Using Boolean Networks to Model Post-Transcriptional Regulation in Gene Regulatory Networks

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Abstract

Gene Regulatory Networks (GRNs) model some of the mechanisms that regulate gene expression. Among the computational approaches available to model and study GNRs, Boolean Network (BN) emerged as very successful to better understand both the structural and dynamical properties of GRNs. Nevertheless, the most widely used models based on BNs do not include any post-transcriptional regulation mechanism. Since miRNAs have been proved to play an important regulatory role, in this paper we show how the post-transcriptional regulation mechanism mediated by miRNAs has been included in an enhanced BN-based model. We resort to the \textit{miR-7} in two \textit{Drosophila} cell fate determination networks to verify the effectiveness of miRNAs modeling in BNs, by implementing it in our tool for the analysis of Boolean Networks.

Keywords: miRNA, Gene Regulatory Networks, post-transcriptional regulation, Boolean Networks, Complex Systems, Network Analysis

1. Introduction

The genome of an organism plays a crucial role in regulating several cellular processes. With few exceptions, every cell in an organism contains the same genetic material. This implies that, after sequencing the whole genome of several living organisms \cite{1}, and after knowing the functional meaning of thousands of proteins encoded by these genomes \cite{2}, one of the principal remaining challenges is to understand the mechanisms that regulate gene expression within the genome \cite{3}.

The regulation of gene expression is achieved through genetic regulatory systems organized in networks of interactions between genes, proteins and other cellular components. In order to understand complex genetic regulatory networks, a concerted effort among experiments, modeling, and theory is required. In particular, researchers require automated formal modeling techniques and appropriate analysis tools.

Several computational approaches have been proposed and developed in literature (\cite{4} and further references therein) to model Gene Regulatory Networks (GRNs). They include partial/ordinary differential equations \cite{5, 6}, linear models \cite{7}, Bayesian networks \cite{8, 9}, Boolean networks \cite{10, 11} and Petri nets \cite{12}. They can be divided into two groups: 1) \textit{discrete-state} models and 2) \textit{continuous-state} models. Discrete-state models assign a small number of discrete states to each node of the network, avoiding intermediate expression levels. Regulatory interactions between nodes are described by logical functions. Bayesian networks, Boolean networks, and Petri nets belong to this group. Instead, continuous-state models, assume the state of each node of the network to be a continuous function in time of the expression of the input components. Its evolution is modeled resorting to differential rate equations. Partial differential equations, (nonlinear) ordinary differential equations, and linear models belong to this group. Their main limitation consists in the difficulty of properly modeling all biochemical reactions governing the interaction among the nodes of the network.

Boolean Networks (BN), introduced by Kauffman \cite{10}, have been proved successful in modeling real regulatory networks (e.g., see \cite{13, 14, 15, 16, 17, 18}). In a BN the state of a biochemical entity, i.e., a gene, is described by a Boolean variable. The transition from one state to another state is computed by means of a Boolean function of the states of other genes in the network. Transitions between states are \textit{deterministic}, which means that, given an initial state, the next state is always the same. Although the approach seems to set a strong simplification compared to reality, BNs enable to study high-level properties of complex networks (e.g., robustness to background noise, behavior under different initial conditions, etc.).

Recent researches suggest that several realistic biological problems may be analyzed resorting to the BN formalism. In this cases, BNs allow for a simulation of a real biological GRN’s dynamic behavior (e.g., the Drosophila case),
emphasizing the functional relevance of the resulting network topology [19]. Biologist are thus able to characterize the network and evaluate its dynamics, identifying topological elements such as feedback circuits, i.e., cascades of regulatory interactions, negative circuits that buffer gene effects, and positive circuits that may constitute developmental switches enabling alternative developmental pathways [20, 21]. Moreover, it is possible to study properties of the biological GRN by computing and analyzing the attractors of the network. An attractor is a single state or a set of states towards which a system tends to converge over time [22, 23].

Most published BN based models focus on high-level gene/gene or gene/protein interactions, neglecting post-transcriptional regulatory activities carried out by small non-coding RNA sequences, such as micro RNA (miRNA).

