136.2 ± 18.1 | | | |
| Hardwood | 600 | 147.0 | 9.80 | NA | 0.176 | AD slurry | 500–580 | 114.2 | | | |
| Corncoobs | - | 23.0 | 8.92 | NA | 0.098 | - | - | 108.9 | [159] | | |
| Mixed sawdust | - | 6.80 | 8.60 | NA | 0.038 | - | - | 24.7 | | | |

| | | | | | | | | | |
|---|---------|---------|------|----|-------|-------------------------------|------------------------------|----------------------------|-------|
| Mixed wood | 600 | 273.623 | 9.80 | NA | 0.176 | Swine manure AD slurry | 1400 mg N L ⁻¹ | 44.64 ± 0.602 | |
| Rice husk | / | 10.995 | 7.80 | NA | 0.038 | - | - | 39.8 ± 0.54 | [155] |
| 1:2 (v/v) mix: - paper sludge - wheat husks | 500 | NA | NA | NA | NA | During anaerobic digestion | 1626 mg TAN kg ⁻¹ | 2.4 mg TAN g ⁻¹ | |
| | | - | - | - | - | - | 2126 mg TAN kg ⁻¹ | 2.0 mg TAN g ⁻¹ | |
| | | - | - | - | - | - | 3126 mg TAN kg ⁻¹ | 4.2 mg TAN g ⁻¹ | [41] |
| | | - | - | - | - | - | 4126 mg TAN kg ⁻¹ | 4.5 mg TAN g ⁻¹ | |
| | | - | - | - | - | - | 6626 mg TAN kg ⁻¹ | 6.8 mg TAN g ⁻¹ | |
| Fruitwood | 800-900 | NA | NA | NA | NA | NH ₄ Cl solution | NA | 2-3 mg N g ⁻¹ | [44] |

563 4.3. *Effects on microbial populations*

564 Many authors reported the positive effects of BC on microbial populations in AD (Table
565 6). In semi-continuous and continuous AD experiments, BC addition improved methane
566 production [34] keeping it stable even with rising OLRs [29,49] and resulted in higher
567 COD removal rates [43,50,51].

568 Although there is a good agreement among the studies in terms of enhancement of AD
569 process performances (11-30 % reduction of lag phases and 11-50 % increments of
570 methane production rates) (Table 6), there is not a common hypothesis to explain BC
571 influence on microbial populations, and different possible mechanisms are proposed as
572 follows.

573 4.3.1. *Microbial attachment and acclimation*

574 Immobilization and acclimation of balanced microbial consortia on various support media
575 are commonly adopted for counteracting various drawbacks of AD processes [160],
576 thanks to the intensification of syntrophic conversion relationships, along with a major
577 resistance to inhibition effects [161]. Wang et al. (2018) [50] found that BC acted as an
578 inert core for microbial aggregation, resulting in a higher microbial growth rate and
579 accelerating sludge granulation. Cooney et al. 2016 [53] investigated the possibility of
580 accelerating biofilm formation by adding BC during the start-up of a packed bed
581 anaerobic digester at pilot scale. In a relatively short time, the AD system reached stable
582 and good performances, suggesting the rapid development of biofilms rich in active
583 methanogens.

584 High SSA, proper porosity structure and particle size, superficial hydrophobicity seemed
585 to be important properties favouring microbial immobilization [12,52]. Further,
586 conductive materials may act as ideal support media due to their surface hydrophobicity

587 and porous structure [54,55]. In particular, macropores can help the attachments of
588 bacterial cells [19,162,163]. Lü et al. (2016) [44] explored the influence of different
589 particle sizes of BC (2-5 mm, 0.5-1 mm, 75-150 μm) on the microbial distribution during
590 the AD of glucose under ammonium stress. They stated that bacteria could access more
591 easily fine particles than coarse particles. As a consequence, the attachment and
592 colonization of microbial populations on BC can limit the risks of wash-out, accelerate
593 the acclimation of microbes during substrate-induced inhibition, reduce the distance
594 between syntrophic bacteria and methanogens, facilitate interspecies electron transfer and
595 exchanges of VFAs or other metabolites [16,36,137]. Li et al. (2018) [46] found that
596 methanogens survived under acidic stress in presence of BC during co-digestion of FW
597 and WAS. Further, the immobilization of microbes could significantly attenuate ammonia
598 inhibition [43,44]. The colonization of porous materials by microbes can alter the
599 dominant species, making them more resistant and more rapidly acclimatized to substrate-
600 induced inhibition [29,36,164]. Magnetic BC favoured the enrichment of acido/acetogens
601 and methanogens absorbed on its surface shortening the microbial contact distance, thus
602 VFAs produced by acido/acetogen bacteria could be more quickly transported to
603 methanogens than in control digesters during AD of OFMSW in batch tests [16,139]. The
604 distance of less than 1 μm has been reported to be essential for the oxidation of VFAs and
605 hydrogen production [19,165].

606 4.3.2. *Selective colonization of functional microbes*

607 The effect of BC addition on microbial communities was investigated with the aim of
608 identifying the most abundant bacterial and archaeal populations and, indirectly, the main
609 interspecies interactions (Table 6). In detail, most studies focused on: biofilm formation
610 [26,48,53]; shifts of microbial populations [28,34,46,47]; selective enrichment of

611 microbial -DIET partners [38,44,49,50]; promotion of DIET [38,44–46,49,51,139]. The
612 relevant anaerobic bacteria and archaea enriched in BC amended digesters are reported
613 in Table 7, together with BC properties, substrate and inoculum used in AD tests. Many
614 bacterial species were found in reactors supplemented with BCs, none of them identified
615 as more recurrent. Among archaeae, most studies identified *methanosaeta*,
616 *methanosarcina*, *methanobacterium*, and *methanolinea* species in BC amended reactors.
617 Different studies [44,48,51] investigated the spatial distribution of bacteria and archaea
618 by dividing sludge samples into different fractions, from suspended to attached to BC. Lü
619 et al. (2016) [44] postulated an explanation for the spatial distribution of methanogens
620 into BC pores by their cell morphology and dimension. The short fibrous form of
621 *Methanosaeta* (0.8-7 μm in size) could explain its attachment into internal and external
622 pores, while the long fibrous form of *Methanobacterium* (1.2-120 μm in length) could
623 limit its penetration into BC pores [44,166].

624 4.3.3. Promotion of syntrophic metabolisms

625 Many studies (Table 7) suggested that BC addition may improve electron transfer
626 mechanisms between anaerobic bacteria and archaea closely attached to BC surface. The
627 overall AD efficiency depends on effective syntrophic interactions between bacteria and
628 methanogens exchanging electrons to satisfy their energy requirements [56], happening
629 through various routes:

- 630 - Indirect interspecies electron transfer (IIET) via soluble (i.e. hydrogen, formate,
631 acetate) [167,168] and insoluble (humic substances) [169,170] compounds;
- 632 - Direct interspecies electron transfer (DIET) via electrical conductive pili, membrane-
633 bound electron transport proteins, and conductive materials (i.e. magnetite, biochar,
634 granular activated carbon, carbon cloth) [171–173].

635 In IIET hydrogen and formate operate as electron shuttles between syntrophic-producing
636 bacteria and consuming-methanogens [56]. Diffusion regulates the transfer of a
637 metabolite between microorganisms, as defined by Fick's Law [174]: the shorter the
638 distance, the higher the flux of metabolites between microbes. Thereby, when cells
639 aggregate the rate of interspecies hydrogen transfer is enhanced by the moment anaerobic
640 bacteria and methanogenic archaea form compact structures acting as an organ [174,175].
641 However, the diffusion of soluble metabolites is considered a relatively slow mechanism
642 of energy and information transfer [173] and hydrogen IET is regarded as a bottleneck in
643 methane production [171]. On the contrary, DIET consists in the formation of an electric
644 current between electron-donating and electron-acceptor microorganisms without the
645 mediation of electron shuttles [56]. DIET could be faster and more specific compared to
646 IIET [173,176]. Park et al. (2018) [171] reported that direct exchange of electrons via
647 conductive pili was observed in co-cultures between *geobacter metalliriducens* and
648 *geobacter sulfurreducens* [176], *methanosaeta harundinacea* [177], or *methanosarcina*
649 *barkeri* [178]. Conductive materials such as magnetite [179], GAC [180], and BC [57]
650 were shown to effectively mediate DIET between syntrophic partners.

651 Based on changes in the microbial community composition, many studies justified the
652 enhancement of AD activity by means of the improvement of hydrogen and formate
653 interspecies transfer mechanisms or, more frequently, by DIET via conductive biochar
654 (Table 7). These findings are usually based on indirect observations, i.e. the enrichment
655 of bacterial and archaeal species able to participate to DIET function as potential partners.
656 Martinez et al. (2018) [47] found an enrichment of homoacetogenic bacteria, as
657 *Clostridium*, *Eubacterium* and *Syntrophomonas*, and H₂ using methanogens through the
658 analysis of microbial communities in digesting WAS and orange peels with BC,

659 suggesting the formation of co-cultures enhancing methane production. Zhao et al. (2016)
660 [51] observed the selective enrichment on BC of *Geobacter* and *Methanosaeta* during
661 AD of synthetic wastewater with butyrate and propionate in UASB reactors. They
662 suggested that butyrate and propionate could be degraded via DIET in the presence of a
663 conductive material, and they found abundance of *Syntrophomonas* and *Smithella*,
664 concluding that the metabolism via interspecies H₂ transfer for butyrate and propionate
665 degradation was probably present. Wang et al. (2018) [50] showed that the microbial
666 community analysis in a BC amended reactor during AD of synthetic wastewater resulted
667 in the selective enrichment of potential DIET-partners, as *Geobacter* and *Bacteroidetes*,
668 as well as archaea *Methanosaeta* and *Methanosarcina*. They suggested that BC could
669 enhance DIET among electrogenic microbes and archaea, improving the electron transfer
670 characteristics of granular sludge, as well as COD removal and methane yield. Wang et
671 al. (2018) [38] found that the addition of different BC doses increased methane production
672 rate and shortened the lag phase during mesophilic AD of dewatered WAS and FW.
673 According to the authors, BC counteracted the pH decrease due to VFAs accumulation
674 through its buffering capacity, and it seemed to promote DIET.

675 Aside from physical properties as SSA and porosity, favouring microbial colonization,
676 other chemical and conductive characteristics of BC may be crucial in the promotion of
677 electrons transfer. The role of electrical conductivity (EC) of BC in AD has been
678 investigated in literature[59]. EC of digestate has been reported to increase in presence of
679 BC [50,151]. However, the EC of digestate seemed unrelated to the conductivity of BC,
680 which varies depending on the metabolism and composition of microbial species [56].
681 The capability of BC in promoting DIET appeared to be comparable to that of GAC, even
682 if the EC of BC was roughly 1000 times lower [57,171]. Barua and Dhar (2017) [181]

683 reported that multi-species aggregates from anaerobic digesters exhibited conductivity
684 ranging 0.2-36.7 $\mu\text{S cm}^{-1}$, suggesting its relation with DIET via conductive pili. Martins
685 et al. (2018) [56] stated that conductive materials could have a similar role of humic
686 substances in DIET, acting as electron shuttles by receiving and donating electrons. Wang
687 et al. (2019) [49] suggested that BC from sawdust may act as a temporary electron
688 acceptor for VFAs oxidation during thermophilic AD. They found a significantly higher
689 and more stable methane yield at higher OLR values during anaerobic co-digestion in
690 semi-continuous mode with the addition of BC from sawdust, while they did not show
691 any enhancement of AD with the addition of BC from sewage sludge. The main effects
692 of sawdust BC seemed to be the enhancements of microbial activities and syntrophic
693 oxidation of VFAs. The EC of both BCs was similar, suggesting it was not a determinant
694 factor in the promotion of syntrophic oxidation of VFAs. Instead, the authors speculated
695 that the presence of redox-active phenazine structures in the BC from sawdust could
696 promote VFAs degradation via DIET. Thereby, for better investigating whether BC from
697 sawdust could substitute hydrogen as electron acceptor in syntrophic oxidation of VFAs,
698 they conducted a series of batch experiments with butyrate or propionate as substrates in
699 which methanogenesis was inhibited. The control reactors did not show degradation of
700 butyrate and propionate, while the addition of BC stimulated their oxidation as well as
701 the production of acetate, supporting the hypothesis of the electron-accepting capacity of
702 BC in the syntrophic process.

703 The adoption of a further control in AD tests amended with a non-conductive material
704 can be crucial to investigate whether the stimulatory effects of BC on methane production
705 can be more closely linked to its physical properties (i.e. SSA and porosity) rather than
706 its electrical properties [56]. This was evident in the study of Cruz Viggi et al. (2017)

707 [45], where they introduced two controls without BC and with non-conductive silica sand
708 for AD of FW. They found that VFAs degradation and methane production were faster
709 in the case of BC amended reactors than both the control reactors, suggesting the
710 predominant influence of the electrical properties of BCs.

711 Table 6. A summary of selected works focusing on positive effects of BC on microbial populations in AD processes

| Biochar | Anaerobic digestion | | | | | | | | | References |
|----------------------|---|------------------------------|--|------------------------------------|------------------|---|---|---|------|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | | |
| Sawdust | Pyrolysis: 500°C 1 h 20°C min ⁻¹ | 10 g L ⁻¹ | Food waste (TS = 94.2 g L ⁻¹) | WAS (TS = 89.1 g L ⁻¹) | 55°C | Batch V = 120 mL S/I (VS) = 0.25 - 3 | By ↑ OLR: ↓ lag time ↑ CH ₄ prod. rate | ↑ buffer capacity Microbial attachment and acclimation on BC Promotion of DIET Selective succession of microbes | [46] | |
| Pine sawdust | Pyrolysis: 650°C 20 min | 8.3 - 33.3 g L ⁻¹ | Food waste (bread) (TS = 61.2%) | WWTP-sludge (heated at 95°C) | 35°C | Batch V = 100 mL TPAD | 1 st phase: ↓ lag phase Faster VFAs generation ↑ H ₂ prod. rate ↑ Cum. H ₂ prod. 2 nd phase: ↓ lag phase (41-45%) Faster VFA degradation ↑ CH ₄ prod. rate | 1 st phase: Promotion biofilm formation Providing temporary nutrients Buffering pH by BC 2 nd phase: Promotion methanogenic biofilm formation No buffering pH by BC | [26] | |
| Macadamia nut shells | Pyrolysis: 350°C 2 h | 33.3 g L ⁻¹ | Food waste + water + (↑ N-NH ₄ conc.) | AD from | Room temperature | CIC reactor V = 4.6 L HRT=16 h | ↑ COD removal by 15% | ↓ NH ₃ inhibition by BC ↑ Alkalinity by BC ↓ VFAs accumulation | [43] | |

| Biochar | | Anaerobic digestion | | | | | | | | References |
|-------------|--|--|---|---|-------------|--|--|---|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature | Experimental mode | Results | Possible mechanisms | | |
| | • 10°C min ⁻¹ | | | | [°C] | | | <ul style="list-style-type: none"> • ↑ Selective enrichment of functional microbes • Microbial attachment and acclimation on BC | | |
| | | | | (SS = 34 g L ⁻¹) | | | | | | |
| Sawdust | Pyrolysis: <ul style="list-style-type: none"> • 500°C • 1.5 h • 10°C min⁻¹ | 2 - 15 g L ⁻¹ | Dewatered WAS <ul style="list-style-type: none"> + Food waste (TS = 8.8%) + water | Meso. AD from brewery factory (TS = 6.8%) | 35°C | Batch <ul style="list-style-type: none"> Serum bottles WV = 90 mL S/I = 0.75 - 3 (VS) | <ul style="list-style-type: none"> • ↓ lag phase by 27.5–64.4% • ↑ Max CH₄ prod. rate by 22.4%–40.3% | <ul style="list-style-type: none"> • Buffer capacity by BC (alleviate ↓ pH due to ↑ VFAs) • ↑ DIET by BC (temporary electron acceptor, due to the richness of surface functional groups) • Selective enrichment of microbial DIET-partners by BC | | [38] |
| Sawdust | Pyrolysis: <ul style="list-style-type: none"> • 500°C | 15 g L ⁻¹ | Food waste + Sewage sludge (TS = 9.2%) <ul style="list-style-type: none"> FW/Sludge = 4:1 (TS) | Brewery Meso. AD (TS = 5.2%) | 55°C | Semi-continuous <ul style="list-style-type: none"> Serum bottles WV = 150 mL ↓ HRT, ↑ OLR | <ul style="list-style-type: none"> • ↑ CH₄ yield by 16.0%–55.2% • Stable CH₄ prod. at ↑ OLRs | <ul style="list-style-type: none"> • ↑ VFAs syntrophic oxidation by BC • High SA of BC → microbial attachment (?), closer association of syntrophic partners • Electroactive functional groups of BC → stimulation of DIET (?) | | [49] |
| Corn stover | Gasification: <ul style="list-style-type: none"> • 850°C | 0.25 - 1.0 g d ⁻¹ (2 nd phase) | Primary sludge + WAS (TS = 7.01%) | WWTP AD | 55°C | Semi-continuous <ul style="list-style-type: none"> V = 500 mL TPAD: | <ul style="list-style-type: none"> • ↑ % CH₄ by 13.7-25.3% • ↑ CH₄ prod. rate by 5.5-36.9% | <ul style="list-style-type: none"> • Adsorption and precipitation of CO₂ by BC • ↓ TAN • ↑ Alkalinity • ↑ Macro-/micro-nutrients in digestate • Shift in bacterial community | | [34,182] |

