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The role of hairs in the adhesion of octopus suckers: a hierarchical peeling approach

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Abstract

Organisms like the octopus or the clingfish are a precious source of inspiration for the design of innovative adhesive systems based on suction cups, but a complete mechanical description of their attachment process is still lacking. In this paper, we exploit the recent discovery of the presence of hairs in the acetabulum roof of octopus suction cups to revise the current model for its adhesion to the acetabulum wall. We show how this additional feature, which can be considered an example of a hierarchical structure, can lead to an increase of adhesive strength, based on the analysis of the cases of a simple tape and an axisymmetrical membrane. Using peeling theory, we discuss in both cases the influence of hierarchical structure and the resulting variation of contact angles on the adhesive energy, highlight how an increase in number of hierarchical levels contributes to its increase, with a corresponding improvement in functionality for the octopus suckers.

1. Introduction

The *Octopus vulgaris* is one of the most intelligent animals that lives on Earth. It uses its suckers to perform many functions ([1], [2]). In particular, octopus suckers are able to generate a maximum pressure difference of about 0.27 MPa that can be reached in a few milliseconds [3]. Other animals, such as clingfish, exploit suction cups with a bed of microfibrils or “micropapillae”, which are tiny soft protuberances that line the cup perimeter, to better adhere to rough rock surfaces underwater [4]. For this reason, these structures represent a remarkable source of inspiration for designing artificial suction cups or adhesives ([5]–[8]). To develop these artificial devices, the full

35 understanding of the adhesion process and the capability to model it correctly is crucial. In the past,
 36 octopus suckers and their interaction with the substrate have been studied mainly by analyzing their
 37 arrangement [9] and structure ([10], [11]). In Tramacere et al.[9], a method to identify the suckers
 38 in the octopus arm was developed in order to better determine its mechanics through imaging.
 39 Moreover, in Tramacere et al. [10], three techniques (MRI, ultrasonography, and histology) were
 40 used to gain a 3D reconstruction of the sucker (Fig. 1). In this context, the acetabulum protuberance
 41 in the acetabulum cavity was discovered for the first time. Experimental studies were also
 42 performed to measure the full mechanical properties of the octopus sucker tissues in [11].
 43 Unfortunately, a reliable value of the Poisson ratio remains to be obtained. Work is in progress to
 44 resolve this issue. The adhesion of the octopus suckers is achieved by exploiting the pressure
 45 difference between the external environment, the acetabulum cavity and the infundibulum cavity
 46 (Fig. 1a) [12]. To maintain this pressure difference, the acetabulum roof and the acetabulum wall
 47 must remain in full contact [10]. More in detail, at the initial stage of adhesion, the infundibulum
 48 is the first part of the sucker in contact with the substrate to form a seal. Then, the acetabular radial
 49 muscles contract to reduce the internal pressure in the sucker with respect to the external one.
 50 Finally, the meridional muscle of the acetabulum contracts to achieve contact between the
 51 acetabulum roof and the acetabulum cavity. At this point, all muscles are contracted. When they
 52 relax, the adhesion is maintained by the adhesive force maintaining the two surfaces in contact (the
 53 acetabulum roof and the acetabulum cavity) [13]. Morphological studies show that the latter does
 54 not present any hairs and can be considered flat.

55 As in other bioadhesion problems, peeling theory has been adopted to describe how these two parts
 56 of the octopus suckers delaminate [14]. The first elastic approach developed in the literature in this
 57 respect was the Kendall model [15], which describes the peeling of a thin elastic tape from a rigid
 58 substrate. The main physical quantity that governs the attachment, or the detachment, of the tape
 59 is the surface energy γ , which is defined as the energy required to generate a unit area of interface
 60 (for a certain crack speed), with Mode I (opening) primary separation mode. In the Kendall model,
 61 the force necessary to detach the membrane can be determined by adopting an energy-based
 62 criterion, imposing the Griffith's balance between the elastic energy, the adhesive energy and the
 63 work of the applied load [16]. The peeling force relative to a tape pulled at an angle θ , is thus:

$$64 \quad F = Etw \left(\cos \alpha_0 - 1 + \sqrt{(1 - \cos \alpha_0)^2 + \frac{2\gamma}{Et}} \right) \quad (1)$$

65 where E is the Young's modulus of the tape, t its thickness and w its width. Introducing $\hat{F} =$
66 $F/(Etw)$, where Etw represents the force necessary to generate a unit strain in the tape, and $\hat{\gamma} =$
67 $\gamma/(Et)$, the relation can be written in non-dimensional form:

$$68 \quad \hat{F} = \cos \alpha_0 - 1 + \sqrt{(1 - \cos \alpha_0)^2 + 2\hat{\gamma}} \quad (2)$$