In this paper we discuss how miRNAs and post-transcriptional regulatory interactions can be modeled resorting to BNs. The proposed BN based post-transcriptional model has been implemented in a software tool able to simulate a BN and to compute the attractors of the network taking into account post-transcriptional activities [24].

We exploited the developed model and the related tool to analyze two real networks related to Drosophila taken from [25]. Results obtained from the network analysis have been then compared with experimental results discussed in [25] to show the capability of the proposed model to provide interesting insights related to the modeled biological process.

2. Background

Modeling general aspects of GRNs, such as genes’ interactions, dates back to the end of 1960s. In [10], Kauffman considered an idealized representation of a typical gene network. The assumption is that genes are equivalent entities, and their interaction forms a directed graph in which each gene receives inputs from a fixed number of selected genes. The set of entities populating the network has been recently extended including gene products (e.g., proteins) by [18]. The state of each regulatory entity, i.e., a gene or a protein, is represented as a Boolean value, either 1, indicating the presence of the entity (e.g., a gene is expressed), or 0 indicating its absence (e.g., a gene is not expressed).

Given a set of N entities, the state of the GRN is represented by a Boolean vector $\hat{X} = [x_1, \cdots, x_N]$, leading to a state space of $2^N$ states. Each component $x_i$ of $\hat{X}$ is described using a Boolean function $f_i (x_i = f_i (\hat{X}))$.

To make the model computationally acceptable, the f behavior of a BN is usually simulated in a synchronous way, i.e., all entities of the network update their state together [13, 26]:

$$\hat{X}^{t+1} = \hat{F} (\hat{X}^t)$$  \hspace{1cm} (1)

where $\hat{X}^{t+1}$ is the next GRN state and $\hat{F}$ is the vector of all functions $f_i$ that map the transition of a single entity from the current state to the next one.

During the simulation of the network dynamics, an initialization state sets all nodes of the network to a known configuration. The network state is then continuously updated by repeatedly evaluating all boolean functions describing the network until a steady state or a state cycle is reached. The sequence of states traversed by a BN during the simulation of its dynamics forms a trajectory of the system, while the final steady state or the final state cycle of the trajectory form a point or a cyclic attractor, respectively. Each network is associated with a set of point and cyclic attractors, depending on its dynamics on different initialization states. In case of a point attractor, the system’s state freezes whenever the network enters the attractor, and it is unable to perform further transitions unless external perturbations are applied. Differently, cyclic attractors show a cyclic behavior of the system. Once a trajectory falls into one of the states belonging the attractor, the system keeps cyclically moving among the attractor’s states.

To properly study the network dynamics all possible initialization states should be simulated. The set of initial states leading to an attractor is called basin of attraction [27]. Attractor’s properties (length of cyclic attractors, basins dimensionality, trajectory lengths, etc.) are commonly studied in order to infer high-level properties of a GRN [18, 27].

Figure 1a gives a simple example of a BN. It contains three entities (e.g., $x_1$, $x_2$, $x_3$) whose next state functions are defined as follows:

$$x_1^{t+1} = \neg x_3^t$$
$$x_2^{t+1} = x_1^t \land x_3^t$$
$$x_3^{t+1} = x_2^t$$

Figure 1b shows the state space of this network. Each node represents a possible network state whereas edges represent legal state transitions for the network.

3. Models and Methods

3.1. Extending the Gene Protein/Product Boolean Network Model

Functional studies indicate that miRNAs participate in the regulation of almost every investigated cellular process like, for instance, cell metabolism, signal transduction, cell differentiation, cell fates and so on [28, 29, 30]. miRNAs regulate gene expression post-transcriptionally by interfering either with a target mRNA’s translation or stability [31]. Moreover, further studies show that they can modulate mRNA-protein interactions, and suppress protein synthesis, although the mechanistic details are still poorly understood.

The interaction between mRNAs and proteins is well modeled by the Gene Protein/Product Boolean Network
Figure 1: Example of a simple boolean network involving three genes ($x_1$, $x_2$ and $x_3$) together with the related space state.

(a) The boolean network represented as a directed graph. Each node of the graph represents a gene while arcs represent gene interactions.

(b) The state space of the network represented as a directed graph where each node represents a state of the network while arcs represent valid state transitions.