| Biochar | | Anaerobic digestion | | | | | | | References |
|-------------|--------------|-------------------------------------|------------------------------------|-----------------|------------------|--------------------------|---|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | |
| k | | | | | | | TPAD: | <ul style="list-style-type: none"> • Inhibition at high BC doses | |
| | | | | | | | 1 ph. (TS = 6.14%) | | |
| | | | | | | | 2 ph. (TS = 3.77%) | | |
| Pine | - | - | - | - | - | - | <ul style="list-style-type: none"> • ↑ % CH₄ by 0.7-9.1% • ↑ CH₄ prod. rate by 2.3-16.6% | | |
| Corn stover | Gasification | 1.82 g BC g ⁻¹ TS sludge | -3.64 g ⁻¹ WWTP sludge | Thermo. WWTP AD | 55°C | Batch V = 600 mL 26 days | <ul style="list-style-type: none"> • ↑ % CH₄ (88.5-96.7%) • ↑ CH₄ yield | <ul style="list-style-type: none"> • ↑ buffer capacity • ↑ electrical conductivity in digester (+37%) → ? ↑ extracellular electron transfer • ↓ NH₃ inhibition • CO₂ sequestration by BC • ↑ macro-/micro-nutrients in digestate | [151] |
| Rice husks | Gasification | 1%, w/w | 3% Cattle manure + water (TS = 5%) | - | 35°C | Batch V = 1400 mL | <ul style="list-style-type: none"> • ↑ Biogas prod. by 31% by 1% BC • ↓ % CH₄ by 7% by 1% BC • No benefits by 3% BC | <ul style="list-style-type: none"> • ? Microbial biofilm formation on BC | [30] |

| Biochar | | Anaerobic digestion | | | | | | | References | |
|-------------------|---|----------------------------|--------------------------------------|--|-------------|--|---|---|------------|--|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature | Experimental mode | Results | Possible mechanisms | | |
| | | | | | [°C] | | | | | |
| Rice husks | Gasification: 900-1000°C | 1%, 3% w/w | Cattle manure + water (Mix: TS = 5%) | - | 25-30°C | Semi-continuous WV = 12 L HRT = 20 days | <ul style="list-style-type: none"> • ↑ Biogas prod. by 4-5% by BC • ↓ % CH₄ | <ul style="list-style-type: none"> • ? Not enough time for biofilm formation | [31] | |
| Dry dairy manure | Pyrolysis: 350°C, 3 h, 10°C min ⁻¹ | 0, 1, 10 g L ⁻¹ | Dry dairy manure | Inoculum from lagoon, fed with dried manure (TS = 115.85 g L ⁻¹) | 20°C | Batch: Serum bottles V = 280 mL S/I = 1 (VS) | <ul style="list-style-type: none"> • ↑ CH₄ yield up to 26.5% • ↓ lag phase • ↓ total VFAs | <ul style="list-style-type: none"> • No biofilm formation on BC (?) • No effects on DIET by BC (?) • ↑ Alkalinity and ↑ pH (buffer capacity) | [27] | |
| - | - | - | - | - | 35°C | - | <ul style="list-style-type: none"> • ↑ CH₄ yield up to 24.9% • ↓ lag phase • ↓ total VFAs | <ul style="list-style-type: none"> • ↑ Alkalinity and ↑ pH (buffer capacity) | | |
| - | - | - | - | - | 55°C | - | <ul style="list-style-type: none"> • ↑ CH₄ yield up to 24.7% • ↓ lag phase • ↓ total VFAs | <ul style="list-style-type: none"> • ↑ Alkalinity and ↑ pH (buffer capacity) | | |
| Vineyard prunings | Pyrolysis: 550°C | 10, 30 g L ⁻¹ | Co-digestion: | WWTP AD | 37°C | Batch: V = 250 mL | <ul style="list-style-type: none"> • ↓ lag phase | <ul style="list-style-type: none"> • Promotion of syntrophic metabolism by BC • Adsorption of inhibitors | [47,183] | |

| Biochar | | Anaerobic digestion | | | | | | | References |
|---------------|--------------------|-------------------------|--|---------------------------------|------------------|---|---|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | |
| | | | <ul style="list-style-type: none"> Orange peels (TS = 311g kg⁻¹) Sewage sludge (TS = 28.7g kg⁻¹) | (TS = 35.5 g kg ⁻¹) | | Semi-continuous: • V = 3 L • HRT = 10-30 days | • ↑ CH ₄ prod. • Adhesion and growth of microorganisms by BC SA | | |
| Coconut shell | Pyrolysis: • 450°C | 1:1 (TS) | Citrus peel (TS = 16.6%) | WWTP AD (TS = 11.0 %) | 35°C | Batch V=500 mL S/I = 0.31 - 0.33 (VS) | • ↓ lag phase • ↑ Cum. CH ₄ prod. | • Limonene adsorption by BC | [40] |
| Rice husk | - | 1:1 (TS) | - | - | - | - | • ↓ lag phase • ↑ Cum. CH ₄ prod. | • Limonene adsorption by BC • Adhesion and growth of microbes on BC | |
| Wood | - | 1:1; 1:2; 1:3; 2:1 (TS) | - | - | - | - | • ↓ lag phase • ↑ Cum. CH ₄ prod. | • Limonene adsorption by BC • Adhesion and growth of microbes on BC | |

| Biochar | | Anaerobic digestion | | | | | | | | References |
|--|--|--------------------------|----------------------------|---------------------------------------|------------------|--|--|--|-------|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | | |
| Rice straw | In solution: FeCl ₃ • 2h | 0.5% w/w | OFMSW + water (TS = 1.64%) | OFMSW + thermo. AD water (TS = 2.19%) | 35°C | Batch | • ↓ lag phase | • No effects on NH ₃ by BC | [139] | |
| (3.2 g FeCl ₃ :10 g rice-straw) | Carbonization: • 500°C • 2h | | | | | Serum bottles V = 500 mL, S/I = 1 (VS) | • ↑ CH ₄ prod. by 11.69% | • No effects on pH by BC | | |
| | | | | | | | | • ↑ syntrophic associations of bacteria on BC | | |
| | | | | | | | | • (?) DIET by high conductivity of BC | | |
| Rice straw | Carbonization: • 500°C • 2h | - | - | - | - | - | • ↓ lag phase | • No effects on NH ₃ by BC | | |
| | | | | | | | • ↓ CH ₄ prod. | • No effects on pH by BC | | |
| Cow manure | Pyrolysis: • 500°C • 4 h • 100°C h ⁻¹ | 2 – 14 g L ⁻¹ | Dry beer lees (TS = 62.5%) | Meso. WWTP AD (TS = 36.7%) | 35°C | Batch | • ↓ lag phase | • Promotion of DIET by BC conductive properties (?) | [28] | |
| | | | | | | WV = 150 mL | • ↑ Max cum. CH ₄ prod. up to 82.9% | • Microbial colonisation and biofilm formation on BC (?) | | |
| | | | | | | TS = 25% | | • ↑ Alkalinity and ↑ pH (buffer capacity) | | |
| | | | | | | S/I = 3 (TS) | | • Selective enrichment of methanogens by BC | | |
| - | - | - | - | Thermo. WWTP AD (TS = 38.5%) | 55°C | - | • ↓ lag phase | - | | |
| | | | | | | | • ↑ Max cum. CH ₄ prod. up to 47.2% | | | |
| Rice straw | Pyrolysis: • 500°C | 4 g L ⁻¹ | Synthetic wastewater | WWTP sludge | 35°C | UASB | UASB: • ↑ COD removal rate | • No effects on pH by BC | [50] | |
| | | | | | | | | • BC inert core for microbial aggregation | | |

| Biochar | | Anaerobic digestion | | | | | | | References |
|-----------|------------|---------------------|--|--------------------------------|------------------|---|--|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | |
| | | | | (SS = 25.2 g L ⁻¹) | | <ul style="list-style-type: none"> Continuous mode V = 5500 mL HRT = 12 h, 12-6 h Batch Serum bottles V = 550 mL | <ul style="list-style-type: none"> ↑ VFAs degradation ↑ gran. sludge conductivity and quality Batch: <ul style="list-style-type: none"> ↓ lag phase by 28.6% ↑ biogas yield ↑ %CH₄ | <ul style="list-style-type: none"> Selective enrichment of microbial DIET-partners by BC | |
| - | Biochar | - | Grease trap wastewater (TSS = 1.04 g L ⁻¹) | - | 37°C | Biochar packed bed anaerobic digester +1500 L+ 1500 L • ↓ HRT (3.1 – 1.8 days) | <ul style="list-style-type: none"> COD removal: 68% Total VFAs: from 4.7 (feed) to 1.46 g L⁻¹ (effluent) %CH₄ > 60% Start-up: 59 days | <ul style="list-style-type: none"> ↑ Methanogenic biofilm communities on BC BS as packing material for growth and retention of biofilm | [53] |

| Biochar | | Anaerobic digestion | | | | | | | | References |
|--------------------|-------------------------|----------------------|--------------------------------|---|------------------|---------------------|--|--|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | | |
| Wheat bran pellets | Pyrolysis: • 800°C • 3h | 25 g L ⁻¹ | Food waste fermentate | Methanogenic culture (from WAS digestate) | 20°C | Batch V = 120 mL | After acclimation: • ↑ rate of VFAs degradation • ↓ lag-phase | • No effects on pH via BC • No effects on NH ₃ via BC • Promotion of IET by BC | | [45] |
| Coppiced woodlands | Pyrolysis: • 500°C | - | - | - | - | - | | - | | |
| Orchard pruning | Pyrolysis: • 500°C | - | - | - | - | - | | - | | |
| Fruitwood | Pyrolysis: • 800-900°C | 10 g L ⁻¹ | Glucose (6 g L ⁻¹) | AD gran. | 35°C | Batch Serum bottles | • ↓ lag phase by 5.9-23.9% • ↑max CH ₄ prod. rate by 23.5-47.1% • Faster VFAs degradation | • NOT NH ₄ adsorption • NOT ↑ buffer capacity • DIET promotion via BC • Affinity of methanogens with BC | | [44] |

| Biochar | | Anaerobic digestion | | | | | | | | References |
|------------|-------------------------------|----------------------|--|---|------------------|--------------------------------|---|---|---------|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | | |
| | | | Ammonium: 0.26, 3.5, 7 g-N L ⁻¹ | | | | | | | |
| Fruit wood | Pyrolysis: • 800°C | 10 g L ⁻¹ | Nutrient solution + glucose (2 - 8 g L ⁻¹) | Meso. pulp sewage digestate (1 g VS L ⁻¹) | 35°C | Batch Serum bottles V = 500 mL | • ↓ lag phase by 11.4-30.3% • ↑ Max CH ₄ prod. rate by 5.2-86.6% • ↑ VFAs production and degradation | • Selective colonization of functional microbes by BC • Not ↑ buffer capacity • ? Biofilm growth on BC • ? Promotion of DIET or Hydrogen IET by BC | [48] | |
| Pine wood | Pyrolysis: • 600°C • 2h | 5 g L ⁻¹ | Synthetic wastewater with butyrate | WAS digestate (TSS = 13.1 g) | 37°C | UASB WV = 1000 mL | • ↑ CH ₄ prod. rate by 25% • ↑ COD removal | • ↑ Butyrate degradation via DIET in UASB via BC • Selective enrichment of microbial DIET-partners by BC | [51,57] | |
| | | | Synthetic wastewater with propionate | | | | • ↑ CH ₄ prod. rate by 16% • ↑ COD removal | • ↑ Propionate degradation via DIET in UASB via BC • Selective enrichment of microbial DIET-partners by BC | | |

| Biochar | | Anaerobic digestion | | | | | | | | References |
|-----------|------------|-----------------------|-----------------------------------|------------------------------|------------------|--|---|---|--|------------|
| Feedstock | Production | Dose of biochar | Substrate | Inoculum | Temperature [°C] | Experimental mode | Results | Possible mechanisms | | |
| - | Biochar | 2.5 g L ⁻¹ | Synthetic wastewater with ethanol | WAS digestate (TSS = 13.1 g) | 37°C | UASB WV = 1000 mL ↓ HRT (24 - 8 h) | <ul style="list-style-type: none"> • ↑ COD removal: ≥ 93% (control: 75-83%) • ↑ CH₄ prod. rate | <ul style="list-style-type: none"> • (?) Promotion syntrophic metabolism via DIET with BC in UASB reactors | | [29] |

CIC: controlling internal circulation; COD: chemical oxygen demand; DIET: direct interspecies electron transfer; FW: food waste; HRT: hydraulic retention time; IET: interspecies electron transfer; OLR: organic loading rate; OFMSW: organic fraction of municipal solid waste; SA: surface area; S/I: substrate to inoculum ratio; SMP: specific methane potential; SS: suspended solids; TPAD: two phased anaerobic digestion; TSS: total suspended solids; UASB: up-flow anaerobic sludge blanket; VFA: volatile fatty acid; WAS: waste activated sludge; WWTP: wastewater treatment plant; WV: working volume; V: volume.

712

713

714 Table 7. A summary of studies reporting the selective enrichment of bacteria and archaea by BC addition during AD processes

| Biochar production | | Biochar properties | | | Anaerobic digestion | | | | | Reference |
|--|-------------------------------|--|---|---|---------------------|-----------------------|--------------------------|---|---|-----------|
| Feedstock | Pyrolysis temperature [°C] | BET-SSA [m ² g ⁻¹] | Electrical conductivity [S m ⁻¹] | Total pore volume [cm ³ g ⁻¹] | Temperature [°C] | Substrate | Identification technique | Enriched Bacteria | Enriched Archaea | |
| Wheat bran pellets | 800 | 55 ± 1 | 49.9 | 0.0445 | 20 | Food waste fermentate | FISH- CLSM | | More Methanosarcina-like Archaea rather than Methanosaeta like Archaea | [45] |
| Coppiced woodlands | 500 | 61 ± 1 | 1.6 | 0.0483 | | | | | | |
| Orchard pruning | 500 | 13.7 ± 0.5 | 0.5 | 0.0165 | | | | | | |
| Macadamia nut shells | 350 | 12.7 | - | - | Room temperature | Food waste + water | 16S rRNA sequencing | Bacteroidales Anaerolineales Syntrophobacterales | Methanoregulaceae Methanotrichaceae Methanobacteriaceae | [43] |
| Fruitwoods | 800-900 | - | - | - | 35 | Glucose | 16S rRNA sequencing | Enterobacteriaceae | Methanobacterium Methanosaeta Methanosarcina | [44] |
| Fruit wood | 800 | - | - | - | 35 | Glucose | 16S rRNA sequencing | Syntrophomonas Clostridium Clostridiaceae | Methanobacterium Methanosaeta Methanosarcina | [48] |
| Rice straw + FeCl ₃ (3.2 g FeCl ₃ :100 g rice-straw) | 500 | 51.14 | - | 0.0328 | 35 | OFMSW + water | 16S rRNA sequencing | Bacteroides Clostridiaceae Porphyromonadaceae Moraxellaceae | Methanosarcina Methanobacterium OTU in Methanomicrobiales Methanosaeta | [139] |
| Rice straw | 500 | 111.5 | - | - | 35 | Synthetic wastewater | 16S rRNA sequencing | Bacteroidetes unclassified (23.65%) Bacteroidales unclassified (9.19%) | Methanosaeta (77.18%) Methanosarcina (11.65%) | [50] |

| | | | | | | | | | | | |
|-------------------|-----|-----------|-----------------------------|--------|----|--------------------------------------|---------------------|---|--|------|--|
| | | | | | | | | | Treponema (6.43%) | | |
| | | | | | | | | | Smithella (5.56%) | | |
| | | | | | | | | | Brooklawnia (5.45%) | | |
| | | | | | | | | | Geobacter (5.42%) | | |
| Sawdust | 500 | 248.6 | - | - | 35 | Dewatered WAS + food waste + water | 16S rRNA sequencing | Anaerolineaceae Porphyromonadaceae | Methanosaeta Methanobacterium Methanolinea | [38] | |
| Cow manure | 500 | 112.6 | - | 0.0156 | 35 | Dry beer lees | 16S rRNA sequencing | Chloroflexi Bacteroides Proteobacteria | Methanospirillum Methanosarcina Methanolinea | [28] | |
| Vineyard prunings | 550 | 240 ± 4.8 | - | - | 37 | Orange peels | 16S rRNA sequencing | Bellilinea Trepomena Clostridium Petrimonas Proteiniphilum Bacteroides | Methanosaeta Thermogymnomonas Methanolinea Methanofollis | [47] | |
| Vineyard prunings | 550 | 240 ± 4.8 | - | - | 37 | Sewage sludge | 16S rRNA sequencing | Clostridium Longilinea Curvibacter Eubacterium Syntrophomonas | Methanosaeta Methanolinea Thermogymnomonas Methanobacterium | | |
| Pinewood | 600 | 209 | .33 μS cm ⁻¹ | - | 37 | Synthetic wastewater with butyrate | 16S rRNA sequencing | Attached sludge: Geobacter Thermanaerovibrio Syntrophomonas | Attached sludge: Methanosaeta Methanosarcina Methanospirillum | [51] | |
| Pinewood | 600 | 209 | 4.33 μS cm ⁻¹ | - | 37 | Synthetic wastewater with propionate | 16S rRNA sequencing | Attached sludge: Geobacter Smithella | Attached sludge: Methanosaeta Methanobacterium | | |

| | | | | | | | | | | |
|-------------|-----|-------------|--------------------------------|------|----|---------------------------------|---------------------|--|---|------|
| | | | | | | | | Syntrophus | Methanosphaerula | |
| Biochar | - | - | - | - | 37 | Grease trap waste wastewater | 16S rRNA sequencing | Attached to BC: Aminobacterium Syntrophomonas Sporanaerobacter Escherichia | Attached to BC: Methanobacterium Methanosarcina Methanobrevibacter | [53] |
| Sawdust | 500 | 248 ± 34 | - | - | 55 | Food waste | 16S rRNA sequencing | Coprothermobacter (36.3%) Fervidobacterium (20.0%) Syntrophothermus Treponema | Methanosaeta (43.9%) Methanosarcina (15.8%) | [46] |
| Corn stover | 850 | 315.2 | - | 0.09 | 55 | Primary sludge + WAS | 16S rRNA sequencing | Firmicutes (Clostridia) Bacteroidetes Proteobacteria | Methanothermobacter (>90%) Methanosarcina | [34] |
| Pine | | 353.1 | - | 0.23 | | | | | | |
| Sawdust | 500 | 248.6 ± 9.4 | 0.11 μS cm ⁻¹ | - | 55 | Food waste + Sewage sludge | 16S rRNA sequencing | DeFluviitoga Tepidimicrobium | Methanothermobacter Methanosarcina | [49] |

NA: not available; CLSM: confocal laser scanning microscopy; FISH: Fluorescence in situ hybridization; OFMSW: organic fraction of municipal solid waste; SSA: surface area; WAS: waste activated sludge.