69 Starting from this approach, a series of more refined models were developed in order to describe
70 various biological mechanisms of adhesion. Among these, the theory of multiple peeling was
71 introduced to model a system of numerous tapes loaded by a single force at a common point [17].
72 This was used in complex adhesive systems, e.g. to describe the adhesive behaviour of spider web
73 anchors [18],[19],[20]. Effects such as tape geometry, viscoelasticity or surface roughness [21], [22]
74 have also been considered, as well as bending stiffness[23]. Moreover, a so-called "hierarchical
75 shear lag model" was introduced to model hierarchical contact splitting occurring in biological
76 adhesive structures such as gecko pads [24], [25], which are suitable for active dynamic short-term
77 attachment, and other approaches have considered the effect of pretension in hierarchical
78 structures [26]. These works showed that hierarchical structuring of the surface also leads to the
79 reduction of stress concentrations and the appearance of multiple separate peeling fronts, with a
80 resulting increase in adhesive capabilities. These examples indicate the possibility of exploiting
81 various types of structures present in nature for enhanced adhesion in artificial adhesives.

82 The recent discovery of the presence of hairs in the acetabulum roof of the octopus' suckers [27]
83 (Fig. 1) suggests a revision of the model outlined in Tramacere et al. [13]. In particular, the peeling
84 model therein can be improved by adding the additional effect due to the presence of hairs on the
85 flat membrane. This work therefore aims to model the peeling process of a membrane equipped
86 with hierarchical hairs, i.e. to analyse how the hairs affect the peeling force. To do this, we apply
87 Yao's approach [28] to the geometry of an axisymmetric membrane, formulating a modified
88 expression for the work of adhesion as a function of the surface energy in a hierarchical structure
89 and deriving the corresponding detachment force of the membrane.

90

91 **2. Theoretical model**

92 *2.1 Hierarchical tape with hairs*

93 We analyse a simple tape with hairs at the interface with the substrate, as shown schematically in
94 Fig. 2, which we define as "hierarchical", meaning that its adhesive properties depend on structures

95 present at two (or more) different size scales. As a first approximation, hairs are considered to be
 96 of the same material of the tape (an incompressible soft material with $\nu = 0.5$). Furthermore, they
 97 are modelled as flat tapes of thickness t_1 , width w_1 detached length L_1 and contact length l_1 . The
 98 distance between two adjacent hairs is ρ along both x and y directions, so that $N = lw/\rho^2$ is the
 99 total number of hairs. The hairs form an angle α_1 with the substrate that is considered to be
 100 constant, and whose relation to the tape contact angle α_0 is discussed below. During the attachment
 101 and detachment phases, we do not consider bunching effects of the hairs and possible variation
 102 effects in the section of the tape. Equation (1) is valid for a simple tape without hairs. The presence
 103 of hairs on the tape surface results in an increase of the equivalent surface energy, since there is
 104 additional elastic energy stored in the hairs themselves that is “dissipated” as kinetic energy
 105 released after detachment ([25], [29]). Thus, Eq. (1) remains valid and the surface energy term can
 106 be modified to

$$107 \quad \gamma' = \gamma + \gamma_H \quad (3)$$

108 where γ' is the total surface energy, γ the surface energy of the flat tape and γ_H the equivalent
 109 surface energy due to the additional elastic energy stored in the hairs. As a first approximation, we
 110 neglected the roughness of the substrate. According to previous work [22], this roughness is not
 111 expected to influence results significantly, unless it is of the order of the microscopic features (i.e.
 112 the hairs) of the adhesive surface, which is not the case considered herein.

113 Since all hairs are assumed identical, γ_H can be considered homogeneous over the whole contact
 114 surface, and can be evaluated as:

$$115 \quad \gamma_H = \frac{l_1 + L_1}{2Ew_1^2t_1l_1} P_1^2 \quad (4)$$

116 where P_1 is the detachment force of a single hair. Using Eq. (1) to compute P_1 , we obtain:

$$117 \quad \gamma_H = \frac{Et_1}{2} \left(1 + \frac{L_1}{l_1}\right) \left(\cos \alpha_1 - 1 + \sqrt{(1 - \cos \alpha_1)^2 + \frac{2\gamma}{Et_1}}\right)^2 \quad (5)$$

118 We can now write Eq. (3) in non-dimensional form:

$$119 \quad \hat{\gamma}' = \hat{\gamma} + \frac{\gamma_H}{Et} = \hat{\gamma} + \frac{t_1}{2t} \left(1 + \frac{L_1}{l_1}\right) \left(\cos \alpha_1 - 1 + \sqrt{(1 - \cos \alpha_1)^2 + \frac{2t}{t_1} \hat{\gamma}}\right)^2 \quad (6)$$