(GPBN) model proposed by Graudenzi et al. in [18], which is a generalization of the classical RBN model. In a GPBN, gene to gene interactions are mediated by the synthesis of proteins and other products. However post-transcriptional regulation carried out by miRNAs is still not fully considered. Figure 2 depicts a very simple example of GRN modeling a cellular regulatory activity that includes all entities that need to be considered in order to extend the GPBN model to include post-transcriptional regulation mechanisms. According to the example, G1 and G2 are transcribed into two mRNA molecules (mRNA1 and mRNA2); P1 and P2 are the resulting proteins. P2 works as an upstream promoter of gene G3, i.e., G2 is a transcription factor of gene G3. miRNA1 (still a product of G1) acts as a post-transcriptional repressor of mRNA2, which results in a translational repression of P2 and therefore in an inhibition effect on gene G3.

To properly model this post-transcriptional interaction, following the GPBN principles, we extended the interaction between genes (G1 and G2 of the example) by explicitly introducing their related products (as previously done for continuous models [32, 33]), thus, including miRNA1 as a product of G1.

Figure 3 shows an extended BN representing the proposed regulatory example. For the sake of simplicity, the translational process that leads to the protein production, starting from the related mRNA molecule, has not been explicitly modeled. Nevertheless, if necessary, the model can be easily extended by adding all actors required to precisely modeling all processes. Networks nodes are depicted with different symbols to identify: (1) genes (circular nodes), (2) mRNA-Protein pairs (rectangular nodes), and (3) miRNA (rhomboidal nodes).

In order to properly model the post-transcriptional regulation mechanisms, the set of boolean functions of each transcriptional product targeted by a miRNA (e.g., the mRNA2_P2 node) must be carefully designed. Post-transcriptional regulation acts at mRNA level, hence, considering the final protein production, it has higher priority compared to gene expression activity. In terms of Boolean
functions, this can be modeled by placing the miRNA expression state in Boolean AND with the mRNA expression state.

When introducing gene products such as proteins and miRNAs in a BN model, the synthesis and decay times for a given product must be taken into account. In our model, we considered an unitary synthesis and decay time for all entities according to the GPBN model. Therefore, the change of state of a gene at time $t$ influences all its products at time $t + 1$.

Although the introduction of miRNA entities into the BN makes it possible to account for their post-transcriptional activity into the dynamics of the system, it is not enough to properly model the whole post-transcriptional activity. If not properly constrained, users may select initialization states for the simulation of the network (that can now include combinations of genes and products) that may be biologically not valid. To deal with illegal states, the description of the BN is expanded, including a set of conditions identifying all illegal states of the network. Once a biologically valid initialization state is selected, if well designed, the network itself will avoid to evolve into biological illegal states. In Figure 3 these conditions are represented by the Boolean equations C1, C2, and C3. Every time an initial state of the network is considered the three conditions must be evaluated. A state is considered legal if all conditions return zero, illegal otherwise. As an example, let us consider G1 and the related protein mRNA1_P1. The protein can be synthesized only if the related gene has been expressed. So, any state in which mRNA1_P1 is equal to 1 (expressed), while G1 is equal to 0 (not expressed) is actually an illegal state.
3.2. Including the model into the Boolean Network Toolkit

We implemented the GRN model presented in Section 3.1 into a software tool able to analyze the network dynamics by computing the network attractors [24]. The actual implementation is based on the Boolean Network Toolkit (BNT) presented in [26]. The BNT implements the BN direct graph using adjacent lists to optimize access speed and reduce memory allocation. The core is entirely built upon the BOOST C++ Library [34], which provides very efficient and cross platform libraries.

Algorithm 1 shows the overall attractors search process that is an iterative process involving probes iterations. Probes is the number of initial states from which the network is simulated to identify its attractors. Whenever the number N of network nodes leads to a state space too large to be exhaustively analyzed, it is possible to keep the computation time under control by selecting a reduced set of randomly or user selected initial states for the network. At each iteration, a candidate initial state is selected and its validity against the transcriptional and post-transcriptional constraints is checked. If the state is legal, the network dynamic is simulated to search for an attractor on the selected trajectory.

```
Data: the Gene Regulatory Network
Result: a Set which contains the attractors
input : probes the number of initial condition to test
for all the probes do
    σ ← generateRandomState;
    isValid ← isValidBooleanState(σ,C);
    if isValid = true then
        anAttractor ← findAttractor(σ);
        grnAttractorSet.add(anAttractor);
    else
        continue;

Algorithm 1: GRN attractors searching algorithm.
```

The isValidBooleanState function (see Algorithm 2) performs the initial state validity check. Given the initial state to analyze and the set of constraints as input, this function evaluates the state against each constraint. If at least one of the constraints is true, it means the given state is not valid and it needs to be discarded. Otherwise, a true value is returned.