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4.4. Effect of biochar on digestate quality

Anaerobic digestate has been considered as soil improver because it is rich in nutrients [184–186]. However, challenges related to digestate management have recently grown in association with EU regulations on ammonia, volatile organic acids, phenolic compounds, heavy metals, PAHs and PCBs [4,187]. So far, most technologies available to exploit digestate as soil improver are based on mechanical/physical (e.g. mechanical dewatering, semipermeable membranes and evaporation) or chemical processes (e.g. ammonia stripping and nutrients adjustment) (Table 8).

Table 8. Advantages and disadvantages of conventional technologies for digestate processing [188–191].

| Methods | Advantages | Disadvantages |
|--|--|--|
| Solid-liquid separation | Rich in phosphorus and suitable for fertilizer application | Liquid fraction has been considered as a surface and ground water pollutions |
| Belt and drum dryers | treatment and upgrading of digestate to a solid or concentrated product | Rapid volatilization of ammonium causes severe ammonia emission |
| High-Tech technologies (e.g. ammonia stripping, membrane process and vacuum evaporation exist) | Production of several streams with different physical and chemical properties. | Expensive |
| Mixing solid digestate with desired nutrient | Increasing value per ton | - |

Dehydrated digestate could be used in non-agricultural markets as a heating fuel, however, this use implies nutrient and metal losses, which in turn have negative impacts

on the environment and crops. Consequently, three approaches have been conventionally applied to reduce diffuse pollution resulting from digestate application to land [19,188]:

- Nutrient recovery from digestate;
- Carbon to Nitrogen ratio adjustment;
- Increasing of nutrient retention capacity using an additive.

One of the key aspects that should be considered when supplying additives to AD process is their effects on the quality of digestate for subsequent uses, especially as soil improver. Despite literature has not explored enough the fate and properties of digestate with BC to land applications, some potential benefits of BC amendment can be identified as follows. BC remaining in digestate after AD acts as a nutrient retention improver and catalyst, mitigates leaching of heavy metals and pollutants via physical and chemical absorption of organics, phosphate, ammonium, nitrate, nitrite, metals and CO₂ [19,192]. The improvements on digestate quality can be related to BC features as SSA, surface functional groups, ash content and presence of metals (Table 9).

Table 9. Biochar properties able to improve digestate quality

| Factors | Function | Literature | Reference |
|---|--|--|-----------|
| Textural properties (surface area, pore size) | Higher content of functional group (e.g. Si-O-Si, O-H and C=O) | HCl, NH ₃ ·H ₂ O and KMnO ₄ modification were performed to obtain functional biochar from Swine manure digestate. | [193] |
| | Increase of the pH and cation exchange capacity | H ₂ O ₂ and KOH modification were performed to obtain functional biochar from domestic sewage sludge digestate | [104] |
| | Ash content | Pinewood and white oak biochars made increase the alkalinity digestate a great alternative to agricultural lime fertilizer. | [33] |

| | | | |
|-----------------------|--|--|-------|
| Metals on the surface | The cation form of the metal can bind with soluble phosphorus | Ex: $3\text{Fe}(\text{OH})_2 + 2\text{H}_3\text{PO}_4^{3-} \rightarrow \text{Fe}_3(\text{PO}_4)_2 + 6\text{H}_2\text{O}$ | [151] |
| Functional group | Negatively charged functional groups forms complexes with heavy metals | Manganese oxide-modified biochar composite derived from corn straws improved heavy metal stabilization in the digestate | [194] |

BC supplement in digesters may be beneficial to the fertilizer value of digestates. Shen et al. [34,151] have found that BC addition can cause a substantial increase of the macro- and micro-nutrients as N, P, K, Ca, Mg, and Fe in digestate. Zhang et al. (2020) [195] reported a similar rise of nutrient content in digestate amended with BC, even if concentrations of certain nutrients did not fully meet limits of EU regulation on fertilizing products (Regulation EU 2019/1009). Research has shown that the joint amendment of BC and anaerobic digestate can reduce atmospheric greenhouses emissions from fields, such as N₂O [196] and CO₂ [197–199]. A mixture of dried anaerobic digestate and BC may be an alternative to standard formulations in horticultural potting media [200]. In addition, BC may contribute to improve the composting of digestate [153,201,202], particularly in terms of process performance, compost quality and its benefits on plants [203–205].

Further research is needed to understand interactions between BC, digestate and soil for the potential use of the mixture as soil improver after AD. Future studies about the use of BC as additive in AD should also consider its effects on the agronomic value of anaerobic sludge (i.e. content in macro- and micro-nutrients, germination and phytotoxicity tests, and others [206]).

4.5. Effects on biogas upgrading

Raw biogas from AD consists mainly of CH₄ (50-70 % v/v) and CO₂ (30-50 % v/v), along with minor compounds as water vapour, H₂S, NH₃, O₂ and N₂ [207]. Upgrading and cleaning biogas are required to meet the requirements and standards for engines and pipelines, although they represent energetic and economic costs up to 55 % of the total biomethane production cost [33,208,209]. To date, conventional technologies involve water scrubbing, cryogenic separation, physico-chemical absorption, and membranes, among the others [210,211]. Recently, BC has been investigated as adsorbent of CO₂ and H₂S for in-situ and ex-situ applications, as addressed in the following paragraphs.

4.5.1. In-situ biogas upgrading

A series of studies [24,33,34,151] investigated the feasibility of in-situ biogas upgrading by the addition of BC, obtaining methane contents up to pipeline quality (Table 10). Shen et al. (2015) [151] investigated the possibility of sequestering CO₂ with BC during thermophilic AD of WAS for in-situ biogas cleaning and upgrading. They reported average methane contents of 88.5-96.7 % in BC amended reactors, compared to 67.9 % in control reactor, reaching CO₂ removals of 54.9-86.3 % and residual H₂S content below 5 ppb. They suggested that CO₂ removal could be promoted by the high porosity of BC, by the large SSA rich of basic sites and of hydrophobic sites. Shen et al. (2016) [33] assessed the AD of WAS with the addition of two woody BCs. They observed average methane content up to 92.3 % and 79.0 % in biogas from BCs amended reactors in mesophilic and thermophilic conditions compared to control reactors, corresponding to CO₂ removals by up to 66.2 % and 32.4 %, respectively. They stated that both BCs owned desirable properties for CO₂ sequestration, and in particular high values of: SSA, porosity,

chemical stability, degree of carbonization and alkaline nature. Linville et al. (2017) [24] investigated the influence of particle size and dose of BC from walnut shell on AD of FW in mesophilic and thermophilic conditions. They found higher CO₂ removals compared to control reactors in the case of smaller particle size of BC (61.0 %) than coarse one (51.0 %), due to the larger SSA and ash content. However, according to other studies [33,151], they observed a reduction of methane production with higher BC doses, concluding that this could lead to inhibition caused by higher concentrations of cations released by the BC. Shen et al. (2017) [34] studied the effects of two BCs from corn stover and pine wood on AD of WAS in two-stage digesters. They reported an average methane content of 81.0-88.6 % in the reactor with BC from corn stover and 72.1-76.6 % with BC from pine wood, compared to around 70.0 % for the control. They stated that BCs would release base cations sequestering CO₂ by chemical sorption and forming bicarbonate/carbonate salts, and that its surface structure would help CO₂ adsorption. Apart from CO₂ adsorption on BC, the major formation of CH₄ depends on the stronger syntrophic cooperation between organic acid-oxidising bacteria and CO₂ reducing methanogens [36,58], underlying the key role of efficient interspecies electron transfers. Further confirmation by other authors to the attractive findings about biogas in-situ upgrading by BC would be beneficial.

4.5.2. Ex-situ biogas cleaning and upgrading

The use of BC and other carbonaceous adsorbents for CO₂ capture from various gaseous streams recently attracted a growing attention [212–214]. Considering ex-situ applications of BC for biogas upgrading and cleaning, Table 11 shows CO₂ and H₂S adsorption capacity of different BCs. Clearly, the adsorption capacity for both CO₂ and H₂S is in a wide range, 0.4-2.3 mmol g⁻¹ and 0.2-19.1 mmol g⁻¹ respectively. Most of the

studies regarding CO₂ capture do not specifically focus on biogas, investigating different BCs eventually subjected to activation. Sethupathi et al. (2017) [215] assessed the adsorption of CH₄, CO₂ and H₂S in a synthetic biogas stream by four BCs in fixed bed adsorbers during continuous experiments. They reported that just CO₂ and H₂S were captured by BCs, which exhibited adsorption capacities up to 0.208 mmol g⁻¹ for H₂S and 0.126 mmol g⁻¹ for CO₂. Creamer et al. (2014) [216] investigated the adsorption of CO₂ into BCs from bagasse and hickory wood. They found that BC could effectively capture CO₂ (adsorption capacity up to 73.55 mg g⁻¹ or 1.67 mmol g⁻¹), suggesting the importance of surface area and superficial nitrogen groups in CO₂ sequestration, mainly through physical adsorption. Creamer and Gao (2016) [217] reported that the main mechanism for CO₂ sequestration by BC is physical adsorption, suggesting the importance of high SSA [100], adequate pore size (0.5-0.8 nm) [218] and pore volume, thanks to Van der Waals and electrostatic forces. However, the adsorption of CO₂ can also be influenced by BC chemical properties, such as the presence of basic surface functional groups or alkali and alkaline earth metals, hydrophobicity and non-polarity [214]. For instance, Xu et al. (2016) [219] found that the adsorption of CO₂ by three BCs during batch equilibrium tests was due to the presence of alkali and alkaline earth metals (Ca, Fe, K, Mg) by CO₂ mineralogical reactions together with physical sorption. Activation and surface treatments of BCs can provide high surface area and micropores for physical sorption and enrich surface functional groups and metal oxides for chemical sorption, leading to remarkable CO₂ adsorption capacities (5.0-7.4 mmol g⁻¹) [220].

Other studies applied BC for removing H₂S from biogas (Table 11). Sahota et al. 2018 [221] found 84.2 % removal of H₂S from biogas with BC from leaf waste. Kanjanarong et al. (2017) [222] obtained a removal of 98 % of H₂S (8.02 mmol g⁻¹) from biogas with

BC, suggesting carboxylic and hydroxide radical groups as responsible of H₂S adsorption. Finally, Pelaez-Samaniego et al. (2018) [37] found that BC from AD digestate could effectively remove H₂S from a synthetic biogas, possibly facilitated by the presence of ash, porosity, or aromatics in BC. In contrast with CO₂ for which adsorption onto BC seemed to be mainly physical, absorption of H₂S seemed to involve many chemical mechanisms with BC surface [223].

Overall, biochar seems to be a promising adsorbent for ex-situ biogas cleaning and upgrading applications. However, additional studies [215] should focus on the adsorption of CO₂ and H₂S along with NH₃ from real or synthetic biogas, considering their competitive adsorptions, along with the influence of water vapour and the eventual removal of CH₄.

Table 10. Physic-chemical properties of biochars used in biogas in-situ upgrading

| Feedstock | Production temperature [°C] | BET-SA [m ² g ⁻¹] | Total pore volume [cm ³ g ⁻¹] | Average pore diameter [nm] | H/C [molar ratio] | O/C [molar ratio] | Ash [% wt] | Reference |
|-------------------|-----------------------------|--|--|----------------------------|-------------------|-------------------|--------------|-----------|
| Corn stover | Gasification | 315.30 | 0.09 | 6.50 | 0.075 ± 0.007 | 0.004 ± 0.001 | 45.18 ± 0.40 | [34,151] |
| Pine pellets | Gasification | 310.19 | 0.19 | 5.07 | 0.078 ± 0.009 | 0.249 ± 0.014 | 18.69 ± 0.44 | [33,34] |
| White oak pellets | Gasification | 296.81 | 0.15 | 4.92 | 0.109 ± 0.026 | 0.051 ± 0.010 | 34.90 ± 0.65 | |
| Walnut shell | Gasification: 900 °C | 86.5 | 0.16 | 7.06 | 0.20 ± 0.01 | 0.06 ± 0.01 | 43.2 ± 0.2 | [24] |

Table 11. Biochar adsorption capacity (mmol g⁻¹) of H₂S and CO₂ during ex-situ applications.

| Feedstock | Pyrolysis/activation | Surface area [m ² g ⁻¹] | Total pore volume [cm ³ g ⁻¹] | Gas | Inlet H ₂ S or CO ₂ concentration [ppm] | H ₂ S Adsorption capacity [mmol g ⁻¹] | CO ₂ Adsorption capacity [mmol g ⁻¹] | Reference |
|-----------|----------------------|--|--|-----|---|--|---|-----------|
|-----------|----------------------|--|--|-----|---|--|---|-----------|

| | | | | | | | |
|------------------------------------|---|-------|-------|-----------------------------------|----------|--------------------|-------------------------|
| AD digestate fibres | PY/500°C/60 min | 134 | 0.037 | Synthetic biogas | 2000 | 3.96 ^a | [37] |
| | PY/600°C/60min | 142 | 0.035 | Synthetic biogas | 2000 | 15.90 ^a | |
| | PY/600°C/60min +Na ₂ CO ₃ | - | - | Synthetic biogas | 2000 | 19.13 ^a | |
| Biomass from black liquor | C/450°C/6min | 60 | | N ₂ +H ₂ S | 1000 | 2.14 ^a | [224] |
| Pig manure | PY/500°C/4hours | 47.4 | | Air + H ₂ S | 1% (v/v) | 1.75 ^a | [225] |
| Sewage sludge | PY/500°C/4hours | 71.6 | | Air + H ₂ S | 1% (v/v) | 1.29 ^a | |
| Potato peel waste | C/500°C/5min | 63 | | N ₂ + H ₂ S | 1000 | 1.56 ^a | [226] |
| Camphor | PY/400°C/5hours | 20 | | | | 3.21 ^a | [227] |
| Rice hull | PY/400°C/5hours | 115 | | | | 11.23 ^a | |
| Bamboo | PY/400°C/5hours | 58 | | | | 9.88 ^a | |
| 80% wood chips 20% AD digestate | PY/600°C | | | Biogas | 1020 | 8.02 ^a | [222] |
| Perilla leaf | PY/700°C | 473.4 | 0.1 | Synthetic biogas | | 0.537 | 2.312 [215] |
| Korean oak | PY/400°C | 270.8 | 0.1 | Synthetic biogas | | 0.178 | 0.597 |
| Japanese oak | PY/500°C | 475.6 | 0.2 | Synthetic biogas | | 0.167 | 0.379 |
| Soybean stover | PY/700°C | 420.3 | 0.2 | Synthetic biogas | | 0.308 | 0.707 |
| Sawdust | G/850°C | 182.0 | 0.003 | N ₂ /CO ₂ | | | 1.08 ^b [228] |
| | G/850°C + monoethanolamine | 3.17 | 0.007 | N ₂ /CO ₂ | | | 1.02 ^b |
| Sugarcane bagasse | PY/600°C | 401.0 | | | | | 1.67 ^b [216] |
| Whitewood | PY/500°C + steam activation | 840 | 0.55 | He/CO ₂ | 30 % mol | | 1.34 ^b [229] |
| Whitewood | PY/500°C + CO ₂ activation | 820 | 0.45 | He/CO ₂ | 30 % mol | | 1.43 ^b |
| Whitewood | PY/500°C + KOH activation | 1400 | 0.62 | He/CO ₂ | 30 % mol | | 1.77 ^b |
| Walnut shell | C900°C/1.5hours | 397 | 0.198 | | | | 1.65 ^b [230] |
| Pig manure | PY/500°C/4hours | 31.57 | 0.044 | N ₂ /CO ₂ | | | 0.53 ^b [219] |
| Wheat straw | PY/500°C/4hours | 20.20 | 0.041 | N ₂ /CO ₂ | | | 0.78 ^b |
| Sewage sludge | PY/500°C/4hours | 10.12 | 0.022 | N ₂ /CO ₂ | | | 0.41 ^b |

5. Economic and environmental assessments

An economic and environmental evaluation of BC production and application in AD was performed, considering specifically four perspectives: 1. BC production according to feedstock composition and 2. to pyrolysis process conditions; 3. BC application in AD compared to current state of the art technologies addressing AD challenges; 4. integration of AD and pyrolysis processes (to our knowledge, specific studies related to the economic and environmental assessment of BC use in AD processes are not available).

5.1. Economic assessment

Considering BC production, the key parameters to evaluate the economic benefits of feedstocks (perspective 1) are: ash and lignin contents and O/C ratio, which affect pyrolysis yield, molecular weight of bio-oil and BC amount production [231]. Li et al. (2017) [232], based on regression model applied to 346 lignocellulosic feedstocks, stated that higher ash content increases BC production in a range of 12.5-15.5 %, reducing bio-oil production, and consequently the incomes coming out from bio-oil trade, which makes pyrolysis economically profitable. With lower ash content and higher O/C ratio of feedstocks, higher biofuel yields are produced, which leads to better economic performance, in fact minimum fuel selling prices for lignocellulosic feedstocks ranged from 0.53-1.1 Euro/L.