120 Substituting this expression for the surface energy in Eq. (2), we obtain the modified non-
 121 dimensional pull-off force as:

122
$$\hat{F} = \cos \alpha_0 - 1 + \sqrt{(1 - \cos \alpha_0)^2 + 2\hat{\gamma} + \kappa_1 \left(\cos \alpha_1 - 1 + \sqrt{(1 - \cos \alpha_1)^2 + \lambda_1 \hat{\gamma}} \right)^2} \quad (7)$$

123 Where $\kappa_1 = \frac{t_1}{2t} \left(1 + \frac{L_1}{l_1} \right)$ and $\lambda_1 = \frac{2t}{t_1}$. Equation (7) thus represents the dimensionless force
 124 necessary to detach a rectangular tape equipped with hairs. Notice that the area fraction, i.e. the
 125 ratio between the contact areas of the tape with/without hairs, respectively, is usually considered
 126 close to 1, i.e., the presence of hairs does not entail a reduction/increase of the contact area[25].
 127 To illustrate the resulting behavior, we plot the peeling force \hat{F} in Fig. 3b for various angles ϵ , having
 128 chosen the following parameters: $\hat{\gamma} = 4 \cdot 10^{-4}$, $w = 10^{-2}m$, $l = 10^{-2}m$, $t = 10^{-3}m$, $w_1 = 10^{-5}m$, $l_1 =$
 129 $10^{-5}m$, $L_1 = 10^{-5}m$, $t_1 = 10^{-5}m$. As expected, the presence of a hierarchical structure, i.e. of
 130 hairs, contributes to an increase of the adhesive properties of the tape for all peeling angles due to
 131 the additional stored elastic energy, which is dissipated during delamination, with an increased
 132 effect for small angles. The peeling force decreases only slightly for increasing ϵ values. For $\alpha_0 = 0$,
 133 and $\alpha_1 = 0$, the tape is sheared parallel to the surface, and the additional dissipated energy due to
 134 the contribution of the hairs is maximum. Conversely, their decreasing effect when the peeling angle
 135 increases and tends to $\pi/2$ is consistent with the qualitative behavior observed in biological
 136 adhesion, where the peeling force needs to be maximized mainly for small peeling angles, while
 137 facilitated detachment is required at larger angles, to achieve the ON/OFF mechanism necessary,
 138 e.g. for motion in animals like geckos or insects like beetles.

139 It should be noted that in Eq.(1) and its derivations, we neglect the effect of the deformation of the
 140 substrate. In previous work, the presence of a soft substrate in peeling problems was seen to give
 141 rise to an overall increase in the detachment force, due to a wider load distribution at the interface,
 142 reducing the load concentration at the peeling line, and a decrease of the local peeling angle [22].
 143 Thus, we expect the soft substrate not to affect the predicted qualitative behavior.

144

145 2.2. Hierarchical axisymmetric membrane

146 The detachment of a single octopus' sucker can be treated as the peeling of an axisymmetric
 147 membrane [13], treated by Afferrante et al. [30], and schematically illustrated in Fig. 4a. The non-
 148 dimensional force necessary to detach the membrane is

149
$$\hat{F} = \left(\frac{32}{27} \right)^{\frac{1}{4}} (\hat{\gamma})^{\frac{3}{4}} (1 + \hat{a}) \quad (8)$$

150 where \hat{F} and \hat{a} are the dimensionless normal load and detached radius, respectively. Equation (8)
 151 predicts a linearly increasing peeling force with the membrane detached radius \hat{a} , i.e. an adhesive
 152 membrane can ideally bear an arbitrary load, provided it is large enough. In this case, the
 153 modification of γ due to the presence of hairs should be also considered. By inserting Eq. (6) in Eq.
 154 (8) we obtain the non-dimensional force necessary to detach the axisymmetric membrane equipped
 155 with hairs, although in this case the latter are assumed to be radially distributed, as shown in Fig.
 156 4b. Making the same assumptions as in the previous Section, we obtain the detachment force of
 157 the axisymmetric membrane as:

$$158 \quad \hat{F} = \left(\frac{32}{27}\right)^{\frac{1}{4}} \left(\hat{\gamma} + \kappa_1 \left(\cos \alpha_1 - 1 + \sqrt{(1 - \cos \alpha_1)^2 + \lambda_1 \hat{\gamma}}\right)^2\right)^{3/4} (1 + \hat{a}) \quad (9)$$

159 The role of the hairs for the axisymmetric membrane can be visualized in Fig. 5. In this case, we plot
 160 the peeling force versus the detached radius \hat{a} for $\hat{\gamma} = 4 \cdot 10^{-4}$, and various values of α_1 . The
 161 dependence is linear, but again, the presence of a hierarchical structure implies a considerable
 162 increase in the adhesive properties of the membrane for a given detached radius. The influence of
 163 the hairs on the peeling force decreases as the angle increases, but the \hat{F} vs. \hat{a} curves remain
 164 considerably larger than that relative to non-hierarchical case, even for large angles, e.g. $\alpha_1 = 0.4$.
 165 This is again consistent with the qualitative behavior observed in biological adhesion, where the
 166 peeling force needs to be maximized mainly for small peeling angles.