Eventually, the tool exports all result of the network analysis (e.g., attractors, state space, trajectories, etc.) using the XGMMML format, which is ready to be visualized and further analyzed with visualization tools such as Cytoscape [35].

```
Result: if the state is admissible or not
input : X the state to check
input : stateConstraintSet the collection of constraints for the given network
output: true if the state being checked is admissible
output: false otherwise
notValid ← false;
foreach anInvalidSchema in stateConstraintSet do
    notValid ← isValid OR anInvalidSchema (X) ;
if notValid = true then
    return false ;
else
    return true ;

Algorithm 2: Algorithm for checking the validity of an initial state.
```

4. Results and Discussion

We tested our model against two real networks discussed in [25]. Both networks analyze the role of miR-7 discussed in [25]. Both networks analyze the role of miR-7

in Drosophila. The two networks have been modeled resorting to the proposed BN post-transcriptional model and then analyzed by computing the exhaustive set of attractors and trajectories. Results obtained from the network analysis have been finally compared with the results reported in [36] that hypothesize a stabilizing role of miR-7 against perturbations that would change the cell fate in terms of development. This comparison provides an interesting example of the type of analysis and results that the proposed model is able to support.

The simulated networks described using the input formalism of our tool, as well as all outputs produced from the network analysis in standard XGMMML format have been provided as additional material to this paper 2. Moreover the source code of the tool used for the analysis, as well as a legacy Cytoscape plugin that can be used to visualize the outputs produced by the tool can be freely downloaded at http://www.testgroup.polito.it/index.php/bio-menu-tools/item/208-boolean-regulatory-network-simulator.

4.1. Networks description

The two considered GRNs that regulate the determination of photoreceptor cells, proprioceptor organs, and olfactory organs in Drosophila are:

1. Photoreceptor determination network (Figure 4a): in this network miR-7 acts in a coherent feed-forward loop cooperating with Pnt-P1 in silencing YAN to create stability against fluctuations of Pnt-P1. This behavior is common for guaranteeing that a cell’s fate change is not spontaneously induced or reverted [37], admitting only oriented transitions from the state YAN ON to the state YAN OFF in which YAN itself

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2We have been unable to submit the supplemental material through the EES submission system. For the review process, this material is therefore available at the following URL: http://orion.polito.it/tmp/JOCS-D-13-00019-AdditionalMaterial.zip.
is fully degraded. After YAN degradation, possible Pnt-P1 fluctuations will not lead to further YAN ON states.

2. Sensory organ precursor (SOP) determination network (Figure 4b): in this network mir-7 belongs to an incoherent feed-forward loop. This type of network motif leads to an accelerated and transient pulse to downstream genes expression [38]. The overall effect is a network in which fluctuating peaks of Atonal (ATO) would result in transient pulses of ATO repression by EsplC. Vice versa the sustained increase of ATO would result in sustained repression of EsplC by miR-7 and a corresponding stabilization of ATO [25].

4.2. Network modeling and attractor analysis

Both networks presented in Figure 4 have been partially redesigned in order to fit our extended GPBN model in which miRNAs target proteins nodes and miRNAs are produced thanks to the expression of the related hosting genes. Figure 5 graphically shows the basic rule exploited during the BN post-transcriptional redesign process. According to [20], if the transcription/translation is active, mRNAs/proteins are synthesized in a one time step. Thus, if the network includes a miRNA node targeting a gene, a new protein node must be inserted in the network. The protein node must be connected to the parent gene node and targeted by the miRNA. Also all gene products (out-going edges) must be re-arranged accordingly. These extensions respect the assumption that both transcription factors and proteins undergoing post-transcriptional modifications decay in a one time step if their mRNAs are not present [20]. Finally, also the miRNA host gene (if not present) must be explicitly inserted in the network.