Considering the pyrolysis process (perspective 2), the investment costs for BC production from lignocellulosic feedstocks are: 43 % for pre-treatment and pyrolysis, 35 % for H₂ generation and 22 % for cooling and fractionations [232]. The average operating cost varies from 0.68 Euro/L for woody biomass to 0.86 Euro/L for straw biomass, due to the higher costs of disposal and pre-treatment of straw biomass, respectively 32 % and 34 %

of total operational costs [233]. Based on the economic analysis performed by Harsono et al. (2013) [234] and Sahoo et al. (2019) [235], the investment and operational costs related to BC production from lignocellulosic feedstock can only be balanced by a BC trade price of 470 Euro/t.

Considering perspective 3, the need to enhance AD feasibility and applicability to unconventional substrates implied higher costs, which should be exceeded by the increased methane production and therefore by the additional electric energy potentially associated. Inorganic and biological additives as iron, micronutrients and ash are conventionally employed to reduce inhibition and facilitate organic matter solubilisation, thus improving methane production. Nevertheless, the application of additives in AD accounts as 3.60-4.10 euro/L of enzyme and as 13-16 euro/L of nutrients [236]. The overall costs of BC, depending on feedstock, pyrolysis process and activating agent (Table 12), could range from 0.2 to 0.5 USD/kg, making BC cheaper than granular activated carbon (GAC), which has production costs ranging between 0.6 and 20 USD/kg.

Table 12. Comparison of granular activated carbon (GAC) and biochar (BC) production costs depending on feedstock, production process and activating agent

| Additive | Feedstock | Production process/ Activating agent | Location | Cost of production (USD kg ⁻¹) | Reference |
|------------------|-------------------------------|--------------------------------------|----------|--|-----------|
| GAC | Poultry litter derived carbon | Steam | USA | 1.44 | [237] |
| GAC | Rice bran | NaOH | China | 3.58-3.77 | [237] |
| GAC | Red oak wood | Steam | USA | 0.62-1.27 | [237] |
| GAC - commercial | - | - | USA | 1.19-16.34 | [237] |
| GAC | Fruit processing waste | Steam | Malaysia | 1.67 | [237] |
| GAC | Rice bran | CO ₂ | Brazil | 3.54 | [237] |

| | | | | | |
|------------------|--|------------------------|-----------------------------|--------------------------|-------|
| GAC | Acid treated rice bran | CO ₂ | Brazil | 20.45 | [237] |
| GAC | Wood | KOH | - | 2.49 | [237] |
| GAC | Charcoal | KOH | - | 1.25 | [237] |
| GAC | Lignite | KOH | - | 2.18 | [237] |
| GAC - commercial | | | | 1.93 | [237] |
| BC | Empty fruit bunches | Slow PY | Malaysia | 0.533 | [234] |
| BC | Straw | Slow PY (large scale) | UK | 0.203 | [18] |
| BC | Straw | Slow PY (medium scale) | UK | 0.447 | [18] |
| BC | Straw | Slow PY (small scale) | UK | 0.351 | [18] |
| BC | Short rotation coppicing, forestry residues, short rotation forestry | Slow PY (large scale) | UK | 0.266 | [18] |
| BC | Short rotation coppicing, forestry residues, short rotation forestry | Slow PY (medium scale) | UK | 0.500 | [18] |
| BC | Short rotation coppicing | Slow PY (small scale) | UK | 0.434 | [18] |
| BC | Forestry residue | Slow PY (large scale) | UK | 0.345 | [18] |
| BC | Forestry residue | Slow PY (medium scale) | UK | 0.584 | [18] |
| BC | Arboricultural arisings | Slow PY (small scale) | UK | 0.213 | [18] |
| BC | Pine wood | Slow PY | USA | 0.220-0.280 | [238] |
| BC | - | - | European market (2014) | 0.662-0.811 ^a | [239] |
| BC | - | - | - | 0.207 | [239] |
| BC | - | - | - | 0.600 | [68] |
| BC | - | - | Global, market price (mean) | 2.06 | [240] |
| BC | - | - | USA, market price (mean) | 2.48 (0.08-13.48) | [241] |

GAC: granular activated carbon; PY: pyrolysis.

(a) exchange rate USD/euro (January 2014): 1.3516

Still considering perspective 3 and moving from the additive's cost to the improvement of AD performances, The economic benefits of the integration of AD and pyrolysis technologies (perspective 4) has been explored by literature. The integrated technologies of AD of waste biomass and pyrolysis of the digestate could increase the net electricity production respect to AD alone [80] and enhance its quality as soil amendment [81] with economic and environmental benefits [242]. However, up to date, there is uncertainty regarding the balance between input costs of BC supplementation and output of energy production from AD. Qiu et al. (2019) [59] proposed the energy input-output LCA method to estimate the total energy input for biogas production, by calculating the energy associated with each component used in AD, and the actual energy return of investment. Zhang et al. (2020) [243] investigated the addition of woody BC to improve the thermophilic AD of FW. They concluded that BC supplementation could be economically feasible to enhance thermophilic AD of FW.

5.2. Environmental assessment

Considering feedstocks (perspective 1), life cycle analysis (LCA) from cradle to cradle of the pyrolysis of lignocellulosic feedstocks stated that GHG emissions for husk/shell/pit ranged from 120-250 g CO₂eq/MJ, while for other organic waste, wood and straw they ranged between 20-50 g CO₂eq/MJ [232]. This difference was due to the dominant contribution of indirect land use change from food production. Feedstocks having higher O/C ratio and 0 – 2 % ash content increased the GHG emissions [244]. Based on GREET database [245], GHG emissions reduction for lignocellulosic feedstocks was 85 - 98 % compared to the GHG emissions for petroleum fuels refining, which was equal to 93

gCO₂eq/MJ. GHG emission reductions for lignocellulosic feedstocks satisfy the 50 % share of renewable fuel standard for GHG emission reduction requirement for advanced fuels [246].

Considering BC production process (perspective 2), LCA from cradle to gate showed that BC produced from palm oil empty fruit bunches through slow pyrolysis had an energy content higher than the energy required for producing BC [234]. Furthermore, LCA cradle-to grave proved the positive energy balance of BC produced from different lignocellulosic materials [247]. Hence, the positive energy balance of BC production and application, due to the high-energy content of lignocellulosic feedstocks, represents a crucial benefit both from economic and environmental perspectives. One of the main issue of BC produced from renewable feedstocks as lignocellulose wastes (i.e. WAS, wood) and not from purpose grown feedstocks is the higher risk of having contaminants as heavy metals or organic compounds like dioxins, PAHs and PCBs [248].

Still considering BC process production (perspective 2), pyrolysis and gasification were compared from the perspective of carbon equivalent abatement (CA): pyrolysis achieved the highest CA, ranging from 0.07 to 1.25 t CO₂ eq/t feedstock, as cardboard and wood waste, while gasification reached the highest electricity generation outputs with 0.9 MWhe/t of feedstocks [249]. Activation was proven to represent a high item cost both in terms of economic and environmental perspectives [250]. BC chemical activation costs, in line with non-renewable GAC activation, range between 1.38 and 1.48 Euro/kg, respectively with acidic and basic treatments [251].

However, perspective 2 could also be explored considering the existing literature related to the conventional applications of BC as soil improver, adsorbent for water and air pollutants, catalyst for syngas upgrading and biodiesel production (Table 13). LCA

studies from cradle-to-grave of conventional BC applications measured in all cases positive environmental benefits compared to conventional perspectives. BC produced from waste forestry feedstock and applied as soil improver contributed to GHGs emission reduction up to 2.74 kg CO₂ eq/ kg BC for the impact categories climate change (CC), natural gas avoided for fossil depletion (FD) and urea avoided for freshwater eutrophication (FE) and human toxicity (HT) [252]. A LCA from cradle to cradle [247] showed that BC production and application as adsorbent presented, compared to GAC deriving from virgin non-renewable feedstock, lower environmental impacts in terms of climate change (CC), fossil depletion (FD), freshwater eutrophication (FE) and terrestrial acidification (TA) impact categories, and that wood and wood chips achieved the highest environmental credits among the others feedstocks. In details, considering CC (expressed as kg CO₂/kg adsorption material), GAC produced 1.44, while wood chips and corn stover had credits ranging between -3.42 to -3.57, whereas oil palm produced 11.1 [253]. Regarding FE (evaluated as kg P eq/ kg adsorption material), BC from lignocellulosic feedstock exhibited higher values than GAC, respectively ranging between 6.2 to 10.9 % [247], due to the organic matter of renewable feedstock [254]. Considering both TA and FD (respectively estimated as kg SO₂ eq/ kg adsorption material and kg oil eq/ kg adsorption material), BC from lignocellulosic feedstock achieved higher credits than GAC, between 9.5 % and 32.0 %.

Considering BC application in AD processes (perspective 3), to our knowledge the available scientific literature only focused on sequential processes, as pyrolysis followed by AD. A LCA from cradle to grave [255] considered BC produced from corn stover applied as carburant and soil amendment with sequential AD, achieving respectively: -

2.47 kg CO₂eq/t and energy saving of -6.53 MJ/ t for the first scenario and - 4.67 kg CO₂eq/t and energy saving of -9.73 MJ/ t for the second scenario.

Table 13. Environmental assessment of biochar (BC) production and uses

| | Positive effects | Negative effects |
|------------------------------------|---|--|
| Feedstock (perspective 1) | Lignocellulosic biomass exhibited more positive effects because of higher energy potential [247] Waste biomasses presented more advantages (e.g. avoided waste management) [256] | Feedstock provision (transport) [93] Potential presence of contaminants (heavy metals, persistent organic pollutants) in waste feedstocks (sewage sludge) [256] |
| Production process (perspective 2) | Renewable energy from syngas and bio-oil [93] Stabilization of C in biomass feedstock [18] | Large centralized pyrolysis units higher transportation distances compared to smaller decentralized units [256] |
| Activation process (perspective 2) | | Activation was proven to represent a high item cost both in terms of economic and environmental perspectives [250]. Use of chemicals and electricity is associated to higher impacts [257]. |
| Applications | | |
| Soil improver | Biochar (C) sequestration in the soil [93] Reduced fertiliser requirements [93] Reduced N ₂ O emissions from soil [93] Enhanced plants growth [18] Reduced fossil fuel use in irrigation and cultivation [18] Enhanced nutrient availability [258] Increased H ₂ O retention [258] Reduced leaching and run-off of nutrients [258] | Binding /deactivation of pesticides, herbicides and nutrients in soil [258] BC as source of potential toxicants (heavy metals, PAHs, organics) [258] |
| Adsorbent | BC lower impacts than activated carbon [247] | |

In conclusion, focusing the environmental assessment on perspective 4, the integration of AD of waste biomass and pyrolysis of digestate, as shown in Figure 2, could increase the

net electricity production respect to AD alone [80] and enhance its quality as soil amendment [81] with economic and environmental benefits [242]. In conclusion, the integrated approach has been investigated by life cycle analysis and exhibited positive environmental outcomes if compared with non-integrated processes [255,259].

6. Conclusions

This review addressed three key issues related to the comprehension of BC role in AD processes:

1. Investigation of the influence of BC properties on AD performances and of their ability to counteract its main challenges. It is understood that BC properties are determined by the feedstock and by pyrolysis and activation processes. The key features were SSA, porous structure and distribution, nature of surface functional groups (related to CEC and adsorption capacity, buffer capacity, ability to immobilize microbial communities), elemental composition and ash content. However, some mechanisms (e.g. BC role in ammonium adsorption and BC influence on microbial mechanisms) still need to be fully understood and explained. Other challenges for future research are related to digestate management; in detail, the influence of BC relatively high doses on the rheological properties of the digestate should be explored, as well as the eventual leaching of pollutants in the environment as a consequence of digestate recovery as soil improver.

2. Assessment of the optimal BC production chain (i.e. feedstock-pyrolysis-activation) to achieve the desired features. Lignocellulosic biomasses, slow pyrolysis and physical activation seemed to be a good combination in general, while other feedstocks and/or chemical activation should be evaluated for specific needs and tailor-made applications.

However, a systematic investigation of the correlations linking BC physico-chemical characteristics and AD performances, carefully exploring one by one the BC effects mentioned in this review, is highly needed for a deep understanding of BC role as additive in AD processes.

3. Evaluation of the economic and environmental advantages connected to BC use in AD processes, compared to conventional solutions applied to address AD challenges. The main research gap related to this issue is the absence of specific literature related to BC use in AD processes, therefore only general statements could be formulated. The use of BC as additive could be cheaper and has less environmental impacts than of conventional AD improvers (e.g. physico-chemical-biological pre-treatments) and of GAC. The integration of AD and pyrolysis achieved economic feasibility and positive environmental performances if compared with non-integrated processes. Future research could investigate the optimization of technical, economic and environmental performances of BC production chain and its integration in AD processes.

Acknowledgements

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors. The authors declare no conflict of interest. Authors' contributions: conceptualization: Silvia Fiore and Marco Chiappero; methodology and supervision: Silvia Fiore; writing of original draft: all authors; review and editing of the manuscript: all authors.

References

[1] Enerdata. World Energy Consumption Statistics. Glob Energy Stat Yearb 2019.

- 1016 <https://yearbook.enerdata.net/total-energy/world-consumption-statistics.html>
1017 (accessed April 28, 2020).
- 1018 [2] International Energy Agency (IEA). Statistics | World - Total Primary Energy
1019 Supply (TPES) by source 2018.
1020 <https://www.iea.org/statistics/?country=WORLD&year=2016&category=Energy>
1021 [supply&indicator=TPESbySource&mode=chart&dataTable=BALANCES](https://www.iea.org/statistics/?country=WORLD&year=2016&category=Energy)
1022 (accessed February 25, 2019).
- 1023 [3] Weiland P. Biogas production: current state and perspectives. *Appl Microbiol*
1024 *Biotechnol* 2010;85:849–60. doi:10.1007/s00253-009-2246-7.
- 1025 [4] Nkoa R. Agricultural benefits and environmental risks of soil fertilization with
1026 anaerobic digestates: a review. *Agron Sustain Dev* 2014;34:473–92.
1027 doi:10.1007/s13593-013-0196-z.
- 1028 [5] Walsh JJ, Jones DL, Edwards-Jones G, Williams AP. Replacing inorganic
1029 fertilizer with anaerobic digestate may maintain agricultural productivity at less
1030 environmental cost. *J Plant Nutr Soil Sci* 2012;175:840–5.
1031 doi:10.1002/jpln.201200214.
- 1032 [6] Ward AJ, Hobbs PJ, Holliman PJ, Jones DL. Optimisation of the anaerobic
1033 digestion of agricultural resources. *Bioresour Technol* 2008;99:7928–40.
1034 doi:10.1016/J.BIORTECH.2008.02.044.
- 1035 [7] Appels L, Lauwers J, Degève J, Helsen L, Lievens B, Willems K, et al.
1036 Anaerobic digestion in global bio-energy production: Potential and research
1037 challenges. *Renew Sustain Energy Rev* 2011;15:4295–301.
1038 doi:10.1016/J.RSER.2011.07.121.
- 1039 [8] European Biogas Association (EBA). Statistical report, 2019 2019.

1040 <https://www.europeanbiogas.eu/eba-statistical-report-2019> (accessed May 8,
1041 2020).

1042 [9] Mata-Alvarez J, Dosta J, Romero-Güiza MS, Fonoll X, Peces M, Astals S. A
1043 critical review on anaerobic co-digestion achievements between 2010 and 2013.
1044 *Renew Sustain Energy Rev* 2014;36:412–27. doi:10.1016/J.RSER.2014.04.039.

1045 [10] Taherzadeh M, Karimi K, Taherzadeh MJ, Karimi K. Pretreatment of
1046 Lignocellulosic Wastes to Improve Ethanol and Biogas Production: A Review.
1047 *Int J Mol Sci* 2008;9:1621–51. doi:10.3390/ijms9091621.

1048 [11] Carrere H, Antonopoulou G, Affes R, Passos F, Battimelli A, Lyberatos G, et al.
1049 Review of feedstock pretreatment strategies for improved anaerobic digestion:
1050 From lab-scale research to full-scale application. *Bioresour Technol*
1051 2016;199:386–97. doi:10.1016/j.biortech.2015.09.007.

1052 [12] Arif S, Liaquat R, Adil M. Applications of materials as additives in anaerobic
1053 digestion technology. *Renew Sustain Energy Rev* 2018;97:354–66.
1054 doi:10.1016/j.rser.2018.08.039.

1055 [13] Ye M, Liu J, Ma C, Li Y-Y, Zou L, Qian G, et al. Improving the stability and
1056 efficiency of anaerobic digestion of food waste using additives: A critical review.
1057 *J Clean Prod* 2018;192:316–26. doi:10.1016/J.JCLEPRO.2018.04.244.

1058 [14] Romero-Güiza MS, Vila J, Mata-Alvarez J, Chimenos JM, Astals S. The role of
1059 additives on anaerobic digestion: A review. *Renew Sustain Energy Rev*
1060 2016;58:1486–99. doi:10.1016/j.rser.2015.12.094.

1061 [15] González J, Sánchez M, Gómez X. Enhancing Anaerobic Digestion: The Effect
1062 of Carbon Conductive Materials. *J Carbon Res* 2018;4:59.
1063 doi:10.3390/c4040059.

