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Nanotechnology for Angiogenesis: Opportunities and Challenges

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Abstract

Angiogenesis plays a critical role within the human body, from the early stages of life (i.e., embryonic development) to life-threatening diseases (e.g., cancer, heart attack, stroke, wound healing). Many pharmaceutical companies have expended huge efforts on both stimulation and inhibition of angiogenesis. During the last decade, the nanotechnology revolution has made a great impact in medicine, and regulatory approvals are starting to be achieved for nanomedicines to treat a wide range of diseases. Angiogenesis therapies involve the inhibition of angiogenesis in oncology and ophthalmology, and stimulation of angiogenesis in wound healing and tissue engineering. This review aims to summarize nanotechnology-based strategies that have been explored in the broad area of angiogenesis. Lipid-based, carbon-based and polymeric nanoparticles, and a wide range of inorganic and metallic nanoparticles are covered in detail. Theranostic and imaging approaches can be facilitated by nanoparticles. Many preparations have been reported to have a bimodal effect where they stimulate angiogenesis at low dose and inhibit it at higher doses.

Keywords: Nanotechnology; Nanoparticles; Angiogenesis; Vascularization; Cancer therapy; Imaging; Tissue engineering; Regenerative medicine

Abbreviations in alphabetic order:

ABIN-2: A20-binding inhibitor of NF-kappaB 2 AKT: Protein kinase B (PKB) Ang: Angiopoietin BAD: Bcl-2 associated agonist of cell death BAX: Bcl-2-associated X protein BC: Breast cancer Bcl-2: B-cell lymphoma 2 bFGF: Basic fibroblast growth factor BMSCs: Bone marrow mesenchymal stem cells BSA: Bovine serum albumin CAM assay: Chick Chorioallantoic Membrane Assay CA4: Combretastatin A-4 CML: Chronic myeloid leukemia COX: Cyclooxygenase COX-2: Cyclooxygenase-2 CNTs: Carbon nanotubes CPT: 20-(S)-Camptothecin CRC: Colorectal cancer CSF-1: Colony stimulating factor 1 CSFR1: Colony stimulating factor 1 receptor CT: Computerized tomography DAG: Diacylglycerol DDSs: Drug delivery systemes EC: Endothelial cells ECM: Extracellular matrix EGF: Epidermal growth factor EGFR: Epidermal growth factor receptor EMT: Epithelial-mesenchymal transition eNOS: Endothelial nitric oxide synthase EOC: Epithelial ovarian cancer, EPC: Endothelial progenitor cell EPCR: Endothelial cell protein C receptor-dependent ERK: Extracellular-signal-regulated kinase EPR: Enhanced permeability and retention EP1: Prostaglandin E2 receptor 1 ERK: Extracellular signal-regulated kinase FAK: Focal adhesion kinase FDA: Food and drug administration FGF: Fibroblast growth factors FGFR1: fibroblast growth factors 1 FL: Follicular lymphoma FLT-3: Fms-like tyrosine kinase 3

GAC: Gastric adenocarcinoma **GBM:** Glioblastoma GEJAC: Gastroesophageal junction adenocarcinoma GF: Growth factor GIST: Gastrointestinal stromal tumor **GLUTs:** Glucose transporters HBP: Heparanase-binding protein HB-GFs: Heparin-binding growth factors HCC: Hepatocellular carcinoma HER2: Human epidermal growth factor receptor 2 HIF-1 α : Hypoxia-inducible factor 1-alpha hMSCs: Human mesenchymal stem cells HREs: Hypoxia-responsive elements HSA: Human serum albumin Hsp90: Heat shock protein 90 HUVEC: Human umbilical vein endothelial cells H1R: Histamine receptor 1 IPF: Idiopathic pulmonary fibrosis IGFs: Insulin-like growth factors IL-6: Interleukin 6 IL-8: Interleukin 8 iNGR: CRNGRGPDC peptide JNKs: c-Jun N-terminal kinases KATP channel: ATP-sensitive potassium channel LRP: Lipoprotein receptor-related protein MAPK: Mitogen-activated protein kinase MBGs: Mesoporous bioactive glasses MCL: Mantle cell lymphoma MDS: Myelodysplastic syndromes MEK: Mitogen-activated protein kinase kinase MM: Multiple myeloma MMPs: Matrix metallopeptidases MRI: Magnetic resonance imaging (MRI), MTC: Medullary thyroid cancer, mTOR: Mammalian target of rapamycin MWCNTs: Multi-walled carbon nanotubes MZL: Marginal zone lymphoma NETs: Pancreatic neuroendocrine tumors NO: Nitric oxide NPs: Nanoparticles NSCLC: Non-small cell lung cancer NF-Kb: Nuclear factor kappa-light-chain-enhancer of activated B cells **OPN: Osteopontin OVA:** Ovalbumin

PAC: Polyacrylic acid PAMAM: Polyamidoamine PC: Pericyte PCL: Polycaprolactone PDGF: Platelet-derived growth factor PDGFR α : Platelet-derived growth factor receptor α PDGFR β : Platelet-derived growth factor receptor β PDLLA: Poly(D, L-lactic acid) PEG: Polyethylene glycol PEI: Polyethyleneimine PEO: Poly(ethylene oxide) PET: Positron-emission tomography PGE2: Prostaglandin E2 PHD: Proline hydroxylases PIGF: Placenta growth factor PIP2: Phosphatidylinositol biphosphate PKC: Protein Kinase C PLA: Poly(D, L-lactic acid) PLC: Phospholipase C PLGA: Poly(lactic-co-glycolic acid) PPO: Poly(p-phenylene oxide) PTEN: Phosphatase and tensin homolog Ph+ALL: philadelphia chromosome positive acute lymphoblastic leukemia pVHL: Von Hippel-Lindau tumor suppressor protein RCC: Renal cell carcinoma RET: Glial cell-line derived neurotrophic factor receptor RGD: Arginine (Arg)- Glycine (Gly)- Aspartic acid (Asp) **ROS:** Reactive oxygen species rGO: Reduced graphene oxide SEDDSs: Self-emulsifying drug delivery systems SEM: Scanning electron microscope SLNs: Solid-lipid nanoparticles SMCs: Smooth muscle cells SMEDDS: Self-micro-emulsifying agents Sox2: Sex determining region Y-box 2 SPECT: Single-photon emission computed tomography SPIONs: Superparamagnetic iron oxide nanoparticles STS: Soft tissue sarcoma SWCNTs: Single-wall carbon nanotubes SWNH: Single-walled carbon nanohorn TC: Thyroid cancer **TEM:** Transmission Electron Microscope TEOS: Tetraethyl orthosilicate TGF: Transforming growth factor