While this redesign phase has been performed manually on-going work is being carried out to make it automatic within our simulation tool.

Figure 6a and Figure 7a show the redesigned networks considered in our analysis. It is worth to point out here that there is no evidence in [39] regarding the way genes cooperate in enhancing/silencing their products. In order to take into account different options two variants of each network have been considered assuming that:

- a gene is expressed when all its parent nodes are concurrently expressed (ON): this condition models enhancer complexes composed by multiple proteins that cooperate together. Resulting boolean functions describe this condition as a set of terms computed by the \( \land \) operator.
- a gene is expressed when at least one of its parent nodes is expressed (ON): this represents the behavior of multiple transcription factors that autonomously induce the expression of their targets. Resulting boolean functions describe this condition as a set of terms computed by the \( \lor \) operator.

In both networks, these two options have been modeled by introducing the generic Boolean operator \( <\text{op}>\in\{\land,\lor\} \).

Moreover, in order to highlight the contribution of mir-7 to the behavior of the network, we compared the two variants of the redesigned networks reported in Figure 6a and Figure 7a, with two equivalent networks in which the...
miR-7 node and related edges have been removed (Figure 6b and Figure 7b).

We analyzed each network using our tool, collecting all attractors, trajectories and basins of attraction. Interestingly, the two options introduced by the \( \langle op \rangle \) operator lead to different behaviors of the network, which still maintains the same topology. In each simulation the exhaustive set of initial states has been analyzed with a reasonable average computation time (~20-30 msec to analyze each network).

The next sections discuss results obtained from the network analysis. The considered networks have been simulated under several assumptions in order to better understand the properties of the computed attractors, along with the role of miR-7 in the network.

4.2.1. Photoreceptor determination network

Due to the network topology, miR-7 is expected to cooperate with ERK and Pnt-P1 in maintaining the network steady against YAN fluctuations.

Given this premise, the comparison of the network attractors between the network with miR-7 (Figure 6a) and the one without miR-7 (Figure 6b), does not show any difference when the \( \langle op \rangle \Rightarrow \wedge \) option is selected. The set of common attractors with and without miR-7 is reported in Table ???. In fact, the mandatory concurring presence of all silencers of YAN somehow masks the role of the miRNA. This is not surprising because, based on their discrete nature, BNs perform a qualitative analysis instead of measuring the gene expression rate from a quantitative perspective. This simplification makes the analysis of complex network feasible, but it possibly masks certain configurations and their intermediate equilibrium. In this scenario, the antagonist roles of Pnt-P1 and miR-7 concur in preserving the expected pathway behavior (e.g., the complete degradation of YAN), possibly masking the miR-7 fine tuning effect.

Things change when the network is analyzed under the \( \langle op \rangle \Rightarrow \lor \) option. In this case, the tool returns 3 point attractors common in both the network with miR-7 (Figure 6a) and the one without miR-7 (Figure 6b) plus a set of specific attractors. The set of common attractors is reported in Table ???. These attractors are not discussed here since they do not show any difference between the two considered networks. We identified three point attractors specific of the network with miR-7 and one point attractor specific of the network without miR-7:

- Attractors of the network with miR-7 (see Table 1): the 3 attractors confirm the expression of miR-7 and the expected degradation of the YAN protein. Interestingly, the three attractors show the suppression of YAN\( _P \) by the miRNA, with no regards to the expression of the signaling genes (Notch, EGFR), and the YAN’s antagonists (ERK, Pnt-P1).

- Attractors of the network without miR-7 (see Table ??): the point attractor shows no degradation...
of the YAN protein even if both YAN’s antagonists (ERK, Pnt-P1) are expressed. The ectopic expression of YAN_P may confirm the regulatory role of miR-7 for the network’s stabilization.

4.2.2. SOP determination network

The topology of the SOP determination network suggests two behaviors: 1) transient pulses of downstream genes expression, 2) specific polarization of upstream genes that leads to the stabilization of the entire network.