167 It should be noted that in Eq.(1) and its derivations, we neglect the effect of the deformation of the
 168 substrate. In previous work [22], the presence of a soft substrate in peeling problems was seen to
 169 give rise to an overall increase in the detachment force, due to a wider load distribution at the
 170 interface, reducing the load concentration at the peeling line, and a decrease of the local peeling
 171 angle. Thus, we expect the soft substrate not to affect the predicted qualitative behaviour.

172

173 *2.3 Additional levels of hierarchy*

174 The previous model can be extended to additional levels of hierarchy, as illustrated schematically in
 175 Fig. 3a. In this case, Eq. (3) can be extended as follows:

$$176 \quad \gamma' = \gamma + \gamma_1 + \gamma_2 + \dots + \gamma_n \quad (10)$$

177 where γ_1 coincides with the previously introduced γ_H . The total force necessary to detach this type
 178 of tape/membrane can be computed as previously, by recursively adding the terms relative to the

179 appropriate hierarchical level. For example, the second level of hierarchy can be described by adding
 180 to $\hat{\gamma}_1$ another term of the form

$$181 \quad \hat{\gamma}_2 = \kappa_2 \left(\cos \alpha_2 - 1 + \sqrt{(1 - \cos \alpha_2)^2 + \lambda_2 \hat{\gamma}} \right)^2 \quad (11)$$

182 where κ_2 , λ_2 and α_2 are analogous to the first level parameters κ_1 , λ_1 and α_1 , respectively.
 183 Analogous expressions can be written for $i > 2$. In order to compute the κ_i and λ_i and α_i parameters,
 184 it is necessary to consider the geometry (i.e. geometry and contact angles at the various hierarchical
 185 levels) of the new system. The approach outlined in the previous sections can then be adopted to
 186 determine higher order surface energy values γ_i to the adhesive energy due to the additional
 187 hierarchical levels, and the corresponding peeling force. Given the small bending stiffness of the
 188 tapes at the various hierarchical levels, the angle variations from one hierarchical level to the next
 189 are in all cases small. Therefore, the corrections decrease in magnitude for an increasing number of
 190 levels, i.e. the adhesive energy and force values do not diverge. This can be seen in results illustrated
 191 in Fig. 6. Here, we consider as previously a perturbation ϵ on the contact angle from one level to the
 192 next, and assume for simplicity that the perturbation is of the same order for each level, i.e.
 193 $\cos \alpha_{i+1} = \cos(\alpha_i + \epsilon)$, $\forall i$. Thus, an increase of the hierarchical level also implies an increase in
 194 the overall perturbation on the initial peeling angle α_0 . Figures 6a and 6b show the effect of an
 195 increasing number of hierarchical levels for the \hat{F} vs. α_0 and \hat{F} vs. \hat{a} plots in the case of a hierarchical
 196 tape and a hierarchical axisymmetric membrane, respectively. For 3 levels of hierarchy, at $\alpha_0 = 0.1$
 197 the adhesive force is increased by approximately 6 times with respect to the non-hierarchical case.
 198 It is apparent that the main increase takes place for the first hierarchical levels, as is clearly visible
 199 in Figs. 6c and 6d, where \hat{F} is plotted as a function of the number of hierarchical levels for fixed θ
 200 and \hat{a} values, again in the case of a hierarchical tape and a hierarchical axisymmetric membrane,
 201 respectively. We can compute the gain in adhesive force at level i by dividing F_i by the force at level
 202 $i-1$ (F_{i-1}).

$$203 \quad Gain = \frac{F_i}{F_{i-1}} \quad (12)$$

204 Plotting the gain values versus the hierarchical level for the simple tape and the axisymmetric
 205 membrane (Fig. 6 e, f), we see that after 2 or 3 levels, there is no further significant gain. Therefore,
 206 we can state that 2 or 3 hierarchical levels are sufficient to optimize adhesive force. A further
 207 increase in hierarchical levels could be detrimental, since the smallest features would become of
 208 the order of the characteristic size of the substrate roughness, leading to a decrease of adhesion

209 [22]. This is consistent with observations on biological adhesive structures found in nature, such as
210 beetle legs or gecko toes [16][31], which typically display 2 or 3 levels of hierarchy. In the case of
211 octopus's sucker membranes, hairs appear to be present at most at three levels of hierarchy.