TIE2: Tyrosine-protein kinase receptor for ANGPT1-2 and 4

TKIs: Tyrosine kinase inhibitors

TNF: Tumor necrosis factor

TNP-470: O-(chloroacetylcarbamoyl)fumagillol

Ub: ubiquitin

VCAM: Vascular cell adhesion molecule 1

VEGF: Vascular endothelial growth factor

VEGFR1: Vascular endothelial growth factor 1

VEGFR2: Vascular endothelial growth factor 2

VEGFR3: Vascular endothelial growth factor 2

VPF: Vascular permeability factor

5-HT: 5-hydroxytryptamine receptors

1. Introduction to Angiogenesis

Any living mammalian tissue needs oxygen and nutrients to ensure cell survival in vivo conditions; therefore, blood vessels play a pivotal role in sustaining life. Endothelial cells (ECs) form the main component of small blood vessels, while pericytes and smooth muscle cells (SMCs) surround larger vessels that are lined with ECs ^{1, 2}. Formation of new blood vessels (neovascularization) within the human body can be achieved via two distinct biological processes. One is called vasculogenesis while the other is called angiogenesis. Vasculogenesis refers to the formation of new vessels de novo from ECs generated by differentiation of progenitor cells (e.g., angioblasts), which self-assemble into lumens and form primitive blood vessels. On the other hand, angiogenesis means the formation of new blood vessels by sprouting from preexisting vasculature ³. A series of molecular and cellular processes are involved in angiogenesis, which can be divided into different steps, including EC activation in response to pro-angiogenic factors, capillary wall degradation via the action of extracellular proteinase enzymes, and formation of a branch point in the vessel walls, ECs migrate into the extracellular matrix (ECM) towards the source of the angiogenic stimulus, and then form tubules with a central lumen that create a vessel network (anastomosis) via the interconnection of the new tubules (Fig. 1).

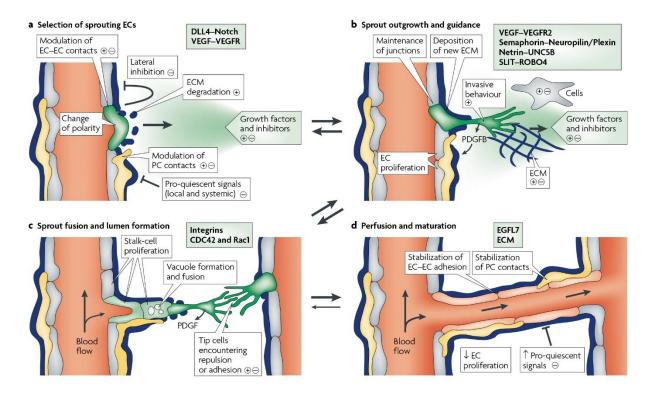


Fig. 1 Schematic representation of different steps of angiogenic sprouting. A) The balance between pro-angiogenic signals (+) (e.g., VEGF), and anti-angiogenic factors (-) (e.g., tight pericyte (PC; yellow) contact), certain ECM molecules and VEGF inhibitors can control the sprouting. Under the appropriate conditions of angiogenesis, ECs can sprout (green), while others inhibit this phenomenon (grey). It has been well documented that the sprouting process needs to flip the apical-basal EC polarity, induce motile and invasive activity, modulate cell-cell contacts and degrade the local ECM. B) Attractive (+) or repulsive (-) cues from cells in the tissue environment are responsible for the growing EC sprouts. C) The fusion of adjacent sprouts into vessels occurs after adhesive or repulsive interactions between the cells at the tip. The fusion of vacuoles facilitates lumen formation in stalk ECs. D) A continuous lumen results from the fusion processes at the EC–EC interface; blood flow enhances oxygen delivery and subsequently reduces the hypoxia-induced pro-angiogenic signals. Maturation processes (e.g., the stabilization of cell junctions, matrix deposition, and tight PC attachment) is likely promoted by increased perfusion. Reproduced with permission from ref ⁴.

Angiogenesis is a critical process involved in embryogenesis and also in maintaining normal homeostasis, including repair and regeneration of injured tissues. Angiogenesis may be deregulated in many pathological conditions. Although angiogenesis remains quiescent during adulthood, it becomes physiologically active in normal conditions such as the cycling ovary and the placenta during pregnancy. Furthermore, angiogenesis regularly occurs via the activation of ECs in response to some specific stimuli (e.g., hypoxia) occurring during the wound healing process to accelerate tissue reconstruction ⁵. However, there is another story about unwanted angiogenesis that occurs in many diseases and disorders, i.e., an imbalance between angiogenic stimulators and inhibitors leads to triggering an angiogenic on-and-off switch. For instance, the angiogenesis process is switched on in the case of malignancies and some inflammatory disorders. On the contrary, insufficient angiogenesis is observed in other pathological conditions such as ischaemic heart tissue, in which healing and regeneration are impaired as a result of dysfunction of ECs, and vessel malformation or regression. To detect and evaluate angiogenesis process, a series of in vitro (e.g., a cell scratch wound), ex vivo (aortic ring assay), and in vivo (chick chorioallantoic membrane (CAM)) assays have been developed and applied that are considered as relible ways towards the translation of results from the laboratory to the clinic ^{6, 7}.

An imbalance in angiogenesis is found in a series of diseases and disorders (e.g., retinopathy); however, this review paper mainly focuses on the importance of inhibiting angiogenesis to fight cancer, and stimulation of angiogenesis in tissue engineering and wound healing by using various types of nanoparticles, nanomaterials, and so on.