- 1064 [16] Zhang J, Zhao W, Zhang H, Wang Z, Fan C, Zang L. Recent achievements in
1065 enhancing anaerobic digestion with carbon- based functional materials. *Bioresour*
1066 *Technol* 2018;266:555–67. doi:10.1016/j.biortech.2018.07.076.
- 1067 [17] Nanda S, Dalai AK, Berruti F, Kozinski JA. Biochar as an Exceptional
1068 Bioresource for Energy, Agronomy, Carbon Sequestration, Activated Carbon and
1069 Specialty Materials. *Waste and Biomass Valorization* 2016;7:201–35.
1070 doi:10.1007/s12649-015-9459-z.
- 1071 [18] Lehmann J, Joseph S. Biochar for environmental management: science,
1072 technology and implementation. II. Earthscan from Routledge; 2015.
- 1073 [19] Fagbohunge MO, Herbert BMJ, Hurst L, Ibeto CN, Li H, Usmani SQ, et al. The
1074 challenges of anaerobic digestion and the role of biochar in optimizing anaerobic
1075 digestion. *Waste Manag* 2017;61:236–49. doi:10.1016/j.wasman.2016.11.028.
- 1076 [20] Pecchi M, Baratieri M. Coupling anaerobic digestion with gasification, pyrolysis
1077 or hydrothermal carbonization: A review. *Renew Sustain Energy Rev* 2019.
1078 doi:10.1016/j.rser.2019.02.003.
- 1079 [21] Fabbri D, Torri C. Linking pyrolysis and anaerobic digestion (Py-AD) for the
1080 conversion of lignocellulosic biomass. *Curr Opin Biotechnol* 2016;38:167–73.
1081 doi:10.1016/j.copbio.2016.02.004.
- 1082 [22] Hübner T, Mumme J. Integration of pyrolysis and anaerobic digestion - Use of
1083 aqueous liquor from digestate pyrolysis for biogas production. *Bioresour Technol*
1084 2015;183:86–92. doi:10.1016/j.biortech.2015.02.037.
- 1085 [23] Salman CA, Schwede S, Thorin E, Yan J. Enhancing biomethane production by
1086 integrating pyrolysis and anaerobic digestion processes. *Appl Energy*
1087 2017;204:1074–83. doi:10.1016/j.apenergy.2017.05.006.

- 1088 [24] Linville JL, Shen Y, Ignacio-de Leon PA, Schoene RP, Urgun-Demirtas M. In-
 1089 situ biogas upgrading during anaerobic digestion of food waste amended with
 1090 walnut shell biochar at bench scale. *Waste Manag Res* 2017;35:669–79.
 1091 doi:10.1177/0734242X17704716.
- 1092 [25] Torri C, Fabbri D. Biochar enables anaerobic digestion of aqueous phase from
 1093 intermediate pyrolysis of biomass. *Bioresour Technol* 2014;172:335–41.
 1094 doi:10.1016/j.biortech.2014.09.021.
- 1095 [26] Sunyoto NMS, Zhu M, Zhang Z, Zhang D. Effect of biochar addition on
 1096 hydrogen and methane production in two-phase anaerobic digestion of aqueous
 1097 carbohydrates food waste. *Bioresour Technol* 2016;219:29–36.
 1098 doi:10.1016/j.biortech.2016.07.089.
- 1099 [27] Jang HM, Choi YK, Kan E. Effects of dairy manure-derived biochar on
 1100 psychrophilic, mesophilic and thermophilic anaerobic digestions of dairy manure.
 1101 *Bioresour Technol* 2018;250:927–31. doi:10.1016/j.biortech.2017.11.074.
- 1102 [28] Sun C, Liu F, Song Z, Wang J, Li Y, Pan Y, et al. Feasibility of dry anaerobic
 1103 digestion of beer lees for methane production and biochar enhanced performance
 1104 at mesophilic and thermophilic temperature. *Bioresour Technol* 2019;276:65–73.
 1105 doi:10.1016/J.BIORTECH.2018.12.105.
- 1106 [29] Zhao Z, Zhang Y, Woodard TL, Nevin KP, Lovley DR. Enhancing syntrophic
 1107 metabolism in up-flow anaerobic sludge blanket reactors with conductive carbon
 1108 materials. *Bioresour Technol* 2015;191:140–5.
 1109 doi:10.1016/j.biortech.2015.05.007.
- 1110 [30] Inthapanya S, Preston TR, Leng RA. Biochar increases biogas production in a
 1111 batch digester charged with cattle manure. *Livest Res Rural Dev* 2012;24.

- 1112 [31] Inthapanya S, Preston TR. Biochar marginally increases biogas production but
1113 decreases methane content of the gas in continuous-flow biodigesters charged
1114 with cattle manure. *Livest Res Rural Dev* 2013;25.
- 1115 [32] Bruun EW, Müller-Stöver D, Ambus P, Hauggaard-Nielsen H. Application of
1116 biochar to soil and N₂O emissions: potential effects of blending fast-pyrolysis
1117 biochar with anaerobically digested slurry. *Eur J Soil Sci* 2011;62:581–9.
1118 doi:10.1111/j.1365-2389.2011.01377.x.
- 1119 [33] Shen Y, Linville JL, Ignacio-de Leon PAA, Schoene RP, Urgun-Demirtas M.
1120 Towards a sustainable paradigm of waste-to-energy process: Enhanced anaerobic
1121 digestion of sludge with woody biochar. *J Clean Prod* 2016;135:1054–64.
1122 doi:10.1016/j.jclepro.2016.06.144.
- 1123 [34] Shen Y, Forrester S, Koval J, Urgun-Demirtas M. Yearlong semi-continuous
1124 operation of thermophilic two-stage anaerobic digesters amended with biochar
1125 for enhanced biomethane production. *J Clean Prod* 2017;167:863–74.
1126 doi:10.1016/j.jclepro.2017.05.135.
- 1127 [35] Codignole Luz F, Cordiner S, Manni A, Mulone V, Rocco V. Biochar
1128 characteristics and early applications in anaerobic digestion-a review. *J Environ*
1129 *Chem Eng* 2018;6:2892–909. doi:10.1016/j.jece.2018.04.015.
- 1130 [36] Masebinu SO, Akinlabi ET, Muzenda E, Aboyade AO. A review of biochar
1131 properties and their roles in mitigating challenges with anaerobic digestion.
1132 *Renew Sustain Energy Rev* 2019;103:291–307.
1133 doi:10.1016/J.RSER.2018.12.048.
- 1134 [37] Pelaez-Samaniego MR, Smith MW, Zhao Q, Garcia-Perez T, Frear C, Garcia-
1135 Perez M. Charcoal from anaerobically digested dairy fiber for removal of

1136 hydrogen sulfide within biogas. *Waste Manag* 2018;76:374–82.
 1137 doi:10.1016/j.wasman.2018.03.011.

1138 [38] Wang G, Li Q, Gao X, Wang XC. Synergetic promotion of syntrophic methane
 1139 production from anaerobic digestion of complex organic wastes by biochar:
 1140 Performance and associated mechanisms. *Bioresour Technol* 2018;250:812–20.
 1141 doi:10.1016/j.biortech.2017.12.004.

1142 [39] Wang D, Ai J, Shen F, Yang G, Zhang Y, Deng S, et al. Improving anaerobic
 1143 digestion of easy-acidification substrates by promoting buffering capacity using
 1144 biochar derived from vermicompost. *Bioresour Technol* 2017;227:286–96.
 1145 doi:10.1016/j.biortech.2016.12.060.

1146 [40] Fagbohunge MO, Herbert BMJ, Hurst L, Li H, Usmani SQ, Semple KT. Impact
 1147 of biochar on the anaerobic digestion of citrus peel waste. *Bioresour Technol*
 1148 2016;216:142–9. doi:10.1016/j.biortech.2016.04.106.

1149 [41] Mumme J, Srocke F, Heeg K, Werner M. Use of biochars in anaerobic digestion.
 1150 *Bioresour Technol* 2014;164:189–97. doi:10.1016/j.biortech.2014.05.008.

1151 [42] Shanmugam SR, Adhikari S, Nam H, Kar Sajib S. Effect of bio-char on methane
 1152 generation from glucose and aqueous phase of algae liquefaction using mixed
 1153 anaerobic cultures. *Biomass and Bioenergy* 2018;108:479–86.
 1154 doi:10.1016/j.biombioe.2017.10.034.

1155 [43] Su C, Zhao L, Liao L, Qin J, Lu Y, ... QD-J of C, et al. Application of biochar in
 1156 a CIC reactor to relieve ammonia nitrogen stress and promote microbial
 1157 community during food waste treatment. *J Clean Prod* 2019;209:353–62.
 1158 doi:10.1016/j.jclepro.2018.10.269.

1159 [44] Lü F, Luo C, Shao L, He P. Biochar alleviates combined stress of ammonium and

acids by firstly enriching *Methanosaeta* and then *Methanosarcina*. *Water Res* 2016;90:34–43. doi:10.1016/j.watres.2015.12.029.

[45] Cruz Viggi C, Simonetti S, Palma E, Pagliaccia P, Braguglia C, Fazi S, et al. Enhancing methane production from food waste fermentate using biochar: The added value of electrochemical testing in pre-selecting the most effective type of biochar. *Biotechnol Biofuels* 2017;10:1–13. doi:10.1186/s13068-017-0994-7.

[46] Li Q, Xu M, Wang G, Chen R, Qiao W, Wang X. Biochar assisted thermophilic co-digestion of food waste and waste activated sludge under high feedstock to seed sludge ratio in batch experiment. *Bioresour Technol* 2018;249:1009–16. doi:10.1016/j.biortech.2017.11.002.

[47] Martínez EJ, Rosas JG, Sotres A, Moran A, Cara J, Sánchez ME, et al. Codigestion of sludge and citrus peel wastes: Evaluating the effect of biochar addition on microbial communities. *Biochem Eng J* 2018;137:314–25. doi:10.1016/j.bej.2018.06.010.

[48] Luo C, Lü F, Shao L, He P. Application of eco-compatible biochar in anaerobic digestion to relieve acid stress and promote the selective colonization of functional microbes. *Water Res* 2015;68:710–8. doi:10.1016/j.watres.2014.10.052.

[49] Wang G, Li Q, Gao X, Wang XC. Sawdust-Derived Biochar Much Mitigates VFAs Accumulation and Improves Microbial Activities To Enhance Methane Production in Thermophilic Anaerobic Digestion. *ACS Sustain Chem Eng* 2019;acssuschemeng.8b04789. doi:10.1021/acssuschemeng.8b04789.

[50] Wang C, Liu Y, Gao X, Chen H, Xu X, Zhu L. Role of biochar in the granulation of anaerobic sludge and improvement of electron transfer characteristics.

1184 Bioresour Technol 2018;268:28–35. doi:10.1016/j.biortech.2018.07.116.

1185 [51] Zhao Z, Zhang Y, Holmes DE, Dang Y, Woodard TL, Nevin KP, et al. Potential
 1186 enhancement of direct interspecies electron transfer for syntrophic metabolism of
 1187 propionate and butyrate with biochar in up-flow anaerobic sludge blanket
 1188 reactors. Bioresour Technol 2016;209:148–56.
 1189 doi:10.1016/j.biortech.2016.03.005.

1190 [52] Yu HQ, Tay JH, Fang HHP. Effects of Added Powdered and Granular Activated
 1191 Carbons on Start-Up Performance of UASB Reactors. Environ Technol
 1192 1999;20:1095–101. doi:10.1080/09593332008616906.

1193 [53] Cooney MJ, Lewis K, Harris K, Zhang Q, Yan T. Start up performance of
 1194 biochar packed bed anaerobic digesters. J Water Process Eng 2016;9:e7–13.
 1195 doi:10.1016/j.jwpe.2014.12.004.

1196 [54] Dang Y, Sun D, Woodard TL, Wang LY, Nevin KP, Holmes DE. Stimulation of
 1197 the anaerobic digestion of the dry organic fraction of municipal solid waste
 1198 (OFMSW) with carbon-based conductive materials. Bioresour Technol
 1199 2017;238:30–8. doi:10.1016/j.biortech.2017.04.021.

1200 [55] Sasaki K, Morita M, Hirano S, Sasaki D, Ohmura N, Igarashi Y. Efficient
 1201 degradation of rice straw in the reactors packed by carbon fiber textiles. Appl
 1202 Microbiol Biotechnol 2010;87:1579–86. doi:10.1007/s00253-010-2667-3.

1203 [56] Martins G, Salvador AF, Pereira L, Alves MM. Methane Production and
 1204 Conductive Materials: A Critical Review. Environ Sci Technol 2018;52:10241–
 1205 53. doi:10.1021/acs.est.8b01913.

1206 [57] Chen S, Rotaru AE, Shrestha PM, Malvankar NS, Liu F, Fan W, et al. Promoting
 1207 interspecies electron transfer with biochar. Sci Rep 2014;4.

- doi:10.1038/srep05019.
- [58] Pan J, Ma J, Zhai L, Luo T, Mei Z, Liu H. Achievements of biochar application for enhanced anaerobic digestion: A review. *Bioresour Technol* 2019;292:122058. doi:10.1016/j.biortech.2019.122058.
- [59] Qiu L, Deng YF, Wang F, Davaritouchaee M, Yao YQ. A review on biochar-mediated anaerobic digestion with enhanced methane recovery. *Renew Sustain Energy Rev* 2019;115:109373. doi:10.1016/J.RSER.2019.109373.
- [60] Nanda S, Azargohar R, Kozinski JA, Dalai AK. Characteristic Studies on the Pyrolysis Products from Hydrolyzed Canadian Lignocellulosic Feedstocks. *BioEnergy Res* 2014;7:174–91. doi:10.1007/s12155-013-9359-7.
- [61] Azargohar R, Nanda S, Kozinski JA, Dalai AK, Sutarto R. Effects of temperature on the physicochemical characteristics of fast pyrolysis bio-chars derived from Canadian waste biomass. *Fuel* 2014;125:90–100. doi:10.1016/j.fuel.2014.01.083.
- [62] Azargohar R, Nanda S, Rao BVSK, Dalai AK. Slow Pyrolysis of Deoiled Canola Meal: Product Yields and Characterization. *Energy & Fuels* 2013;27:5268–79. doi:10.1021/ef400941a.
- [63] Mohanty P, Nanda S, Pant KK, Naik S, Kozinski JA, Dalai AK. Evaluation of the physiochemical development of biochars obtained from pyrolysis of wheat straw, timothy grass and pinewood: Effects of heating rate. *J Anal Appl Pyrolysis* 2013;104:485–93. doi:10.1016/j.jaap.2013.05.022.
- [64] Kan T, Strezov V, Evans TJ. Lignocellulosic biomass pyrolysis: A review of product properties and effects of pyrolysis parameters. *Renew Sustain Energy Rev* 2016;57:126–1140. doi:10.1016/j.rser.2015.12.185.
- [65] Garcia-Nunez JA, Pelaez-Samaniego MR, Garcia-Perez ME, Fonts I, Abrego J,

1232 Westerhof RJM, et al. Historical Developments of Pyrolysis Reactors: A Review.
 1233 Energy and Fuels 2017;31:5751–75. doi:10.1021/acs.energyfuels.7b00641.

1234 [66] Bridgwater A V. Biomass Conversion Technologies: Fast Pyrolysis Liquids from
 1235 Biomass: Quality and Upgrading, Springer, Cham; 2017, p. 55–98.
 1236 doi:10.1007/978-3-319-48288-0_3.

1237 [67] Meier D, Van De Beld B, Bridgwater A V., Elliott DC, Oasmaa A, Preto F.
 1238 State-of-the-art of fast pyrolysis in IEA bioenergy member countries. Renew
 1239 Sustain Energy Rev 2013;20:619–41. doi:10.1016/j.rser.2012.11.061.

1240 [68] Yoder J, Galinato S, Granatstein D, Garcia-Pérez M. Economic tradeoff between
 1241 biochar and bio-oil production via pyrolysis. Biomass and Bioenergy
 1242 2011;35:1851–62. doi:10.1016/j.biombioe.2011.01.026.

1243 [69] Brown RC. Thermochemical processing of biomass conversion into fuels,
 1244 chemicals and power. Wiley; 2011.

1245 [70] Hossain MM, Scott IM, Berruti F, Briens C. Application of Novel Pyrolysis
 1246 Reactor Technology to Concentrate Bio-oil Components with Antioxidant
 1247 Activity from Tobacco, Tomato and Coffee Ground Biomass. Waste and
 1248 Biomass Valorization 2018;9:1607–17. doi:10.1007/s12649-017-9943-8.

1249 [71] Li D, Berruti F, Briens C. Autothermal fast pyrolysis of woody residues and
 1250 wastes for the production of wood adhesives. In: M. Kuehle-Weidemeier and K.
 1251 Buescher Eds., editor. Waste-to-Resources 2017, Proc. VII Int. Symp. MBT
 1252 MRF, Hann. Ger., Cuvillier Verlag Gottingen; 2017, p. 215–26.

1253 [72] Tan X fei, Liu Y guo, Gu Y ling, Xu Y, Zeng G ming, Hu X jiang, et al. Biochar-
 1254 based nano-composites for the decontamination of wastewater: A review.
 1255 Bioresour Technol 2016;212:318–33. doi:10.1016/j.biortech.2016.04.093.