212

213 **Conclusions**

214 Understanding of the effect of a layer of hairs on the adhesive properties of octopus' suckers is
215 important for the design of artificial suction cups with improved adhesion for various applications,
216 such as smart-skin attachable skin patches [32] or biorobotic adhesive discs [33]. Here, we have
217 evaluated the effect of hierarchical structure, i.e. the presence of hairs, on the adhesion and
218 detachment of a simple tape and of an axisymmetric membrane, in order to gain insight into the
219 adhesion mechanism of octopus' suckers (in particular the detachment of the acetabulum roof from
220 the acetabulum wall). The model is based on a number of simplifying assumptions, e.g. that there is
221 no hair bunching and that the peeling angle does not vary significantly between structures at one
222 hierarchical level and those at the next. Furthermore, delamination is assumed to take place from a
223 rigid substrate, whereas the real biological tissue considered is soft and relatively deformable.
224 However, these assumptions are not expected to qualitatively modify the analysis herein.

225 Results for the simple tape case indicate that the presence of hairs can improve the adhesive
226 properties by more than 30% at small peeling angles, with the effect decreasing for larger angles.
227 This is consistent with observations on biological adhesion, where typically adhesive forces need to
228 be enhanced at small peeling angles. The main parameter determining this increase is the initial
229 detached length of the hairs, which has an upper limit in lengths for which there is an onset of
230 bunching effects. The detachment force for an axisymmetric membrane also increases in the
231 presence of hierarchical structuring. We show that the model can be easily extended to the analysis
232 to multiple levels of hierarchy. Here, results indicate that the first hierarchical levels are the ones
233 that contribute more to an increase in adhesive force. In terms of convergence, we find that after
234 the third level of hierarchy there is no longer a significant change in peeling force.

235 This paper provides a possible explanation for the role of the hairs in octopus' suckers, correctly
236 accounting for their role in determining the ON/OFF behavior during adhesion. Currently, further
237 studies are under way to evaluate other possible functions of these hairs (e.g. sensing) that could
238 be fundamental to the octopus functionality. Our work can also help the design of artificial suction

239 cups by providing a model that predicts the potential benefits of a hierarchical surface in terms of
240 improved and angle-dependent adhesive properties.

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252 **Author Contributions**

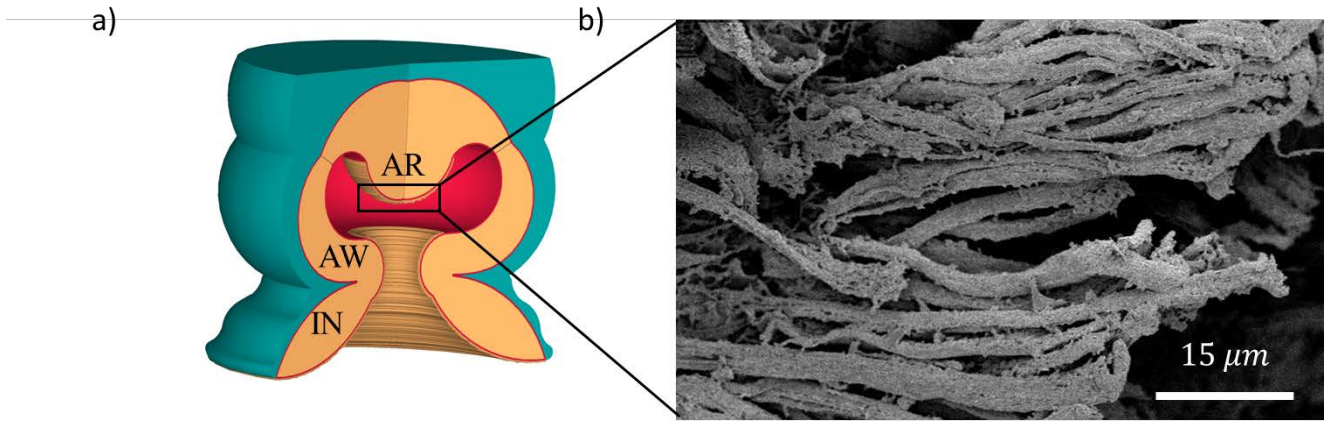
253 N.M.P designed the study and supervised the work, G.G. wrote the first draft of the manuscript and
254 generated diagrams supervised also by F.B.. All the authors finalized the manuscript.

255 **Additional Information**

256 **Competing Interests:** The authors declare that they have no competing interests.

257 **Data availability:** The authors declare that the data supporting the findings of this study
258 are available within the article and its supplementary information files.