2. Angiogenesis mediators

The molecular mediators of angiogenesis consist of different growth factors and cytokines (e.g., VEGF and FGF), matrix metalloproteinases (MMPs), and molecules involved in intracellular signaling pathways (Rho GTPases) (see **Fig. 2** and **Table. 1**) ⁸. There are specific types of receptors on the surface of cells (e.g., ECs) responding to angiogenic biomolecules; receptor tyrosine kinases (RTKs) are among the largest and most well-known receptor families ⁹. VEGF

receptors (VEGFR1-3), FGF receptors (FGFRs), PDGF receptors (PDGFRs), IGF receptors (IGFRs), and the Tie receptors (Tie1 and Tie2) are different classes of RTKs mediating angiogenesis through the activation of relevant signaling pathways after receiving the appropriate signals. For instance, the coupling the IGF to its receptor (IGFRs) triggers two distinct signaling pathways in the cells, resulting improved angiogenesis in the hypoxia condition (see **Fig. 3**).

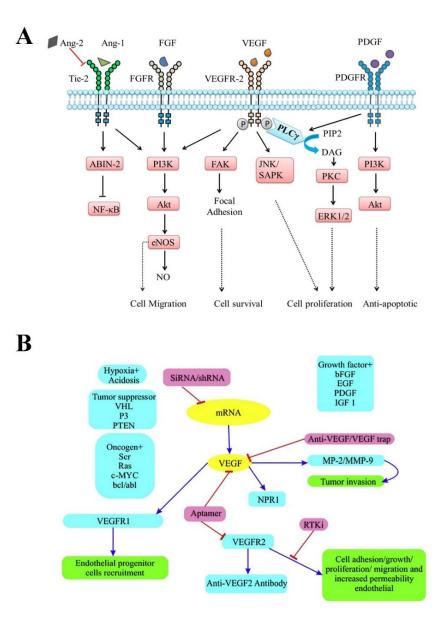


Fig. 2 Schematic illustration of (A) pro-angiogenic mediators and pathways involved in the activation of ECs and (B) the main clinical and preclinical factors involved in anti-angiogenic therapy ^{10, 11}.

Angiogenesis inhibitors can be divided into two distinct classes, including those directly targeting the microvascular ECs, and those indirectly targeting the pro-angiogenic communication pathways between the cancer cells and ECs ¹². A number of direct inhibitors (e.g., angiostatin) have been identified and used to inhibit angiogenesis in cancer treatment. The main action of these inhibitors is to prevent proliferation and migration of ECs stimulated by angiogenesis inducers (e.g., VEGF) ¹³. An inhibitory effect on integrin receptors and subsequent signaling pathways is another mechanism proposed for the action of direct angiogenic inhibitors by which they prevent the proliferation of ECs ¹⁴.

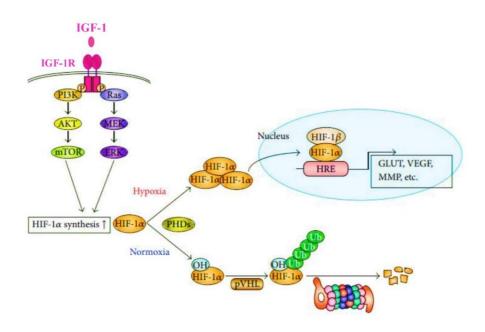


Fig. 3 The binding the IGF, an angiogenic molecule, to IGF-1R receptor on the cell surface activates two cell signaling pathways, leading to increased synthesis of HIF-1 α by which the production of VEGF and thereby improved angiogenesis occur in the hypoxia condition. With some modifications from ref ¹⁵

The U.S. FDA has approved several angiogenesis inhibitors for the treatment of cancer (see section 1.3). R. K. Jain reported that for both direct and indirect anti-angiogenic therapy, the balance between pro-angiogenic and anti-angiogenic factors could be restored through the

reduction of vessel permeability and hypoxia, and enhancement of the homogeneity of blood

flow and perivascular cell coverage ¹⁶.

Category	Molecules	cognate receptor	Effects*	
Growth factors	VEGF	Tyrosine kinase receptors (VEGFR1, VEGFR2, and VEGFR3)		
	PDFG	Tyrosine kinase receptors (PDGFR α and β)		
FGF Tyrosine kinase		Tyrosine kinase receptors (FGFR1, FGFR2, FGFR3, and FGFR4)	PA	
	EGF	Tyrosine kinase receptors: EGFR (ErbB1, HER1), ErbB2 (HER2), ErbB3 (HER3) and ErbB4 (HER4)		
	TGF	Serine/threonine kinase receptors (type I and type II)		
	TNF	Tyrosine kinase receptors (TNFRI and TNFRII)		
	Angiopoetin	Tyrosine kinase receptors (Tie-1 and Tie-2)		
Cytokines	IL-8	CXCR1 and CXCR2 and thereby VEGFR2	PA	
	CSF-1	CSFR1, CSFR 2, and CXCR4	PA	
Bioactive lipids	PGE2	EP1-4 receptors	PA	
Matrix-degrading enzymes	MMPs	Low-density LRP	PA	
	Heparanases	НВР	PA	
Small mediators	NO	Tyrosine kinase receptors (VEGFR1, VEGFR2)	PA	
	Serotonin	5- HT1 and 5-HT2	PA	
	Histamine	H1R and H2R	PA	
Chemotherapeutic agents	Cyclophosphamide	Induces EC apoptosis and decreases circulating EPC		
	Paclitaxel	Microtubule	AA	
VEGF-targeted therapy	Bevacizumab	VEGF-A		
	VEGF-Trap	VEGF-A, VEGF-B, and PIGF	AA	
	Sunitinib	VEGFR1–3, PDGFR- α , PDGFR- β , c-Kit, CSF-1R and Flt-3	AA	
	Sorafenib	VEGFR1–3, PDGFR-β, Raf-1, B-Raf	AA	
	Vatalanib	VEGFR1–3, PDGFR-β and c-Kit	AA	
	Axitinib	VEGFRs, PDGFR- β , and c-Kit	AA	
	SU6668	VEGFR2, FGFR1 and PDGF-β		
FGF-targeted therapy	AZD4547	FGFR1-3	AA	
	Ponatinib	FGFR1-4	AA	
	SSR	FGFRs	AA	
	Brivanib	VEGFRs and FGFRs	AA	
	Dovitinib	FEGFRs, VEGFRs, and PDGFR	AA	

 Table. 1 Pro- and anti-angiogenic factors and receptors. With some modifications from Ref ¹².