- 1256 [73] Shahkarami S, Dalai AK, Soltan J. Enhanced CO₂ Adsorption Using MgO-
1257 Impregnated Activated Carbon: Impact of Preparation Techniques. *Ind Eng*
1258 *Chem Res* 2016;55:5955–64. doi:10.1021/acs.iecr.5b04824.
- 1259 [74] Dehkhoda AM, Gyenge E, Ellis N. A novel method to tailor the porous structure
1260 of KOH-activated biochar and its application in capacitive deionization and
1261 energy storage. *Biomass and Bioenergy* 2016;87:107–21.
1262 doi:10.1016/j.biombioe.2016.02.023.
- 1263 [75] Briens C, Piskorz J, Berruti F. Biomass Valorization for Fuel and Chemicals
1264 Production -- A Review. *Int J Chem React Eng* 2008;6. doi:10.2202/1542-
1265 6580.1674.
- 1266 [76] Ekström C, Lindman N, Pettersson R. Catalytic Conversion of Tars, Carbon
1267 Black and Methane from Pyrolysis/Gasification of Biomass. *Fundam.*
1268 *Thermochem. Biomass Convers.*, Dordrecht: Springer Netherlands; 1985, p. 601–
1269 18. doi:10.1007/978-94-009-4932-4_34.
- 1270 [77] Lira CS, Berruti FM, Palmisano P, Berruti F, Briens C, Pécora AAB. Fast
1271 pyrolysis of Amazon tucumã (*Astrocaryum aculeatum*) seeds in a bubbling
1272 fluidized bed reactor. *J Anal Appl Pyrolysis* 2013;99:23–31.
1273 doi:10.1016/j.jaap.2012.11.005.
- 1274 [78] Suliman W, Harsh JB, Abu-Lail NI, Fortuna AM, Dallmeyer I, Garcia-Perez M.
1275 Modification of biochar surface by air oxidation: Role of pyrolysis temperature.
1276 *Biomass and Bioenergy* 2016;85:1–11. doi:10.1016/j.biombioe.2015.11.030.
- 1277 [79] Zhu L, Zhao N, Tong L, Lv Y, Li G. Characterization and evaluation of surface
1278 modified materials based on porous biochar and its adsorption properties for 2,4-
1279 dichlorophenoxyacetic acid. *Chemosphere* 2018;210:734–44.

doi:10.1016/j.chemosphere.2018.07.090.

[80] Monlau F, Sambusiti C, Antoniou N, Barakat A, Zabaniotou A. A new concept for enhancing energy recovery from agricultural residues by coupling anaerobic digestion and pyrolysis process. *Appl Energy* 2015;148:32–8. doi:10.1016/j.apenergy.2015.03.024.

[81] Monlau F, Francavilla M, Sambusiti C, Antoniou N, Solhy A, Libutti A, et al. Toward a functional integration of anaerobic digestion and pyrolysis for a sustainable resource management. Comparison between solid-digestate and its derived pyrochar as soil amendment. *Appl Energy* 2016;169:652–62. doi:10.1016/j.apenergy.2016.02.084.

[82] Pan J, Ma J, Liu X, Zhai L, Ouyang X, Liu H. Effects of different types of biochar on the anaerobic digestion of chicken manure. *Bioresour Technol* 2019;275:258–65. doi:10.1016/j.biortech.2018.12.068.

[83] Wen C. Anaerobic Digestion of Aqueous Pyrolysis Condensate. Electronic Thesis and Dissertation Repository, 2018.

[84] Yang Y, Heaven S, Venetsaneas N, Banks CJ, Bridgwater A V. Slow pyrolysis of organic fraction of municipal solid waste (OFMSW): Characterisation of products and screening of the aqueous liquid product for anaerobic digestion. *Appl Energy* 2018;213:158–68. doi:10.1016/j.apenergy.2018.01.018.

[85] Corton J, Donnison IS, Patel M, Böhle L, Hodgson E, Wachendorf M, et al. Expanding the biomass resource: Sustainable oil production via fast pyrolysis of low input high diversity biomass and the potential integration of thermochemical and biological conversion routes. *Appl Energy* 2016;177:852–62. doi:10.1016/j.apenergy.2016.05.088.

- 1304 [86] Elsamadony M, Tawfik A, Suzuki M. Surfactant-enhanced biohydrogen
1305 production from organic fraction of municipal solid waste (OFMSW) via dry
1306 anaerobic digestion. *Appl Energy* 2015;149:272–82.
1307 doi:10.1016/j.apenergy.2015.03.127.
- 1308 [87] Monlau F, Sambusiti C, Antoniou N, Zabaniotou A, Solhy A, Barakat A.
1309 Pyrochars from bioenergy residue as novel bio-adsorbents for lignocellulosic
1310 hydrolysate detoxification. *Bioresour Technol* 2015;187:379–86.
1311 doi:10.1016/j.biortech.2015.03.137.
- 1312 [88] Erdogan E, Atila B, Mumme J, Reza MT, Toptas A, Elibol M, et al.
1313 Characterization of products from hydrothermal carbonization of orange pomace
1314 including anaerobic digestibility of process liquor. *Bioresour Technol*
1315 2015;196:35–42. doi:10.1016/j.biortech.2015.06.115.
- 1316 [89] Dufour A, Castro-Diaz M, Brosse N, Bouroukba M, Snape C. The Origin of
1317 Molecular Mobility During Biomass Pyrolysis as Revealed by In situ ^1H NMR
1318 Spectroscopy. *ChemSusChem* 2012;5:1258–65. doi:10.1002/cssc.201100442.
- 1319 [90] Dufour A, Castro-Díaz M, Marchal P, Brosse N, Olcese R, Bouroukba M, et al.
1320 In Situ Analysis of Biomass Pyrolysis by High Temperature Rheology in
1321 Relations with ^1H NMR. *Energy & Fuels* 2012;26:6432–41.
1322 doi:10.1021/ef301310x.
- 1323 [91] Montoya J, Pecha B, Janna FC, Garcia-Perez M. Identification of the fractions
1324 responsible for morphology conservation in lignocellulosic pyrolysis:
1325 Visualization studies of sugarcane bagasse and its pseudo-components. *J Anal*
1326 *Appl Pyrolysis* 2017;123:307–18. doi:10.1016/j.jaap.2016.11.015.
- 1327 [92] Rangabhashiyam S, Balasubramanian P. The potential of lignocellulosic biomass

precursors for biochar production: Performance, mechanism and wastewater application—A review. *Ind Crops Prod* 2019;128:405–23. doi:10.1016/j.indcrop.2018.11.041.

[93] Meyer S, Glaser B, Quicker P. Technical, economical, and climate-related aspects of biochar production technologies: A literature review. *Environ Sci Technol* 2011;45:9473–83. doi:10.1021/es201792c.

[94] Rousset P, Aguiar C, Labbé N, Commandré JM. Enhancing the combustible properties of bamboo by torrefaction. *Bioresour Technol* 2011;102:8225–31. doi:10.1016/j.biortech.2011.05.093.

[95] Wilk M, Magdziarz A, Kalemba I. Characterisation of renewable fuels' torrefaction process with different instrumental techniques. *Energy* 2015;87:259–69. doi:10.1016/j.energy.2015.04.073.

[96] Kumar A, Saini K, Bhaskar T. Hydrochar and biochar: Production, physicochemical properties and techno-economic analysis. *Bioresour Technol* n.d. doi:10.1016/j.biortech.2020.123442.

[97] Román S, Nabais JM V, Laginhas C, Ledesma B, González JF. Hydrothermal carbonization as an effective way of densifying the energy content of biomass. *Fuel Process Technol* 2012;103:78–83. doi:10.1016/j.fuproc.2011.11.009.

[98] Sabio E, Álvarez-Murillo A, Román S, Ledesma B. Conversion of tomato-peel waste into solid fuel by hydrothermal carbonization: Influence of the processing variables. *Waste Manag* 2016;47:122–32. doi:10.1016/j.wasman.2015.04.016.

[99] Liu Z, Zhang FS, Wu J. Characterization and application of chars produced from pinewood pyrolysis and hydrothermal treatment. *Fuel* 2010;89:510–4. doi:10.1016/j.fuel.2009.08.042.

- 1352 [100] Hao W, Björkman E, Lilliestråle M, Hedin N. Activated carbons prepared from
 1353 hydrothermally carbonized waste biomass used as adsorbents for CO₂. Appl
 1354 Energy 2013;112:526–32. doi:10.1016/J.APENERGY.2013.02.028.
- 1355 [101] Cao X, Zhong L, Peng X, Sun S, Li S, Liu S, et al. Comparative study of the
 1356 pyrolysis of lignocellulose and its major components: Characterization and
 1357 overall distribution of their biochars and volatiles. Bioresour Technol
 1358 2014;155:21–7. doi:10.1016/j.biortech.2013.12.006.
- 1359 [102] Tan X fei, Liu S bo, Liu Y guo, Gu Y ling, Zeng G ming, Hu X jiang, et al.
 1360 Biochar as potential sustainable precursors for activated carbon production:
 1361 Multiple applications in environmental protection and energy storage. Bioresour
 1362 Technol 2017;227:359–72. doi:10.1016/j.biortech.2016.12.083.
- 1363 [103] Cha JS, Park SH, Jung SC, Ryu C, Jeon JK, Shin MC, et al. Production and
 1364 utilization of biochar: A review. J Ind Eng Chem 2016;40:1–15.
 1365 doi:10.1016/j.jiec.2016.06.002.
- 1366 [104] Wongrod S, Simon S, van Hullebusch ED, Lens PNL, Guibaud G. Changes of
 1367 sewage sludge digestate-derived biochar properties after chemical treatments and
 1368 influence on As(III and V) and Cd(II) sorption. Int Biodeterior Biodegrad
 1369 2018;135:96–102. doi:10.1016/j.ibiod.2018.10.001.
- 1370 [105] Dalai AK, Azargohar R. Production of Activated Carbon from Biochar Using
 1371 Chemical and Physical Activation: Mechanism and Modeling. In: Argyropoulos
 1372 DS, editor. Mater. Chem. Energy from For. Biomass, American Chemical
 1373 Society (ACS); 2007, p. 463–76. doi:10.1021/bk-2007-0954.ch029.
- 1374 [106] You S, Ok YS, Chen SS, Tsang DCW, Kwon EE, Lee J, et al. A critical review
 1375 on sustainable biochar system through gasification: Energy and environmental

1376 applications. *Bioresour Technol* 2017;246:242–53.
 1377 doi:10.1016/j.biortech.2017.06.177.

1378 [107] Li H, Dong X, da Silva EB, de Oliveira LM, Chen Y, Ma LQ. Mechanisms of
 1379 metal sorption by biochars: Biochar characteristics and modifications.
 1380 *Chemosphere* 2017;178:466–78. doi:10.1016/j.chemosphere.2017.03.072.

1381 [108] Chen Y, Zhang X, Chen W, Yang H, Chen H. The structure evolution of biochar
 1382 from biomass pyrolysis and its correlation with gas pollutant adsorption
 1383 performance. *Bioresour Technol* 2017;246:101–9.
 1384 doi:10.1016/j.biortech.2017.08.138.

1385 [109] Xie T, Reddy KR, Wang C, Yargicoglu E, Spokas K. Characteristics and
 1386 applications of biochar for environmental remediation: A review. *Crit Rev*
 1387 *Environ Sci Technol* 2015;45:939–69. doi:10.1080/10643389.2014.924180.

1388 [110] Yuan Y, Bolan N, Prévosteau A, Vithanage M, Biswas JK, Ok YS, et al.
 1389 Applications of biochar in redox-mediated reactions. *Bioresour Technol*
 1390 2017;246:271–81. doi:10.1016/j.biortech.2017.06.154.

1391 [111] Qambrani NA, Rahman MM, Won S, Shim S, Ra C. Biochar properties and eco-
 1392 friendly applications for climate change mitigation, waste management, and
 1393 wastewater treatment: A review. *Renew Sustain Energy Rev* 2017;79:255–73.
 1394 doi:10.1016/j.rser.2017.05.057.

1395 [112] Yin Q, Zhang B, Wang R, Zhao Z. Biochar as an adsorbent for inorganic
 1396 nitrogen and phosphorus removal from water: a review. *Environ Sci Pollut Res*
 1397 2017;24:26297–309. doi:10.1007/s11356-017-0338-y.

1398 [113] Trigo C, Cox L, Spokas K. Influence of pyrolysis temperature and hardwood
 1399 species on resulting biochar properties and their effect on azimsulfuron sorption

1400 as compared to other sorbents. *Sci Total Environ* 2016;566–567:1454–64.

1401 doi:10.1016/j.scitotenv.2016.06.027.

1402 [114] Chen T, Zhang Y, Wang H, Lu W, Zhou Z, Zhang Y, et al. Influence of pyrolysis

1403 temperature on characteristics and heavy metal adsorptive performance of

1404 biochar derived from municipal sewage sludge. *Bioresour Technol* 2014;164:47–

1405 54. doi:10.1016/j.biortech.2014.04.048.

1406 [115] Collison M, Collison L, Sakrabani R, Tofield B. *Biochar and Carbon*

1407 *Sequestration: A Regional Perspective* A report prepared for East of England

1408 Development Agency (EEDA) Biochar and Carbon 2009.

1409 [116] Antal MJ, Grønli M. The Art, Science, and Technology of Charcoal Production.

1410 *Ind Eng Chem Res* 2003;42:1619–1640. doi:10.1021/IE0207919.

1411 [117] Novak JM, Busscher WJ, Laird DL, Ahmedna M, Watts DW, Niandou MAS.

1412 Impact of Biochar Amendment on Fertility of a Southeastern Coastal Plain Soil.

1413 *Soil Sci* 2009;174:105–12. doi:10.1097/SS.0b013e3181981d9a.

1414 [118] Fiore S, Berruti F, Briens C. Investigation of innovative and conventional

1415 pyrolysis of ligneous and herbaceous biomasses for biochar production. *Biomass*

1416 *and Bioenergy* 2018.

1417 [119] Suliman W, Harsh JB, Abu-Lail NI, Fortuna AM, Dallmeyer I, Garcia-Perez M.

1418 Influence of feedstock source and pyrolysis temperature on biochar bulk and

1419 surface properties. *Biomass and Bioenergy* 2016;84:37–48.

1420 doi:10.1016/j.biombioe.2015.11.010.

1421 [120] Tan Z, Lin CSK, Ji X, Rainey TJ. Returning biochar to fields: A review. *Appl*

1422 *Soil Ecol* 2017;116:1–11. doi:10.1016/j.apsoil.2017.03.017.

1423 [121] Ahmed MB, Zhou JL, Ngo HH, Guo W. Insight into biochar properties and its

1424 cost analysis. *Biomass and Bioenergy* 2016;84:76–86.

1425 doi:10.1016/j.biombioe.2015.11.002.

1426 [122] Nguyen BT, Lehmann J. Black carbon decomposition under varying water

1427 regimes. *Org Geochem* 2009;40:846–53. doi:10.1016/j.orggeochem.2009.05.004.

1428 [123] Klüpfel L, Keiluweit M, Kleber M, Sander M. Redox Properties of Plant

1429 Biomass-Derived Black Carbon (Biochar). *Environ Sci Technol* 2014;48:5601–

1430 11. doi:10.1021/es500906d.

1431 [124] Singh, Balwant, Camps-Arbestain M, Lehmann J, (Eds). *Biochar: a guide to*

1432 *analytical methods*. CSIRO Publishing; 2017.

1433 [125] Appels L, Baeyens J, Degreè J, Dewil R. Principles and potential of the

1434 anaerobic digestion of waste-activated sludge. *Prog Energy Combust Sci*

1435 2008;34:755–81. doi:10.1016/j.pecs.2008.06.002.

1436 [126] Zhang C, Su H, Baeyens J, Tan T. Reviewing the anaerobic digestion of food

1437 waste for biogas production. *Renew Sustain Energy Rev* 2014;38:383–92.

1438 doi:10.1016/J.RSER.2014.05.038.

1439 [127] Ren Y, Yu M, Wu C, Wang Q, Gao M, Huang Q, et al. A comprehensive review

1440 on food waste anaerobic digestion: Research updates and tendencies. *Bioresour*

1441 *Technol* 2018;247:1069–76. doi:10.1016/J.BIORTECH.2017.09.109.

1442 [128] Di Maria F, Sordi A, Cirulli G, Micale C. Amount of energy recoverable from an

1443 existing sludge digester with the co-digestion with fruit and vegetable waste at

1444 reduced retention time. *Appl Energy* 2015;150:9–14.

1445 doi:10.1016/j.apenergy.2015.01.146.

1446 [129] Elalami D, Carrere H, Monlau F, Abdelouahdi K, Oukarroum A, Barakat A.

1447 Pretreatment and co-digestion of wastewater sludge for biogas production:

Recent research advances and trends. *Renew Sustain Energy Rev* 2019;114:109287. doi:10.1016/j.rser.2019.109287.

[130] Nghiem LD, Koch K, Bolzonella D, Drewes JE. Full scale co-digestion of wastewater sludge and food waste: Bottlenecks and possibilities. *Renew Sustain Energy Rev* 2017;72:354–62. doi:10.1016/j.rser.2017.01.062.

[131] Kang X, Sun Y, Li L, Kong X, Yuan Z. Improving methane production from anaerobic digestion of Pennisetum Hybrid by alkaline pretreatment. *Bioresour Technol* 2018;255:205–12. doi:10.1016/j.biortech.2017.12.001.

[132] Novais RM, Gameiro T, Carvalheiras J, Seabra MP, Tarelho LAC, Labrincha JA, et al. High pH buffer capacity biomass fly ash-based geopolymer spheres to boost methane yield in anaerobic digestion. *J Clean Prod* 2018;178:258–67. doi:10.1016/j.jclepro.2018.01.033.

[133] Novais RM, Buruberri LH, Seabra MP, Bajare D, Labrincha JA. Novel porous fly ash-containing geopolymers for pH buffering applications. *J Clean Prod* 2016;124:395–404. doi:10.1016/j.jclepro.2016.02.114.

[134] Wei W, Guo W, Ngo HH, Mannina G, Wang D, Chen X, et al. Enhanced high-quality biomethane production from anaerobic digestion of primary sludge by corn stover biochar. *Bioresour Technol* 2020;306:123159. doi:10.1016/j.biortech.2020.123159.

[135] Ambaye TG, Rene ER, Dupont C, Wongrod S, van Hullebusch ED. Anaerobic Digestion of Fruit Waste Mixed With Sewage Sludge Digestate Biochar: Influence on Biomethane Production. *Front Energy Res* 2020;8:31. doi:10.3389/fenrg.2020.00031.