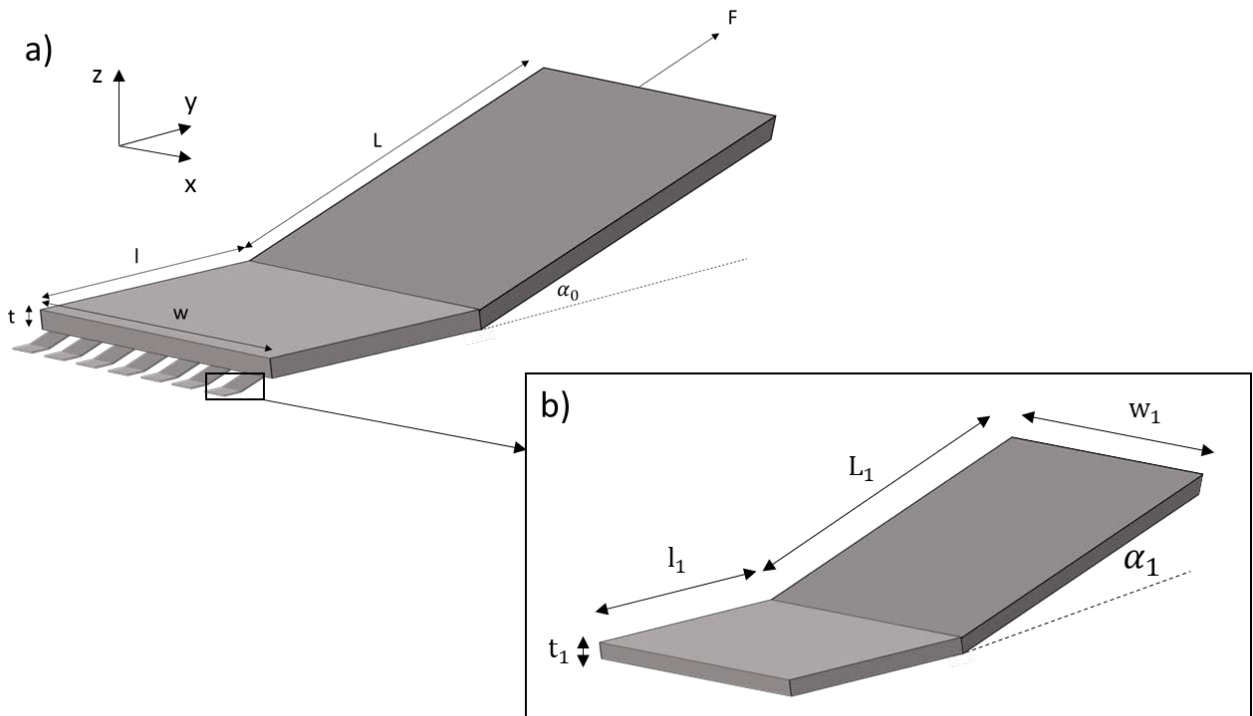
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260

261 *Figure 1: a) Schematic of the octopus' sucker: Acetabular Roof (AR), Acetabular Wall (AW) and Infundibulum*
 262 *(IN). b) Hairs present on the surface of the AR that is attached to the AW during adhesion.*

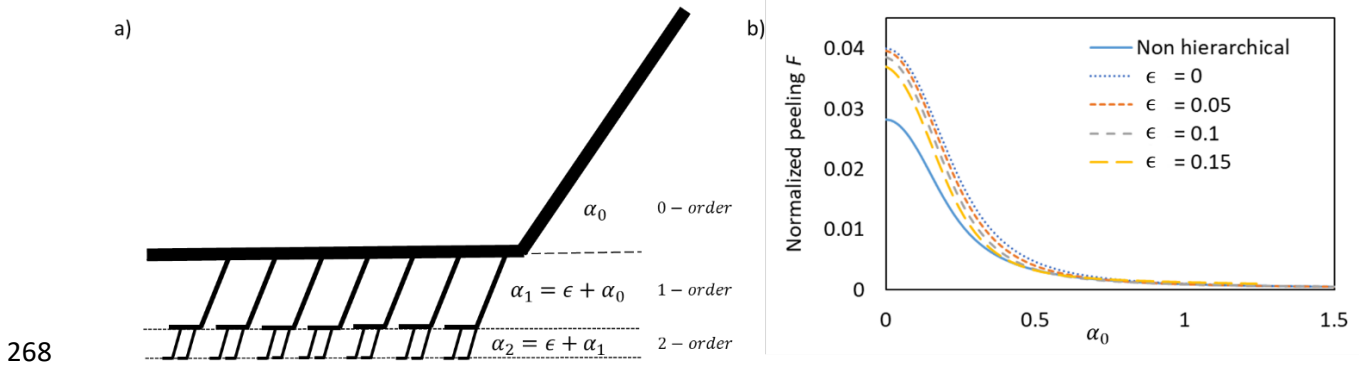
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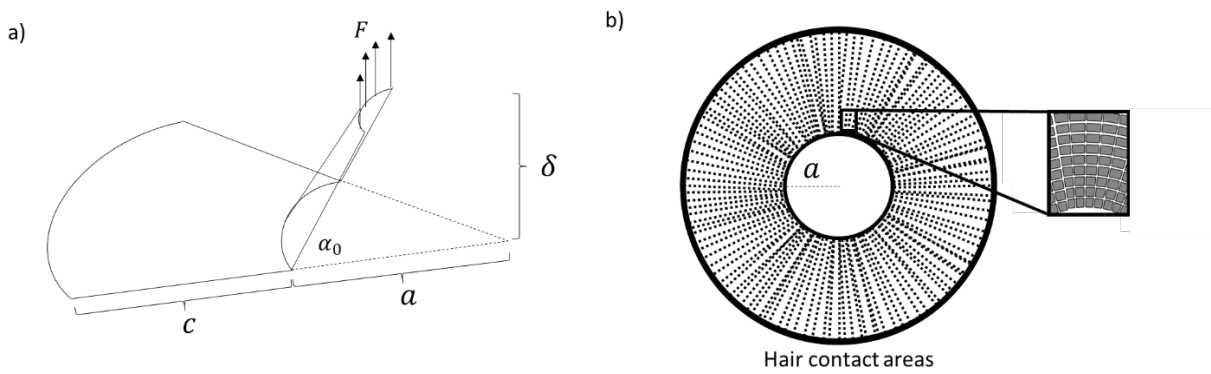
265 *Figure 2: a) Schematic of the peeling of an elastic tape equipped with hairs; b) schematic of a single*
 266 *second-level tape (hair).*