	Nintedanib	VEGFRs, FGFRs, and PDGFR	AA
Oncogene-targeted therapy/signaling transduction-targeted therapy	Dasatinib	Src and indirectly VEGF, IL-8	AA
	Tipifarnib	MMP-1	AA
	NVP-AUY922	Hsp90	AA
	Bortezomib	NF-κB-dependent release of VEGF and IL-8	AA
	Gossypol	VEGF and IL-8 release	AA
	Dacinostat	Histone deacetylase	AA
Matrix degrading and remodelling-targeted therapy	DX-2400	MMP-14	AA
	PI-88	Heparanase	AA
	Thrombospondins	CD36 and CD47	AA
Tumor-associated stromal cell-targeted therapy	JNJ-28312141	CSF-1R	AA
	Zoledronic acid	TAM-associated production of VEGF	AA
	Anti-BV8 antibody	Neutrophils recruitment	AA
CAMs-targeted therapy	Cilengitide	$\alpha\nu\beta3$ and $\alpha\nu\beta5$ integrins ligation to matrix proteins	AA
	Volociximab	$\alpha\nu\beta1$ integrin interaction with fibronectin	
	ADH-1	N-cadherin	AA
Inflammatory angiogenesis- targeted therapy	Ibuprofen	COX1/2	AA
	Celecoxib	COX-2	AA
	Repertaxin	CXCR1 and CXCR2	AA

*Note: PA and AA refer to pro-angiogenic and anti-aniogenic effect, respectively.

3. Angiogenesis as a promising target in medicine

Nowadays, controlling unwanted vessel outgrowth is considered as an important therapeutic strategy in the medical setting. Accordingly, a large number of approaches have been developed and approved to suppress aberrant angiogenesis $^{17, 18}$. The molecular mechanisms (signaling pathways, mediators, and receptors) involved in the angiogenesis process, including VEGF/VEGFR, PDGFB/PDGFR- β , and the angiopoietins (Angs) are often considered potential targets $^{19, 20}$. As an example, bevacizumab (Avastin^{*}), a recombinant humanized monoclonal antibody, targets the VEGF/VEGFR signaling pathway to suppress angiogenesis in glioblastoma, and the clinical data have shown improvement in both

progression-free and overall survival of patients ²¹v²². However, it should be noted that development of resistance to anti-angiogenic therapies is common due to activation of alternative pro-angiogenic signaling pathways ²³⁻²⁷. It should be mentioned that cancer cells, which harbor many mutations, often activate compensatory signaling pathways in response to inhibition of a particular pathway, thus rendering cancer cells therapy-resistant. Therefore, there is a rationale for combination therapy (simultaneously targeting multiple pathways) in cancer treatment in order to reduce drug resistance and cancer recurrence ^{28, 29}. In brief, the main antiangiogenic drugs developed to inhibit cancer progression in various types of malignancies include monoclonal antibodies, small-molecule tyrosine kinase inhibitors (TKIs), and non-TKI small-molecule inhibitors (**Table. 2**).

The expression of many pro-angiogenic factors (including VEGF as a key factor) and their cognate receptors is upregulated in the tumor microenvironment. Anti-angiogenic monoclonal antibodies (mAbs) act by blocking the interaction between pro-angiogenic ligands and their cognate receptors hindering the downstream signaling pathways promoting angiogenesis ³⁰. Small-molecule tyrosine kinase inhibitors (TKIs) as anti-angiogenic drugs act by blocking the ATP binding site in a pro-angiogenic receptor and, hence, inhibiting phosphorylation of the tyrosine residue of that receptor, which eventually hinders downstream pro-angiogenic signaling pathways. Compared to anti-angiogenic mAbs, TKI usually targets not only the VEGF/VEGFR pathway but also other pro-angiogenic pathways such as platelet-derived growth factor receptor (PDGFR), mesenchymal epithelial transition factor receptor (c-MET) and TIE-2 ³¹.

Table. 2 FDA-approved anti-angiogenic drugs used to treat different cancers.

Classification	Drug name	Chemical formulation	Mechanism of action	Clinical usage	Re f (s)
Monoclonal Antibodies	Bevacizumab (Avastin®, Genentech)	C6538H10034N1716O2033S44, Mw = 149 kDa	- Hinders the interaction between VEGF-A and VEGFR2 via targeting VEGF-A	A variety of cancers including CRC, BC, GBM, NSCLC, RCC, and EOC	32, 33
	Ramucirumab (Cyramza®, ImClone Systems Incorporated)	C6374H9864N1692O1996S46, Mw = 143.6 kDa	- Hinders the interaction between VEGFR2 ligands (i.e., VEGF-A, VEGF-C, and VEGF-D) and VEGFR2 via targeting VEGFR2	GAC, GEJAC and NSCLC	34, 35
	Aflibercept (Zaltrap®, Regeneron pharmaceuticals)	C4318H6788N1164O1304S32, Mw = 115 kDa	- Suppresses angiogenesis via hindering the interaction between VEGF isoforms, mainly VEGF-A, VEGF-B, and PIGF, and their cognate receptors VEGFR-1 and VEGFR- 2	Metastatic CRC and age-related macular degeneration	36, 37
	Olaratumab (Lartruvo®, Eli Lilly)	C6554H10076N1736O2048S40, Mw = 154 kDa	- Acts against the external domain of human PDGFR- α , blocking its ligand binding hindering activation of downstream signaling molecules protein kinase B (Akt) and MAPK	STS	38, 39
Small- molecule tyrosine kinase inhibitors	Axitinib (Inlyta®, Pfizer)	C22H18N4OS, Mw = 386.47 Da	- Inhibits the VEGFR-1, VEGFR-2, VEGFR-3	RCC	40
	Cabozantinib (Cabometyx®, Exelixis)	C28H24FN3O5, Mw = 501.514 Da	- A multi-kinase TKI of various receptors including VEGFR-1, -2 and -3, KIT, FLT-3, AXL, RET, MET, and TIE- 2	RCC and MTC	41
	Lenvatinib (Lenvima®, Eisai)	C21H19ClN4O4, Mw = 426.86 Da	- A multi-kinase inhibitor of VEGFR 1, -2 and -3, fibroblast growth factor receptor FGFR 1, -2 and -3, PDGFRα, KIT, and RET	Radioiodine refractory differentiated TC, advanced RCC, and HCC	42
	Nintedanib (Ofev®, Boehringer Ingelheim Pharmaceuticals)	C31H33N5O4, Mw = 539.6248 Da	- A multi-kinase inhibitor of VEGFR 1, -2 and -3, FGFR 1, -2 and -3, PDGFR α/β , and FLT3	IPF	43
	Pazopanib (Votrient®, GlaxoSmithKline)	C21H23N7O2S, Mw = 437.518 Da	- A multi-kinase inhibitor of several kinases including VEGFR 1, -2 and -3, FGFR 1, -2 and -3, PDGFR α/β , and KIT	Advanced RCC and advanced soft tissue sarcoma	44
	Ponatinib (Iclusig®, Ariad Pharmaceuticals)	C29H27F3N6O, Mw = 532.5595 Da	- A multi-kinase inhibitor of several kinases mainly BCR-ABL, BCR-ABL T315I, VEGFR2, PDGFR α , FGFR1, -2 and - 3, ephrin receptor EPHR, SRC family kinases, KIT, RET, TIE2, and FLT3	CML or Ph+ALL resistant to previous TKI therapies. The drug is also effective for CML or Ph+ ALL patients with positive T315I mutation	45, 46