Let us first consider the network attractors under the $<op> = \land$ assumption. The network with miR-7 (Figure 7a) and the one without miR-7 (Figure 7b) manifest a set of five point and four common cyclic attractors (reported in Table ??). In all of them, EspIC and its protein are suppressed, regardless the state of Notch. In fact, the suppression of EspIC is an obvious outcome because it needs the simultaneous expression of both Notch and ATO (since they are joined with the $<op> = \land$). The Notch silencing also leads to a similar result in the set of common cyclic attractors: all of them show transient pulses of ATO only when the Notch chain is turned OFF.

Moreover, the network with miR-7 introduces one specific point attractor (see Table ??). The point attractor leads to the complete degradation of EspIC_P, while EspIC, miR-7 and ATO are still expressed. Since only the EspIC protein is silenced whilst its encoding gene remains expressed, this attractor confirms the regulatory role of the miRNA.

Instead, when considering the $<op> = \lor$ assumption, the network with miR-7 (Figure 7a) and the one without miR-7 (Figure 7b) manifest two common point attractors only. They show two outcomes compatible with expectancies in [39]: i) when Notch is ON and EGFR is OFF, miR-7 is also OFF, avoiding the silencing of EspIC_P; ii) when both Notch and EGFR are ON, miR-7 is expressed, correctly degrading EspIC_P. The analysis of the basins of common attractors provides additional information to understand these two behaviors: the size of each basin decreases when miR-7 is introduced within the network, supporting the hypothesis that the presence of the miRNA makes the network less prone to enter in one of the common attractors.

Together with the common attractors (see Table ??) the network also includes now a set of specific attractors:

- **Attractors of the network with miR-7 (see Table ??):** the set of attractors is composed of three point attractors. All of them show the expected stabilization of the network. The sustained expression of ATO and miR-7 causes the degradation of EspIC_P, regardless the fact that EGFR and Notch are expressed or not.

- **Attractors of the network without miR-7 (see Table ??):** The three point attractors show the same pattern of expression: ATO, ATO_P, EspIC, and EspIC_P are always turned ON, regardless of the expression of both EGFR and Notch. The ATO expression seems only regulated by Sense because it remains still turned ON even if its signaling chain is turned OFF. Looking at the set of four cyclic attractors, all of them share the degradation of EGFR and the expression of Notch. This condition leads to the expression of both EspIC and EspIC_P, unexpected in [39]. The pulses of ATO are only driven by the buffer effect of Sense.

For this network only, it can be easily noticed that the $<op> = \lor$ and $<op> = \land$ option produces results that are more coherent with experimental findings reported in the literature.

Overall, given the different attractors obtained with the $<op> = \lor$ and $<op> = \land$ operators, this preliminary work highlights how the description of the boolean function of any given node plays a crucial role when dealing with GRNs: a better estimate of the operator should eventually lead to a more reliable set of attractors.

5. Conclusions

In this paper we discussed an extended BN model to account for post-transcriptional regulation in GRN simulation. Thanks to this extended model, we discussed the set of attractors of two biologically confirmed networks, focusing on the regulatory role of miR-7. Attractors have been compared with networks in which the miRNA was removed. The central role of the miRNA for increasing the
network stability has been highlighted in both the networks, confirming the cooperative stabilizing role of miR-7.

The enhanced BN model presented in this paper is only a first step towards a more realistic analysis of the high-level functional and topological characteristics of GRNs. Resorting to the tool facilities, the dynamics of real networks can be analyzed. Thanks to the extended model that includes post-transcriptional regulations, not only the network simulation can be more reliable, but also it can offer new insights on the role of miRNAs from a functional perspective, and this improves the current state-of-the-art, which mostly focuses on high-level gene/gene or gene/protein interactions, neglecting post-transcriptional regulations.

Due to its discrete nature, the BN model may still neglect some regulatory fine adjustments. However, the largest number of the computed attractors, now including miRNAs, still represents meaningful states of the network. The simple glimpse into the complexity of the network dynamics, that the toolkit is able to provide, could be used not only as a validation of in vitro experiments, but as a real System Biology tool able to raise new questions and drive new experiments.