267



269 *Figure 3: a) Schematic of hierarchical levels up to the second order. b) Normalized peeling force vs. peeling*
 270 *angle (Eq. (7)) for different ϵ parameter values and $\hat{\gamma}=4 \cdot 10^{-4}$.*

271

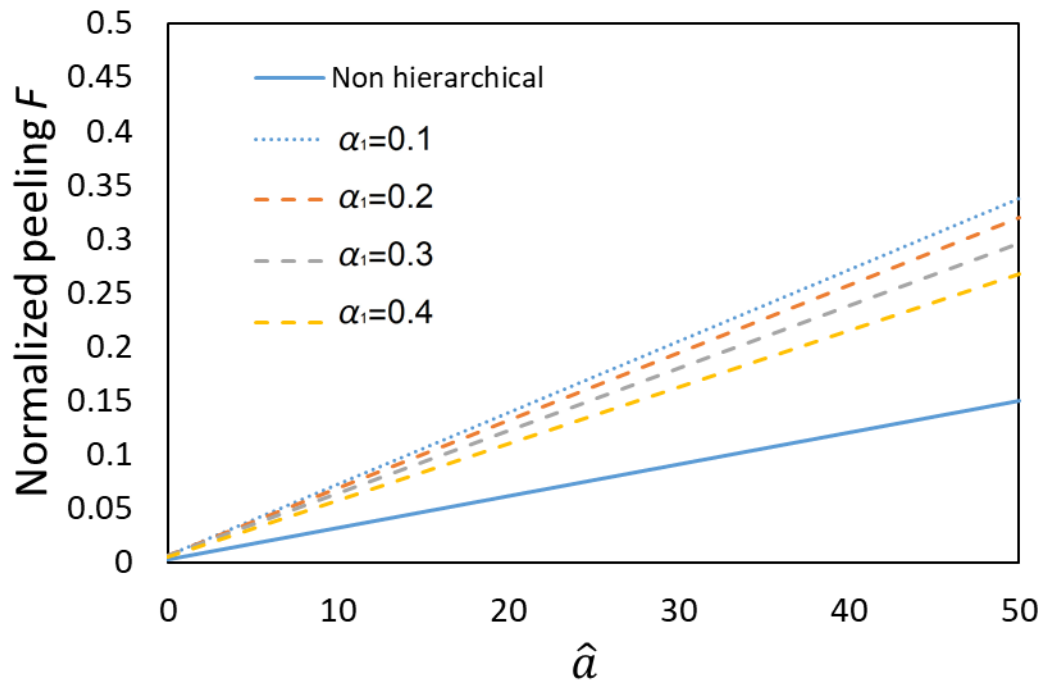


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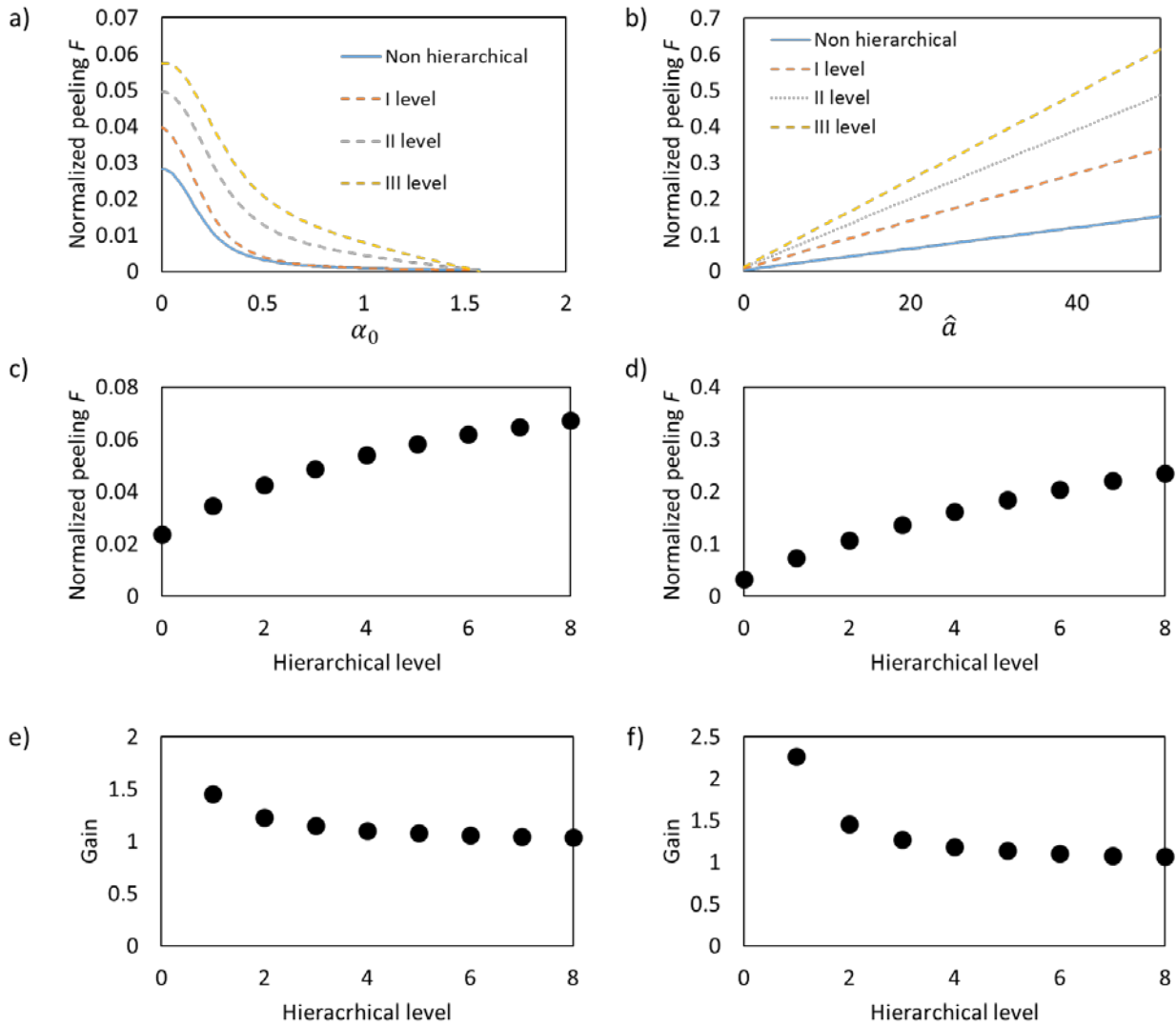
274 *Figure 4: a) Schematic of the peeling of an axisymmetric membrane and b) schematic the contact region*
 275 *between the hairs of the axisymmetric membrane and the substrate.*

276



278

279 Figure 5: Peeling force for an axisymmetric membrane vs. detached radius \hat{a} (Eq. (9)) for different α_1 values
 280 ($\hat{\gamma} = 4 \cdot 10^{-4}$).



281

282 *Figure 6: a) Normalized peeling force vs. peeling angle for increasing hierarchical levels in the case*
 283 *of a simple tape ($\varepsilon = 0.05$); b) Normalized peeling force vs normalized detached radius for*
 284 *increasing hierarchical levels in the case of an axisymmetric membrane ($\varepsilon = 0.05$ and $\alpha_0 = 0.1$). c)*
 285 *Normalized peeling force as a function of number of hierarchical levels in the case of a simple tape*
 286 *($\varepsilon = 0.05$ and $\alpha_0 = 0.1$). d) Normalized peeling force as a function of number of hierarchical levels in*
 287 *the case of an axisymmetric membrane ($\varepsilon = 0.05$, $\hat{a} = 10$). e) Plot of the gain (Eq. 12) versus the*
 288 *hierarchical level for the simple tape and f) the axisymmetric membrane.*

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