	Regorafenib (Stivarga®, Bayer)	C21H15ClF4N4O3, Mw = 482.815 Da	- A multi-kinase inhibitor of several kinases including VEGFR 1, -2 and -3, FGFR 1, -2, PDGFRα/β, RET, KIT, TIE2, Eph2A, BCR-ABL, B-RAF, and B-RAF V600E	Metastatic CRC, locally advanced, unresectable or metastatic GIST previously treated with imatinib or sunitinib, and HCC	47
	Sorafenib (Nexavar®, Bayer)	C21H16ClF3N4O3, Mw = 464.825 Da	 A multi-kinase inhibitor of several kinases including BRAF, BRAF V600E, KIT, FLT-3, VEGFR-2, -3, and PDGFR-ß Targets the Raf/Mek/Erk pathway inhibiting downstream signaling pathways leading to cancer hallmarks such as cell proliferation, apoptosis evasion, angiogenesis, invasion, and metastasis 	Unresectable HCC, advanced RCC and differentiated TC refractory to radioactive iodine	48
	Sunitinib (Sutent®, Pfizer)	C22H27FN4O2, Mw = 398.4738 Da	A multi-kinase inhibitor of several kinases including VEGFR 1, -2 and -3, PDGFRα/β, KIT, FLT3, colony stimulating factor receptor Type 1(CSF-1R), and RET	Advanced RCC, GIST resistant to imatinib and NETs	49
	Vandetanib (Caprelsa®, Genzyme Corporation)	C22H24BrFN4O2, Mw = 475.354 Da	- Multi-kinase inhibitor of several kinases mainly EGFR, VEGFR2, and RET	Locally advanced or metastatic MTC	50
Non-TKI small- molecule inhibitors	Thalidomide (Thalidomide®, Celgene)	C13H10N2O4, Mw = 258.2295 Da	- Inhibition of the production of TNF- α and VEGF	Multiple myeloma (MM)	51
	Lenalidomide (Revlimid®, Celgene)	C13H13N3O3, Mw = 259.2606	- Inhibiting the expression of COX-2	MM, MDS and MCL, FL, and MZL	52
	Temsirolimus (Torisel®, Wyeth Pharmaceuticals)	C56H87NO16, Mw = 1030.2871 Da	 An inhibitor of mTOR Inhibition of mTOR suppresses angiogenesis by reducing levels of the hypoxia-inducible factors HIF-1 and HIF-2, and the VEGF 	RCC	53
	Everolimus (Afinitor®, Novartis)	C53H83NO14, Mw = 958.24 Da	An inhibitor of mTOR. Inhibition of mTOR suppresses angiogenesis by reducing levels of the HIF-1 and HIF-2, and the VEGF	Some malignancies mainly RCC, advanced HR+ (hormone receptor), HER2- BC; progressive neuroendocrine tumors of pancreatic, gastrointestinal or lung origin	54

4. The pivotal role of chemistry towards the angiogenic design of nanomaterials

Nowadays, the design and development of nanotechnology-based therapies by using organic and inorganic materials form a substantial part of the modern medicine; indeed, chemistry plays a central role in this sense ⁵⁵. There are huge numbers of commercially available nanotechnology-based products (e.g., nanopharmaceuticals) on the market, which are used in a broad range of applications including cancer therapy ⁵⁶. Still, more research is needed to progress towards novel and more efficient nanomaterials/nano-systems-based cancer therapies, which will be key to overcome the limitations of current treatments (e.g., drug resistance). Therefore, it is an undeniable evidence that the medicinal chemistry will play a critical role in imaginable achievments in the near future ⁵⁷.

The pro- and anti-angiogenic potential of nanomaterials could be straightforwardly controlled by chimestry rules, from simple adjustments in the synthesis and structural manipulation to complicated surface modifications, self-assembly, processing and integration to make smart materials in the concept of advanced healthcare materials. As an illustration, making mesoporous bioactive glass (MBG) nanoparticles with the ability to carry biomolecules (e.g., pro- or anti-angiogenic agents) is simply applicable via a wet-chemical technique, i.e. the sol-gel process ⁵⁸⁻⁶⁰. Targeted cancer therapy is of utmost importance to reduce side effects of chemotherapy as well as to improve the clinical outcomes. Targeting angiogenesis via nano-structured materials is one of the most interesting issues in cancer therapy ^{61, 62}. Chemical coupling of various biomolecules including antibodies, peptides (e.g. RGD), and peptidomimetics to nanomaterials has provided the opportunity to target vascular integrins (e.g., $\alpha_v\beta_3$ integrin) and subsequent targeted cancer therapy ⁶³⁻⁶⁶.