[136] Qiang H, Lang DL, Li YY. High-solid mesophilic methane fermentation of food

1472 waste with an emphasis on Iron, Cobalt, and Nickel requirements. *Bioresour*
 1473 *Technol* 2012;103:21–7. doi:10.1016/j.biortech.2011.09.036.

1474 [137] Chen Y, Cheng JJ, Creamer KS. Inhibition of anaerobic digestion process: A
 1475 review. *Bioresour Technol* 2008;99:4044–64.
 1476 doi:10.1016/j.biortech.2007.01.057.

1477 [138] Wang G, Li Q, Li Y, Xing Y, Yao G, Liu Y, et al. Redox-active biochar
 1478 facilitates potential electron transfer between syntrophic partners to enhance
 1479 anaerobic digestion under high organic loading rate. *Bioresour Technol*
 1480 2020;298:122524. doi:10.1016/j.biortech.2019.122524.

1481 [139] Qin Y, Wang H, Li X, Cheng JJ, Wu W. Improving methane yield from organic
 1482 fraction of municipal solid waste (OFMSW) with magnetic rice-straw biochar.
 1483 *Bioresour Technol* 2017;245:1058–66. doi:10.1016/j.biortech.2017.09.047.

1484 [140] Rajagopal R, Massé DI, Singh G. A critical review on inhibition of anaerobic
 1485 digestion process by excess ammonia. *Bioresour Technol* 2013;143:632–41.
 1486 doi:10.1016/j.biortech.2013.06.030.

1487 [141] Zhang Y, Zamudio Cañas EM, Zhu Z, Linville JL, Chen S, He Q. Robustness of
 1488 archaeal populations in anaerobic co-digestion of dairy and poultry wastes.
 1489 *Bioresour Technol* 2011;102:779–85. doi:10.1016/j.biortech.2010.08.104.

1490 [142] Ho L, Ho G. Mitigating ammonia inhibition of thermophilic anaerobic treatment
 1491 of digested piggery wastewater: Use of pH reduction, zeolite, biomass and humic
 1492 acid. *Water Res* 2012;46:4339–50. doi:10.1016/j.watres.2012.05.016.

1493 [143] Poirier S, Madigou C, Bouchez T, Chapleur O. Improving anaerobic digestion
 1494 with support media: Mitigation of ammonia inhibition and effect on microbial
 1495 communities. *Bioresour Technol* 2017;235:229–39.

doi:10.1016/j.biortech.2017.03.099.

[144] Yenigün O, Demirel B. Ammonia inhibition in anaerobic digestion: A review. *Process Biochem* 2013;48:901–11. doi:10.1016/j.procbio.2013.04.012.

[145] Yun YM, Kim DH, Cho SK, Shin HS, Jung KW, Kim HW. Mitigation of ammonia inhibition by internal dilution in high-rate anaerobic digestion of food waste leachate and evidences of microbial community response. *Biotechnol Bioeng* 2016;113:1892–901. doi:10.1002/bit.25968.

[146] Fang C, Boe K, Angelidaki I. Anaerobic co-digestion of desugared molasses with cow manure; focusing on sodium and potassium inhibition. *Bioresour Technol* 2011;102:1005–11. doi:10.1016/j.biortech.2010.09.077.

[147] Wang Q, Yang Y, Yu C, Huang H, Kim M, Feng C, et al. Study on a fixed zeolite bioreactor for anaerobic digestion of ammonium-rich swine wastes. *Bioresour Technol* 2011;102:7064–8. doi:10.1016/j.biortech.2011.04.085.

[148] Wang W, Ren X, Yang K, Hu Z, Yuan S. Inhibition of ammonia on anaerobic digestion of synthetic coal gasification wastewater and recovery using struvite precipitation. *J Hazard Mater* 2017;340:152–9. doi:10.1016/j.jhazmat.2017.07.002.

[149] Zhang Y, Angelidaki I. Counteracting ammonia inhibition during anaerobic digestion by recovery using submersible microbial desalination cell. *Biotechnol Bioeng* 2015;112:1478–82. doi:10.1002/bit.25549.

[150] Ghyselbrecht K, Monballiu A, Somers MH, Sigurnjak I, Meers E, Appels L, et al. Stripping and scrubbing of ammonium using common fractionating columns to prove ammonium inhibition during anaerobic digestion. *Int J Energy Environ Eng* 2018;9:447–55. doi:10.1007/s40095-018-0283-7.

- 1520 [151] Shen Y, Linville JL, Urgun-Demirtas M, Schoene RP, Snyder SW. Producing
1521 pipeline-quality biomethane via anaerobic digestion of sludge amended with corn
1522 stover biochar with in-situ CO₂ removal. *Appl Energy* 2015;158:300–9.
1523 doi:10.1016/j.apenergy.2015.08.016.
- 1524 [152] Ciccoli R, Sperandei M, Petrazzuolo F, Broglia M, Chiarini L, Correnti A, et al.
1525 Anaerobic digestion of the above ground biomass of Jerusalem Artichoke in a
1526 pilot plant: Impact of the preservation method on the biogas yield and microbial
1527 community. *Biomass and Bioenergy* 2018;108:190–7.
1528 doi:10.1016/j.biombioe.2017.11.003.
- 1529 [153] Lü F, Hua Z, Shao L, He P. Loop bioenergy production and carbon sequestration
1530 of polymeric waste by integrating biochemical and thermochemical conversion
1531 processes: A conceptual framework and recent advances. *Renew Energy*
1532 2018;124:202–11. doi:10.1016/j.renene.2017.10.084.
- 1533 [154] Takaya CA, Fletcher LA, Singh S, Anyikude KU, Ross AB. Phosphate and
1534 ammonium sorption capacity of biochar and hydrochar from different wastes.
1535 *Chemosphere* 2016;145:518–27. doi:10.1016/j.chemosphere.2015.11.052.
- 1536 [155] Kizito S, Wu S, Kirui WK, Lei M, Lu Q, Bah H, et al. Evaluation of slow
1537 pyrolyzed wood and rice husks biochar for adsorption of ammonium nitrogen
1538 from piggery manure anaerobic digestate slurry. *Sci Total Environ*
1539 2015;505:102–12. doi:10.1016/j.scitotenv.2014.09.096.
- 1540 [156] Wang Z, Guo H, Shen F, Yang G, Zhang Y, Zeng Y, et al. Biochar produced
1541 from oak sawdust by Lanthanum (La)-involved pyrolysis for adsorption of
1542 ammonium (NH₄⁺), nitrate (NO₃⁻), and phosphate (PO₄³⁻). *Chemosphere*
1543 2015;119:646–53. doi:10.1016/j.chemosphere.2014.07.084.

- 1544 [157] Zeng Z, Zhang S, Li T, Zhao F, He Z, Zhao H, et al. Sorption of ammonium and
1545 phosphate from aqueous solution by biochar derived from phytoremediation
1546 plants. *J Zhejiang Univ Sci B* 2013;14:1152–61. doi:10.1631/jzus.B1300102.
- 1547 [158] Zhang Y, Li Z, Mahmood IB. Recovery of NH_4^+ by corn cob produced biochars
1548 and its potential application as soil conditioner. *Front Environ Sci Eng*
1549 2014;8:825–34. doi:10.1007/s11783-014-0682-9.
- 1550 [159] Kizito S, Wu S, Wandera SM, Guo L, Dong R. Evaluation of ammonium
1551 adsorption in biochar-fixed beds for treatment of anaerobically digested swine
1552 slurry: Experimental optimization and modeling. *Sci Total Environ* 2016;563–
1553 564:1095–104. doi:10.1016/j.scitotenv.2016.05.149.
- 1554 [160] Lopez RJ, Higgins SR, Pagaling E, Yan T, Cooney MJ. High rate anaerobic
1555 digestion of wastewater separated from grease trap waste. *Renew Energy*
1556 2014;62:234–42. doi:10.1016/j.renene.2013.06.047.
- 1557 [161] Lettinga G. Anaerobic digestion and wastewater treatment systems. *Antonie Van*
1558 *Leeuwenhoek* 1995;67:3–28. doi:10.1007/BF00872193.
- 1559 [162] Laine J, Simoni S, Calles R. Preparation of activated carbon from coconut shell
1560 in a small scale cocurrent flow rotary kiln. *Chem Eng Commun* 1991;99:15–23.
1561 doi:10.1080/00986449108911575.
- 1562 [163] Watanabe Y, Tanaka K. Innovative sludge handling through
1563 pelletization/thickening. *Water Res* 1999;33:3245–52. doi:10.1016/S0043-
1564 1354(99)00045-7.
- 1565 [164] Sawayama S, Tada C, Tsukahara K, Yagishita T. Effect of ammonium addition
1566 on methanogenic community in a fluidized bed anaerobic digestion. *J Biosci*
1567 *Bioeng* 2004;97:65–70. doi:10.1016/S1389-1723(04)70167-X.

- 1568 [165] Stams AJM. Metabolic interactions between anaerobic bacteria in methanogenic
1569 environments. *Antonie Van Leeuwenhoek* 1994;66:271–94.
1570 doi:10.1007/BF00871644.
- 1571 [166] Whitman WB, Bowen TL, Boone DR. *The Methanogenic Bacteria*. The
1572 Prokaryotes, New York, NY: Springer New York; 2006, p. 165–207.
1573 doi:10.1007/0-387-30743-5_9.
- 1574 [167] Morris BEL, Henneberger R, Huber H, Moissl-Eichinger C. Microbial syntrophy:
1575 Interaction for the common good. *FEMS Microbiol Rev* 2013;37:384–406.
1576 doi:10.1111/1574-6976.12019.
- 1577 [168] Schink B, Montag D, Keller A, Müller N. Hydrogen or formate: Alternative key
1578 players in methanogenic degradation. *Environ Microbiol Rep* 2017;9:189–202.
1579 doi:10.1111/1758-2229.12524.
- 1580 [169] Lovley DR, Fraga JL, Coates JD, Blunt-Harris EL. Humics as an electron donor
1581 for anaerobic respiration. *Environ Microbiol* 1999;1:89–98. doi:10.1046/j.1462-
1582 2920.1999.00009.x.
- 1583 [170] Roden EE, Kappler A, Bauer I, Jiang J, Paul A, Stoesser R, et al. Extracellular
1584 electron transfer through microbial reduction of solid-phase humic substances.
1585 *Nat Geosci* 2010;3:417–21. doi:10.1038/ngeo870.
- 1586 [171] Park JH, Kang HJ, Park KH, Park HD. Direct interspecies electron transfer via
1587 conductive materials: A perspective for anaerobic digestion applications.
1588 *Bioresour Technol* 2018;254:300–11. doi:10.1016/j.biortech.2018.01.095.
- 1589 [172] McGlynn SE, Chadwick GL, Kempes CP, Orphan VJ. Single cell activity reveals
1590 direct electron transfer in methanotrophic consortia. *Nature* 2015;526:531–5.
1591 doi:10.1038/nature15512.

- 1592 [173] Lovley DR. Happy together: microbial communities that hook up to swap
1593 electrons. *ISME J* 2017;11:327–36. doi:10.1038/ismej.2016.136.
- 1594 [174] Stams AJM, Plugge CM. Electron transfer in syntrophic communities of
1595 anaerobic bacteria and archaea. *Nat Rev Microbiol* 2009;7:568–77.
1596 doi:10.1038/nrmicro2166.
- 1597 [175] Schink B, Thauer R. Energetics of syntrophic methane formation and the
1598 influence of aggregation. In: Lettinga G, Zehnder, A.J.B. Grotenhuis, J.T.C.
1599 Hulshoff LW, editors. *Granul. Anaerob. sludge; Microbiol. Technol. Proc.*
1600 *GASMAT - Work.* Lunteren, Netherlands, 25-27 Oct. 1987 , Centre for
1601 Agricultural Publishing and Documentation; 1988, p. 5–17.
- 1602 [176] Summers ZM, Fogarty HE, Leang C, Franks AE, Malvankar NS, Lovley DR.
1603 Direct Exchange of Electrons Within Aggregates of an Evolved Syntrophic
1604 Coculture of Anaerobic Bacteria. *Science* (80-) 2010;330:1413–5.
1605 doi:10.1126/science.1196526.
- 1606 [177] Rotaru AE, Shrestha PM, Liu F, Shrestha M, Shrestha D, Embree M, et al. A new
1607 model for electron flow during anaerobic digestion: Direct interspecies electron
1608 transfer to *Methanosaeta* for the reduction of carbon dioxide to methane. *Energy*
1609 *Environ Sci* 2014;7:408–15. doi:10.1039/c3ee42189a.
- 1610 [178] Rotaru A-E, Shrestha PM, Liu F, Markovaite B, Chen S, Nevin KP, et al. Direct
1611 interspecies electron transfer between *Geobacter metallireducens* and
1612 *Methanosarcina barkeri*. *Appl Environ Microbiol* 2014;80:4599–605.
1613 doi:10.1128/AEM.00895-14.
- 1614 [179] Liu F, Rotaru A-E, Shrestha PM, Malvankar NS, Nevin KP, Lovley DR.
1615 Magnetite compensates for the lack of a pilin-associated *c* -type cytochrome in

extracellular electron exchange. *Environ Microbiol* 2015;17:648–55.
doi:10.1111/1462-2920.12485.

[180] Liu F, Rotaru A-E, Shrestha PM, Malvankar NS, Nevin KP, Lovley DR.
Promoting direct interspecies electron transfer with activated carbon. *Energy*
Environ Sci 2012;5:8982. doi:10.1039/c2ee22459c.

[181] Barua S, Dhar BR. Advances towards understanding and engineering direct
interspecies electron transfer in anaerobic digestion. *Bioresour Technol*
2017;244:698–707. doi:10.1016/j.biortech.2017.08.023.

[182] Cheah S, Malone SC, Feik CJ. Speciation of Sulfur in Biochar Produced from
Pyrolysis and Gasification of Oak and Corn Stover. *Environ Sci Technol*
2014;48:8474–80. doi:10.1021/es500073r.

[183] Rosas JG, Gómez N, Cara J, Ubalde J, Sort X, Sánchez ME. Assessment of
sustainable biochar production for carbon abatement from vineyard residues. *J*
Anal Appl Pyrolysis 2015;113:239–47. doi:10.1016/J.JAAP.2015.01.011.

[184] Coelho JJ, Prieto ML, Dowling S, Hennessy A, Casey I, Woodcock T, et al.
Physical-chemical traits, phytotoxicity and pathogen detection in liquid anaerobic
digestates. *Waste Manag* 2018;78:8–15. doi:10.1016/j.wasman.2018.05.017.

[185] Di Maria F, Sordi A, Cirulli G, Gigliotti G, Massaccesi L, Cucina M. Co-
treatment of fruit and vegetable waste in sludge digesters. An analysis of the
relationship among bio-methane generation, process stability and digestate
phytotoxicity. *Waste Manag* 2014;34:1603–8.
doi:10.1016/j.wasman.2014.05.017.

[186] Di Maria F, Gigliotti G, Sordi A, Micale C, Zadra C, Massaccesi L. Hybrid solid
anaerobic digestion batch: biomethane production and mass recovery from the

1640 organic fraction of solid waste. *Waste Manag Res* 2013;31:869–73.
 1641 doi:10.1177/0734242X13477902.
 1642 [187] Moukazis I, Pelleri FM, Gidaracos E. Slaughterhouse by-products treatment
 1643 using anaerobic digestion. *Waste Manag* 2018;71:652–62.
 1644 doi:10.1016/j.wasman.2017.07.009.
 1645 [188] Fuchs W, Drosch B. Assessment of the state of the art of technologies for the
 1646 processing of digestate residue from anaerobic digesters. *Water Sci Technol*
 1647 2013;67:1984–93. doi:10.2166/wst.2013.075.
 1648 [189] Vaneeckhaute C, Lebuf V, Michels E, Belia E, Vanrolleghem PA, Tack FMG, et
 1649 al. Nutrient Recovery from Digestate: Systematic Technology Review and
 1650 Product Classification. *Waste and Biomass Valorization* 2017;8:21–40.
 1651 doi:10.1007/s12649-016-9642-x.
 1652 [190] Egle L, Rechberger H, Zessner M. Overview and description of technologies for
 1653 recovering phosphorus from municipal wastewater. *Resour Conserv Recycl*
 1654 2015;105:325–46. doi:10.1016/j.resconrec.2015.09.016.
 1655 [191] Dahlin J, Nelles M, Herbes C. Biogas digestate management: Evaluating the
 1656 attitudes and perceptions of German gardeners towards digestate-based soil
 1657 amendments. *Resour Conserv Recycl* 2017;118:27–38.
 1658 doi:10.1016/j.resconrec.2016.11.020.
 1659 [192] Bagreev A, Bandosz TJ, Locke DC. Pore structure and surface chemistry of
 1660 adsorbents obtained by pyrolysis of sewage sludge-derived fertilizer. *Carbon N Y*
 1661 2001;39:1971–9. doi:10.1016/S0008-6223(01)00026-4.
 1662 [193] Jiang B, Lin Y, Mbog JC. Biochar derived from swine manure digestate and
 1663 applied on the removals of heavy metals and antibiotics. *Bioresour Technol*

1664 2018;270:603–11. doi:10.1016/j.biortech.2018.08.022.

1665 [194] Li J, Zhang M, Ye Z, Yang C. Effect of manganese oxide-modified biochar
1666 addition on methane production and heavy metal speciation during the anaerobic
1667 digestion of sewage sludge. *J Environ Sci* 2019;76:267–77.
1668 doi:10.1016/j.jes.2018.05.009.

1669 [195] Zhang L, Li F, Kuroki A, Loh KC, Wang CH, Dai Y, et al. Methane yield
1670 enhancement of mesophilic and thermophilic anaerobic co-digestion of algal
1671 biomass and food waste using algal biochar: Semi-continuous operation and
1672 microbial community analysis. *Bioresour Technol* 2020;302:122892.
1673 doi:10.1016/j.biortech.2020.122892.