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URL http://books.google.it/books?id=9bZbDFrO9QC
URL http://dx.doi.org/10.1016/j.semcdb.2009.07.003
URL http://dx.doi.org/10.1371%2Fjournal.pbio.1000238
Table 1: Unique attractors for Photoreceptor Determination Network using the operator $< op >= \wedge$ with miR-7. The Table shows 3 point attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

Table 2: Unique attractors for Photoreceptor Determination Network using the operator $< op >= \wedge$ without miR-7. The Table shows 1 point attractor. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

Table 3: Common attractors for Photoreceptor Determination Network using the operator $< op >= \wedge$. The Table shows 4 point attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

Table 4: Common attractors for Photoreceptor Determination Network using the operator $< op >= \vee$. The Table shows 3 point attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.
Additional files

- PhD_w_mirna_AND_results.zip: Photoreceptor determination network with \( miR-7 \) present and \( < op >= \wedge \) option analysis and simulation results.
- PhD_w_mirna_AND.net: Photoreceptor determination network with \( miR-7 \) present and \( < op >= \wedge \) option description file.
- PhD_w_mirna_OR_results.zip: Photoreceptor determination network with \( miR-7 \) present and \( < op >= \vee \) option analysis and simulation results.
- PhD_w_mirna_OR.net: Photoreceptor determination network with \( miR-7 \) present and \( < op >= \vee \) option description file.
- PhD_w_mirna.constraints: Photoreceptor determination network with \( miR-7 \) present constraint file to be used both for simulation with \( < op >= \wedge \) and \( < op >= \vee \) options.
- PhD_wo_mirna_AND_results.zip: Photoreceptor determination network without \( miR-7 \) present and \( < op >= \wedge \) option analysis and simulation results.
- PhD_wo_mirna_AND.net: Photoreceptor determination network without \( miR-7 \) present and \( < op >= \wedge \) option description file.
- PhD_wo_mirna_OR_results.zip: Photoreceptor determination network without \( miR-7 \) present and \( < op >= \vee \) option analysis and simulation results.
- PhD_wo_mirna_OR.net: Photoreceptor determination network without \( miR-7 \) present and \( < op >= \vee \) option description file.
- SOP_w_mirna_AND_results.zip: SOP determination network with \( miR-7 \) present and \( < op >= \wedge \) option analysis and simulation results.
- SOP_w_mirna_AND.net: SOP determination network with \( miR-7 \) present and \( < op >= \wedge \) option description file.
- SOP_w_mirna_OR_results.zip: SOP determination network with \( miR-7 \) present and \( < op >= \vee \) option analysis and simulation results.
- SOP_w_mirna_OR.net: SOP determination network with \( miR-7 \) present and \( < op >= \vee \) option description file.
- SOP_w_mirna.constraints: SOP determination network with \( miR-7 \) present constraint file to be used both for simulation with \( < op >= \wedge \) and \( < op >= \vee \) options.
- SOP_wo_mirna_AND_results.zip: SOP determination network without \( miR-7 \) present and \( < op >= \wedge \) option analysis and simulation results.
- SOP_wo_mirna_AND.net: SOP determination network without \( miR-7 \) present and \( < op >= \wedge \) option description file.
- SOP_wo_mirna_OR_results.zip: SOP determination network without \( miR-7 \) present and \( < op >= \vee \) option analysis and simulation results.
- SOP_wo_mirna_OR.net: SOP determination network without \( miR-7 \) present and \( < op >= \vee \) option description file.
- SOP_wo_mirna.constraints: SOP determination network without \( miR-7 \) present constraint file to be used both for simulation with \( < op >= \wedge \) and \( < op >= \vee \) options.
Table 5: Unique attractors for SOP Determination Network using the operator < op >= ∧ with miR-7. The Table shows 1 point attractor. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

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Table 6: Common attractors for SOP Determination Network using the operator < op >= ∧. The Table shows 5 point attractors and 4 cyclic attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

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Table 7: Unique attractors for SOP Determination Network using the operator < op >= ∨ with miR-7. The Table shows 3 point attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

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Table 8: Unique attractors for SOP Determination Network using the operator < op >= ∨ without miR-7. The Table shows 3 point attractors and 4 cyclic attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.

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Table 9: Common attractors for SOP Determination Network using the operator < op >= ∨. The Table shows 2 point attractors. Bold/Green labels indicate ON nodes, whereas Italic/Red labels indicate OFF nodes.