It is also worth pointing out that some nano(materials) are able to elicit an inherent pro- or antiangiogenic effect associated to the release of therapeutic ions. In this regard, materials composition and chemistry strongly govern the biological response. However, there are some critical factors that limit the progress in the field of angiogenesis modulation utilizing ions. Essentially, this is because (i) ions can easily diffuse to other non-target cells or tissues and stimulate unwanted responses, and (ii) the biochemical/biomolecular effects elicited by such ions may be partly unpredictable. Hence, at least a couple of key questions in "ionic research" need to be addressed before clinical translation, namely: How can the non-specific side effects of ion-based therapeutics be minimized? What is the signaling cascade of these ions on angiogenesis? We cannot ignore that our current biochemical/biomolecular knowledge is still incomplete and unable to provide an exhaustive response to these questions; the goal of this review is to draw a structured picture of the relevant state-of-the-art, on which researchers can further build new knowledge and plan experiments to bridge the gaps.

5. Nanotechnology meets angiogenesis

Loading and delivery of various natural and synthetic pro-angiogenic or anti-angiogenic substances by using nanostructured vehicles is recognized as one of the most promising approaches in medicine ⁶⁷⁻⁶⁹. There is strong evidence for the utility of nano-sized delivery systems for therapeutic drugs, since they can overcome the limited tissue diffusion of drugs, protect them in the blood circulation, and lower the risk of systemic toxicity. In other words, targeted therapy using nano-scale vehicles helps drug-loaded nanostructures more easily reach the desired sites in the body (cells, tissues, and organs) and the drug release profile occurs in a

more controlled manner ⁷⁰. In addition, it has been well-documented that organic and inorganic nanoparticles can display pro-angiogenic and anti-angiogenic characteristics depending on their nano-sized design (see **Fig. 4**) ^{71, 72}. In the following sections we introduce and discuss the types of nano-sized particles (organic and inorganic), as well as nanotechnology-based systems that have been designed and developed for pro- and anti-angiogenic applications.

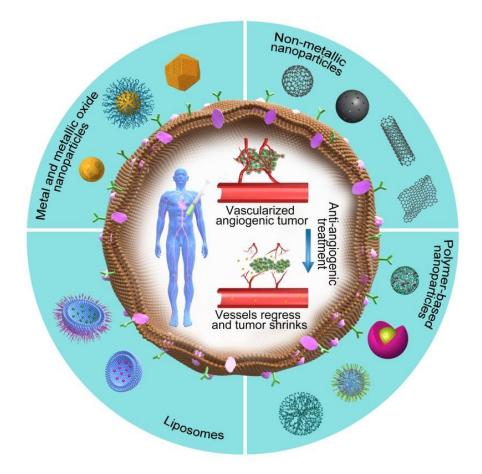


Fig. 4 Different types of NPs that have been used as therapeutics for anti-angiogenesis and vessel regression. From ref ⁷².

6. Polymeric nanoparticles as carriers for the delivery of anti-angiogenic biomolecules

The most commonly-reported natural and synthetic materials for constructing nanoparticle carriers are polymers, liposomes, micelles, and inorganic nanoparticles ⁷³. The use of polymers in

drug delivery strategies (DDSs) has been proved to be a successful approach; some of them have been on the market since the early 1990s ⁷⁴. However, polymeric nanoparticles used as drug delivery vehicles are considered to be newer members of DDSs, bringing new hope in medicine thanks to their properties, such as higher bioavailability, low toxicity, and controllable drug release kinetics ^{75, 76}. Although polymeric nanoparticles have been used in the treatment of some diseases (e.g., arthritis, multiple sclerosis), the main focus is still on cancer therapy ⁷⁷. In this sense, numerous nano-sized polymers have been used to load and deliver anti-angiogenic chemicals and drugs, in the fight against various cancers (see **Table. 3**) ⁷². Among them, PEG, PLA, PCL, PLGA, chitosan, heparin, gelatin, and albumin have extensively being using for therapeutic angiogenesis, either in bare or modified form.

PEG is a non-ionic water-soluble polymer, which has been extensively used in drug delivery applications due to its biocompatibility. More than 35 FDA-approved nanoparticles incorporating PEG are presently on the market, designed for both imaging and therapeutic purposes ⁷⁸. There are some experimental studies in which PEG has been used in combination with other biocompatible polymers for targeted delivery of angiogenic substances ^{79, 80}.

PLA is an FDA and EMA-approved material thanks to its excellent properties such as biocompatibility, biodegradability and lack of any toxic by-products. Several FDA-approved DDSs based on PLA or PGA/PLA copolymers are available on the market, used in nanoparticle or microparticle formulations for the treatment of different cancers ⁸¹. The use of nano-sized PLA particles has achieved much attention in drug delivery applications ⁸²; one of the first reports on the use of this nano-sized particles in an anti-angiogenic strategy was published by Burt et al. in 1995 ⁸³. The use of nanoparticles containing co-polymers made of PCL and other biocompatible polymers (e.g., PEG) is suggested to improve anti-angiogenic efficacy and thereby anti-cancer potential *in vivo* compared to PLA ^{82, 84}. In contrast, it has been reported that electrospun PLA nanofibers could increase the proliferation of ECs *in vitro* ⁸⁵. Moreover, surface functionalization of PLA has been proposed as an approach to increase its pro-angiogenic properties; polyethylenelmine (PEI) and polyacrylic acid (PAC)-coated electrospun PLA nanofibers significantly promoted angiogenesis both *in vitro* and *in vivo* ⁸⁶. PLA may be a suitable platform for delivery of a range of pro-angiogenic molecules, such as VEGF ⁸⁷.

As an FDA-approved substance, PCL in different formulations has received much attention in controlled drug delivery and tissue engineering applications ^{88, 89}. For example, Niza et al. prepared micro- and nano-sized vehicles based on PCL for doxorubicin delivery to glioblastoma ⁹⁰. However, some limitations have restricted the use of PCL in biomedicine, as compared to PLGA, such as its slow biodegradability ⁹¹. There are few studies in the literature in which PCL was used for pro- and anti-angiogenic applications ^{92, 93}; Jiang et al. could successfully prepare PCL nanofibers containing VEGF-encapsulated gelatin particles to enhance MSCs differentiation and angiogenesis of ECs ⁹³.