1674 [196] Martin SL, Clarke ML, Othman M, Ramsden SJ, West HM. Biochar-mediated
1675 reductions in greenhouse gas emissions from soil amended with anaerobic
1676 digestates. *Biomass and Bioenergy* 2014;79:39–49.
1677 doi:10.1016/j.biombioe.2015.04.030.

1678 [197] Marchetti R, Castelli F. Biochar from Swine Solids and Digestate Influence
1679 Nutrient Dynamics and Carbon Dioxide Release in Soil. *J Environ Qual*
1680 2013;42:893–901. doi:10.2134/jeq2012.0352.

1681 [198] Mukherjee S, Weihermueller L, Tappe W, Vereecken H, Burauel P. Microbial
1682 respiration of biochar- and digestate-based mixtures. *Biol Fertil Soils*
1683 2016;52:151–64. doi:10.1007/s00374-015-1060-x.

1684 [199] Cardelli R, Giussani G, Marchini F, Saviozzi A. Short-term effects on soil of
1685 biogas digestate, biochar and their combinations. *Soil Res* 2018;56:623.
1686 doi:10.1071/SR18017.

1687 [200] Vaughn SF, Eller FJ, Evangelista RL, Moser BR, Lee E, Wagner RE, et al.

1688 Evaluation of biochar-anaerobic potato digestate mixtures as renewable
1689 components of horticultural potting media. *Ind Crops Prod* 2015;65:467–71.
1690 doi:10.1016/j.indcrop.2014.10.040.

1691 [201] Zhang J, Lü F, Shao L, He P. The use of biochar-amended composting to
1692 improve the humification and degradation of sewage sludge. *Bioresour Technol*
1693 2014;168:252–8. doi:10.1016/j.biortech.2014.02.080.

1694 [202] Bustamante MA, Albuquerque JA, Restrepo AP, de la Fuente C, Paredes C,
1695 Moral R, et al. Co-composting of the solid fraction of anaerobic digestates, to
1696 obtain added-value materials for use in agriculture. *Biomass and Bioenergy*
1697 2012;43:26–35. doi:10.1016/j.biombioe.2012.04.010.

1698 [203] Godlewska P, Schmidt HP, Ok YS, Oleszczuk P. Biochar for composting
1699 improvement and contaminants reduction. A review. *Bioresour Technol*
1700 2017;246:193–202. doi:10.1016/j.biortech.2017.07.095.

1701 [204] Xiao R, Awasthi MK, Li R, Park J, Pensky SM, Wang Q, et al. Recent
1702 developments in biochar utilization as an additive in organic solid waste
1703 composting: A review. *Bioresour Technol* 2017;246:203–13.
1704 doi:10.1016/j.biortech.2017.07.090.

1705 [205] Sanchez-Monedero MA, Cayuela ML, Roig A, Jindo K, Mondini C, Bolan N.
1706 Role of biochar as an additive in organic waste composting. *Bioresour Technol*
1707 2018;247:1155–64. doi:10.1016/j.biortech.2017.09.193.

1708 [206] Cristina G, Camelin E, Pugliese M, Tommasi T, Fino D. Evaluation of anaerobic
1709 digestates from sewage sludge as a potential solution for improvement of soil
1710 fertility. *Waste Manag* 2019;99:122–34. doi:10.1016/j.wasman.2019.08.018.

1711 [207] Angelidaki I, Treu L, Tsapekos P, Luo G, Campanaro S, Wenzel H, et al. Biogas

1712 upgrading and utilization: Current status and perspectives. *Biotechnol Adv*
1713 2018;36:452–66. doi:10.1016/J.BIOTECHADV.2018.01.011.

1714 [208] Browne J, Nizami A-S, Thamsiroj T, Murphy JD. Assessing the cost of biofuel
1715 production with increasing penetration of the transport fuel market: A case study
1716 of gaseous biomethane in Ireland. *Renew Sustain Energy Rev* 2011;15:4537–47.
1717 doi:10.1016/J.RSER.2011.07.098.

1718 [209] Petersson A, WeLLInGer A. Biogas upgrading technologies–developments and
1719 innovations. *IEA Bioenergy* 2009;20:1–19.

1720 [210] Sun Q, Li H, Yan J, Liu L, Yu Z, Yu X. Selection of appropriate biogas
1721 upgrading technology-a review of biogas cleaning, upgrading and utilisation.
1722 *Renew Sustain Energy Rev* 2015;51:521–32. doi:10.1016/J.RSER.2015.06.029.

1723 [211] Miltner M, Makaruk A, Harasek M. Review on available biogas upgrading
1724 technologies and innovations towards advanced solutions. *J Clean Prod*
1725 2017;161:1329–37. doi:10.1016/J.JCLEPRO.2017.06.045.

1726 [212] Singh G, Lakhi KS, Sil S, Bhosale S V., Kim IY, Albahily K, et al. Biomass
1727 derived porous carbon for CO₂ capture. *Carbon N Y* 2019;148:164–86.
1728 doi:10.1016/j.carbon.2019.03.050.

1729 [213] Ahmed R, Liu G, Yousaf B, Abbas Q, Ullah H, Ali MU. Recent advances in
1730 carbon-based renewable adsorbent for selective carbon dioxide capture and
1731 separation-A review. *J Clean Prod* 2020;242:118409.
1732 doi:10.1016/j.jclepro.2019.118409.

1733 [214] Dissanayake PD, You S, Igalavithana AD, Xia Y, Bhatnagar A, Gupta S, et al.
1734 Biochar-based adsorbents for carbon dioxide capture: A critical review. *Renew*
1735 *Sustain Energy Rev* 2020;119:109582. doi:10.1016/j.rser.2019.109582.

- 1736 [215] Sethupathi S, Zhang M, Rajapaksha A, Lee S, Mohamad Nor N, Mohamed A, et
1737 al. Biochars as Potential Adsorbers of CH₄, CO₂ and H₂S. Sustainability
1738 2017;9:121. doi:10.3390/su9010121.
- 1739 [216] Creamer AE, Gao B, Zhang M. Carbon dioxide capture using biochar produced
1740 from sugarcane bagasse and hickory wood. Chem Eng J 2014;249:174–9.
1741 doi:10.1016/J.CEJ.2014.03.105.
- 1742 [217] Creamer AE, Gao B. Carbon-Based Adsorbents for Postcombustion CO₂
1743 Capture: A Critical Review. Environ Sci Technol 2016;50:7276–89.
1744 doi:10.1021/acs.est.6b00627.
- 1745 [218] Presser V, McDonough J, Yeon S-H, Gogotsi Y. Effect of pore size on carbon
1746 dioxide sorption by carbide derived carbon. Energy Environ Sci 2011;4:3059.
1747 doi:10.1039/c1ee01176f.
- 1748 [219] Xu X, Kan Y, Zhao L, Cao X. Chemical transformation of CO₂ during its
1749 capture by waste biomass derived biochars. Environ Pollut 2016;213:533–40.
1750 doi:10.1016/j.envpol.2016.03.013.
- 1751 [220] Jung S, Park YK, Kwon EE. Strategic use of biochar for CO₂ capture and
1752 sequestration. J CO₂ Util 2019;32:128–39. doi:10.1016/j.jcou.2019.04.012.
- 1753 [221] Sahota S, Vijay VK, Subbarao PMV, Chandra R, Ghosh P, Shah G, et al.
1754 Characterization of leaf waste based biochar for cost effective hydrogen sulphide
1755 removal from biogas. Bioresour Technol 2018;250:635–41.
1756 doi:10.1016/J.BIORTECH.2017.11.093.
- 1757 [222] Kanjanarong J, Giri BS, Jaisi DP, Oliveira FR, Boonsawang P, Chaiprapat S, et
1758 al. Removal of hydrogen sulfide generated during anaerobic treatment of sulfate-
1759 laden wastewater using biochar: Evaluation of efficiency and mechanisms.

1760 Bioresour Technol 2017;234:115–21. doi:10.1016/J.BIORTECH.2017.03.009.
 1761 [223] Bamdad H, Hawboldt K, MacQuarrie S. A review on common adsorbents for
 1762 acid gases removal: Focus on biochar. Renew Sustain Energy Rev
 1763 2018;81:1705–20. doi:10.1016/J.RSER.2017.05.261.
 1764 [224] Sun Y, Zhang JP, Wen C, Zhang L. An enhanced approach for biochar
 1765 preparation using fluidized bed and its application for H₂S removal. Chem Eng
 1766 Process Process Intensif 2016;104:1–12. doi:10.1016/j.cep.2016.02.006.
 1767 [225] Xu X, Cao X, Zhao L, Sun T. Comparison of sewage sludge- and pig manure-
 1768 derived biochars for hydrogen sulfide removal. Chemosphere 2014;111:296–303.
 1769 doi:10.1016/j.chemosphere.2014.04.014.
 1770 [226] Sun Y, Yang G, Zhang L, Sun Z. Preparation of high performance H₂S removal
 1771 biochar by direct fluidized bed carbonization using potato peel waste. Process Saf
 1772 Environ Prot 2017;107:281–8. doi:10.1016/j.psep.2017.02.018.
 1773 [227] Shang G, Shen G, Liu L, Chen Q, Xu Z. Kinetics and mechanisms of hydrogen
 1774 sulfide adsorption by biochars. Bioresour Technol 2013;133:495–9.
 1775 doi:10.1016/j.biortech.2013.01.114.
 1776 [228] Madzaki H, Karimghani WAWAB, Nurzalikharebitanim, Azilbaharialias.
 1777 Carbon Dioxide Adsorption on Sawdust Biochar. Procedia Eng 2016;148:718–
 1778 25. doi:10.1016/j.proeng.2016.06.591.
 1779 [229] Shahkarami S, Azargohar R, Dalai AK, Soltan J. Breakthrough CO₂ adsorption
 1780 in bio-based activated carbons. J Environ Sci (China) 2015;34:68–76.
 1781 doi:10.1016/j.jes.2015.03.008.
 1782 [230] Lahijani P, Mohammadi M, Mohamed AR. Metal incorporated biochar as a
 1783 potential adsorbent for high capacity CO₂ capture at ambient condition. J CO₂

Util 2018;26:281–93. doi:10.1016/j.jcou.2018.05.018.

[231] Guedes RE, Luna AS, Torres AR. Operating parameters for bio-oil production in biomass pyrolysis: A review. *J Anal Appl Pyrolysis* 2018;129:134–49. doi:10.1016/j.jaap.2017.11.019.

[232] Li W, Dang Q, Brown RC, Laird D, Wright MM. The impacts of biomass properties on pyrolysis yields, economic and environmental performance of the pyrolysis-bioenergy-biochar platform to carbon negative energy. *Bioresour Technol* 2017;241:959–68. doi:10.1016/j.biortech.2017.06.049.

[233] Ji C, Cheng K, Nayak D, Pan G. Environmental and economic assessment of crop residue competitive utilization for biochar, briquette fuel and combined heat and power generation. *J Clean Prod* 2018;192:916–23. doi:10.1016/j.jclepro.2018.05.026.

[234] Harsono SS, Grundman P, Lau LH, Hansen A, Salleh MAM, Meyer-Aurich A, et al. Energy balances, greenhouse gas emissions and economics of biochar production from palm oil empty fruit bunches. *Resour Conserv Recycl* 2013;77:108–15. doi:10.1016/j.resconrec.2013.04.005.

[235] Sahoo K, Bilek E, Bergman R, Mani S. Techno-economic analysis of producing solid biofuels and biochar from forest residues using portable systems. *Appl Energy* 2019;235:578–90. doi:10.1016/J.APENERGY.2018.10.076.

[236] ICIS. Analytics - ICIS Explore 2018. <https://www.icis.com/explore/services/analytics/?intcmp=mega-menu-explore-services-analytics&redirect=english> (accessed January 2, 2019).

[237] Menya E, Olupot PW, Storz H, Lubwama M, Kiros Y. Production and performance of activated carbon from rice husks for removal of natural organic

1808 matter from water: A review. Chem Eng Res Des 2018;129:271–96.
 1809 doi:10.1016/j.cherd.2017.11.008.

1810 [238] Shabangu S, Woolf D, Fisher EM, Angenent LT, Lehmann J. Techno-economic
 1811 assessment of biomass slow pyrolysis into different biochar and methanol
 1812 concepts. Fuel 2014;117:742–8. doi:10.1016/j.fuel.2013.08.053.

1813 [239] Dickinson D, Balduccio L, Buysse J, Ronsse F, van Huylbroeck G, Prins W.
 1814 Cost-benefit analysis of using biochar to improve cereals agriculture. GCB
 1815 Bioenergy 2015;7:850–64. doi:10.1111/gcbb.12180.

1816 [240] Jirka S, Tomlinson T. State of the biochar industry 2014. A survey of commercial
 1817 activity in the biochar field. 2015.

1818 [241] Jirka S, Tomlinson T. 2013 State of the biochar industry. A Survey of
 1819 commercial activity in the biochar field. 2014.

1820 [242] Antoniou N, Monlau F, Sambusiti C, Ficara E, Barakat A, Zabaniotou A.
 1821 Contribution to Circular Economy options of mixed agricultural wastes
 1822 management: Coupling anaerobic digestion with gasification for enhanced
 1823 energy and material recovery. J Clean Prod 2019;209:505–14.
 1824 doi:10.1016/j.jclepro.2018.10.055.

1825 [243] Zhang L, Lim EY, Loh KC, Ok YS, Lee JTE, Shen Y, et al. Biochar enhanced
 1826 thermophilic anaerobic digestion of food waste: Focusing on biochar particle
 1827 size, microbial community analysis and pilot-scale application. Energy Convers
 1828 Manag 2020;209:112654. doi:10.1016/j.enconman.2020.112654.

1829 [244] Sharifzadeh M, Sadeqzadeh M, Guo M, Borhani TN, Murthy Konda NVSN,
 1830 Garcia MC, et al. The multi-scale challenges of biomass fast pyrolysis and bio-oil
 1831 upgrading: Review of the state of art and future research directions. Prog Energy

1832 Combust Sci 2019;71:1–80. doi:10.1016/J.PECS.2018.10.006.

1833 [245] GREET. GREET Models 2018. <https://greet.es.anl.gov/greet.models> (accessed
1834 January 2, 2019).

1835 [246] EPA U. Overview for Renewable Fuel Standard 2017.

1836 [247] Moreira MT, Noya I, Feijoo G. The prospective use of biochar as adsorption
1837 matrix – A review from a lifecycle perspective. Bioresour Technol
1838 2017;246:135–41. doi:10.1016/j.biortech.2017.08.041.

1839 [248] Mohan D, Sarswat A, Ok YS, Pittman CU. Organic and inorganic contaminants
1840 removal from water with biochar, a renewable, low cost and sustainable
1841 adsorbent - A critical review. Bioresour Technol 2014;160:191–202.
1842 doi:10.1016/j.biortech.2014.01.120.

1843 [249] Ibarrola R, Shackley S, Hammond J. Pyrolysis biochar systems for recovering
1844 biodegradable materials: A life cycle carbon assessment. Waste Manag
1845 2012;32:859–68. doi:10.1016/j.wasman.2011.10.005.

1846 [250] Pratt K, Moran D. Evaluating the cost-effectiveness of global biochar mitigation
1847 potential. Biomass and Bioenergy 2010;34:1149–58.
1848 doi:10.1016/j.biombioe.2010.03.004.

1849 [251] Nowrouzi M, Younesi H, Bahramifar N. High efficient carbon dioxide capture
1850 onto as-synthesized activated carbon by chemical activation of Persian Ironwood
1851 biomass and the economic pre-feasibility study for scale-up. J Clean Prod
1852 2017;168:499–509. doi:10.1016/j.jclepro.2017.09.080.

1853 [252] Muñoz E, Curaqueo G, Cea M, Vera L, Navia R. Environmental hotspots in the
1854 life cycle of a biochar-soil system. J Clean Prod 2017;158:1–7.
1855 doi:10.1016/j.jclepro.2017.04.163.

- [253] Hammond J, Shackley S, Sohi S, Brownsort P. Prospective life cycle carbon abatement for pyrolysis biochar systems in the UK. *Energy Policy* 2011;39:2646–55. doi:10.1016/j.enpol.2011.02.033.
- [254] Brassard P, Godbout S, Pelletier F, Raghavan V, Palacios JH. Biomass and Bioenergy Pyrolysis of switchgrass in an auger reactor for biochar production : A greenhouse gas and energy impacts assessment. *Biomass and Bioenergy* 2018;116:99–105. doi:10.1016/j.biombioe.2018.06.007.
- [255] Righi S, Bandini V, Marazza D, Baioli F, Torri C, Contin A. Life Cycle Assessment of high ligno-cellulosic biomass pyrolysis coupled with anaerobic digestion. *Bioresour Technol* 2016;212:245–53. doi:10.1016/j.biortech.2016.04.052.
- [256] Matušík J, Hnátková T, Kočí V. Life cycle assessment of biochar-to-soil systems: A review. *J Clean Prod* 2020;259. doi:10.1016/j.jclepro.2020.120998.
- [257] Hjaila K, Baccar R, Sarrà M, Gasol CM, Blánquez P. Environmental impact associated with activated carbon preparation from olive-waste cake via life cycle assessment. *J Environ Manage* 2013;130:242–7. doi:10.1016/j.jenvman.2013.08.061.
- [258] Kookana RS, Sarmah AK, Van Zwieten L, Krull E, Singh B. Biochar application to soil. agronomic and environmental benefits and unintended consequences. vol. 112. 1st ed. Elsevier Inc.; 2011. doi:10.1016/B978-0-12-385538-1.00003-2.
- [259] Li H, Feng K. Life cycle assessment of the environmental impacts and energy efficiency of an integration of sludge anaerobic digestion and pyrolysis. *J Clean Prod* 2018;195:476–85. doi:10.1016/J.JCLEPRO.2018.05.259.