PLGA is another FDA-approved pharmaceutical product, extensively used as a DDS in imaging, diagnostics, and therapy due to its favorable properties, such as biocompatibility, as well as controlled and sustained release of drugs ⁹⁴. In several studies, researchers have demonstrated the applicability of PLGA nanoparticles (pristine, chemically modified, or hybrids) to load and deliver anti-angiogenic molecules ^{84, 95-97}. A tumor-vessel-recognizing and tumor-penetrating system was developed based on iNGR-modified PEG-PLGA nanoparticles for treating glioma in mice ⁹⁸. The modified nanoparticles could penetrate into the tumor parenchyma and

showed good cellular uptake in HUVECs, resulting in enhanced anti-proliferative and anticapillary tube formation activities of paclitaxel *in vitro*. Moreover, the results showed improved anti-angiogenic acticity of the drug-loaded nano-carriers. It is worth noting that several research groups have used PLGA nanoparticles to load pro-angiogenic biomolecules (e.g., aptamers) and other chemicals to improve angiogenesis and subsequently accelerate tissue healing ⁹⁹⁻¹⁰¹.

As an FDA-approved product, chitosan in micro- and nano-sized formulations is commonly used in a broad range of biomedical applications, from wound healing to drug delivery ^{102, 103}. The biological activities of chitosan can be summarized as antimicrobial, antioxidant, and anti-cancer ¹⁰⁴⁻¹⁰⁶. Furthermore, it has been reported that chitosan nanoparticles can inhibit angiogenesis in a dose- and time-dependent manner in cancer models *in vivo* ¹⁰⁷. The suppression of VEGFR-2 and subsequent blockage of VEGF is proposed to explain the antiangiogenic activity of chitosan nanoparticles. Anti-angiogenic activity was also observed in the case of depolymerized chitosan products, i.e., water-soluble low-molecular-weight chitosan (LMWC) and chito-oligosaccharides (COs) ^{108, 109}. Furthermore, it should be stated that chitosan has also been used as a drug delivery system in pro- and anti-aniogenic applications ¹¹⁰⁻¹¹².

Heparin is a natural water-soluble polysaccharide with a high negative surface charge used for a broad range of applications, from the treatment of thromboembolism to anti-cancer strategies. Although heparin exhibits anticancer effects by inhibition of angiogenesis ¹¹³, the side effects of thrombocytopenia and heart arrhythmias restrict its long-term administration in humans ¹¹⁴⁻¹¹⁶. Chemical modification using deoxycholate or lithocholate could reduce the anticoagulant activity of heparin, encouraging its broader use as an anti-tumor drug carrier. The

use of nano-sized heparin as a conjugate carrier for delivery of a wide range of pro-angiogenic and anti-angiogenic substances has also been proposed ¹¹⁷⁻¹²⁶.

Gelatin is extensively used in biomedical products due to its versatile characteristics, including biocompatibility, biodegradability, non-antigenicity, cost-effectiveness, and easy availability ¹²⁷. A number of experimental studies showed the utility of cationic gelatin in drug delivery strategies, either as pristine or surface-modified forms ¹²⁸. However, the use of nano-sized gelatin in the development of DDSs has been encouraged by several surface modifications to improve the targeted and sustained release of therapeutic genes, drugs, and chemicals ¹²⁹⁻¹³³. The study published by Kommareddy and Amiji is one of the first reports using gelatin nanoparticles as an antiangiogenic strategy ¹³⁴. They used gelatin, thiolated gelatin (SHGel), and PEG-modified gelatin (PEG-Gel) nanoparticles to encapsulate and deliver plasmid DNA encoding the VEGF receptor-1 (VEGFR1 or sFlt-1) in order to entrap excess VEGF produced by tumor cells and thereby reduce the angiogenesis process. On the other hand, gelatin nanoparticles have been used in pro-angiogenic strategies; such as, the sustained release of pro-angiogenic factors (e.g., VEGF and bFGF) loaded into gelatin-based nanoparticles to improve neo-vascularization ¹³⁵, ¹³⁶.

Albumin is one of the most important components of human blood with a half-life of 19 days on average, which is extensively used in various biomedical applications such as drug delivery ¹³⁷. In order to improve the inherent properties of albumin, nano-sized albumin systems can be prepared via different procedures including desolvation (coacervation), emulsification, thermal gelation, nano spray drying, and self-assembly ¹³⁸⁻¹⁴⁴. The FDA approved a nanoparticle formulation (130-nm) of albumin-bound paclitaxel called ABI-007 (Abraxane[®]; Abraxis BioScience

and AstraZeneca), which is used to treat cancers such as breast, non-small-cell lung carcinoma (NSCLC), and pancreatic cancer ^{145, 146}. Albumin nanoparticles (either in pristine or modified forms) have been studied as anti-angiogenic strategies for treating various types of solid tumors in experimental models ¹⁴⁷⁻¹⁵⁰. In addition, albumin could be applied as a suitable platform to deliver anti-angiogenic cargos to tumoric sites ¹⁵¹.

7. Regulation of angiogenesis by chemicals and drugs

7.1 Herbs and Phytochemicals

The use of plant-derived chemicals and drugs for pro-angiogenic and anti-angiogenic strategies has a long history, especially in traditional Chinese medicine. With the emergence of modern technology, chemical optimization of these compounds has led to a substantial improvement in their effectiveness to modulate angiogenesis ^{152, 153}. These natural products affect the angiogenesis process via distinct molecular pathways (Fig. 5) ¹⁰. In the following sections, we introduce and discuss the pro- and anti-angiogenic activities of the most commonly-used plant-derived components, and then show their effectiveness when used in nano-sized format, including nano-carriers.

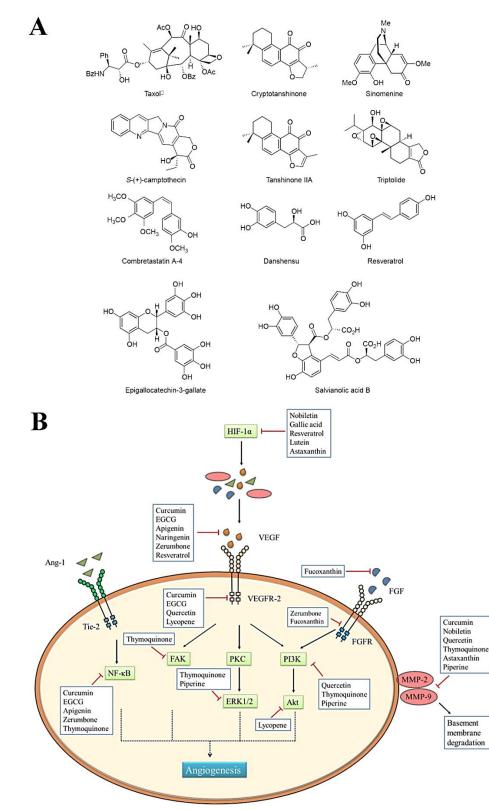


Fig. 5 (A) Chemical structures of some of the most well-known pro- and anti-angiogenic substances derived from medicinal plants, and (B) the main signaling pathways of angiogenesis. Reproduced with permission from ref $^{10, 152}$.

7.2 Curcumin

Curcumin is the principal polyphenolic compound present in turmeric, and is among the most studied organic compounds in biomedical applications. There are conflicting data regarding the potential of curcumin in the new vessel formation; however, most reports seem to suggest the anti-angiogenic activity of this biomolecule. The anti-angiogenic properties of curcumin result from its interaction with multiple cell signaling proteins and pathways ¹⁵⁴. As an illustration, curcumin shows inhibitory effects on the expression or synthesis of some of the most important proteins involved in angiogenesis in solid tumors, including HIF-1 α , VEGF, CD31, and bFGF ^{155, 156}. The inhibitory effects of curcumin on angiogenesis is also related to its activity against cell signal transduction pathways involving PKC and the transcription factors NF-κB and AP-1. Furthermore, curcumin could affect proteinases (MMP and uPA families), which are involved in the angiogenesis process. Some studies showed that curcumin can act as a blocker of cell adhesion molecules that are upregulated in active angiogenesis ¹⁵⁷. The use of nanoformulated curcumin shows promise for overcoming some limitations of curcumin such as its low aqueous solubility, rapid systemic clearance, and low cellular uptake. Although the preparation of curcumin nanoparticles has been previously reported by a process based on a wet-milling technique ¹⁵⁸, most research has been focused on using different nanocarriers (e.g., liposome/lipid nanoparticles, micelles, polymer conjugates, etc.) to efficiently encapsulate and then deliver curcumin to target sites ¹⁵⁹. In the cancer therapy setting, various nano-formulations of curcumin, including micelles and liposomes, have exhibited a significant improvement in antiangiogenic efficacy ¹⁶⁰⁻¹⁶³; Mukerajee et al. introduced targeted nanocurcumin therapy as an effective approach in inhibiting neovascularization ¹⁶⁴. The anti-angiogenic and subsequent anticancer effects of liposomal curcumin have also been evaluated *in vitro* and *in vivo* against human pancreatic cancer ¹⁶⁵. Intraperitoneal injection of 20 mg/kg liposome-encapsulated curcumin into tumor-bearing mice (three times a week for one month) could reduce tumor growth up to 42% in comparison to untreated animals. The histological and immunohistological assessment showed a significant decrease in the formation of blood vessels, as well as expression of VEGF in animals treated with liposomal curcumin.

7.3 Icariin

Icariin is a prenylated flavonol glycoside and one of the main bioactive components of *Epimedium* (family Berberidaceae), which is used in a broad range of medical applications, including cancer therapy ¹⁶⁶. Icaritin is another bioactive chemical found in the ethyl acetate fraction of Epimedii extract. There are several studies in the literature revealing anti-cancer activities mediated by these organic compounds, including apoptosis, cell cycle arrest, antiangiogenesis and anti-metastasis, as well as immunomodulation ¹⁶⁶. It has been proposed that their anti-angiogenic effects could be mediated via inhibition of the ERK signaling pathway ¹⁶⁷. Icariin and icaritin have inhibitory effects on the proliferation, migration, and tube formation of human umbilical vein endothelial cells ¹⁶⁸ and could attenuate angiogenesis in a chick embryo model in a dose-dependent manner ^{169, 170}. In vivo experiments have also shown that both icariin and icaritin exhibit anti-angiogeneic effects in xenograft models of tumors, including hepatocellular and renal carcinoma ^{171, 172}. The inhibition of the VEGF signaling pathway via reduction of the transcriptional activity of HIF-1 α was reported to explain the anti-angiogenic effects of icariin and icaritin in vitro and in vivo ¹⁷³. On the contrary, there are a few studies claiming that icariin can stimulate angiogenesis by activating relevant signaling pathways ^{174, 175}.

For example, Chung et al. reported icariin at a concentration of 5 μ M could activate the MEK/ERK and PI3K/Akt/eNOS-dependent signaling pathways in human endothelial cells; however, it did not affect VEGF signaling pathway. The authors showed that this pro-angiogenic concentration (5 μ M) was comparable to that of 10 ng/ml VEGF. The results of an *ex vivo* experiments on rat aortic rings showed that 5 μ M icariin increased vessel sprouting at the cut edge, three times more than controls. Other research groups also reported that icariin (at concentrations of 7.5, 15 and 30 μ M) via activating eNOS increased the number of sprouting tubules in endothelial progenitor cells (EPCs) ¹⁷⁶. Although icariin has been used in both pro- and anti-angiogenic strategies, there are few experimental studies concerning the use of its nanoformulation ¹⁷⁷⁻¹⁷⁹.

7.4 Resveratrol

Resveratrol (3,5,4'-trihydroxystilbene) is a non-flavonoid polyphenolic compound found in a number of plants, including grapes, peanut roots, and the heartwood of mulberry trees ¹⁸⁰, ¹⁸¹. This compound has been shown to be a cancer chemopreventive agent, as it could inhibit angiogenesis in various tumors ¹⁸²⁻¹⁸⁴. It has been previously well-documented that systemic delivery of resveratrol at concentrations of 2.5–100 mg/kg inhibits tumor-induced neovascularization in animal models ¹⁸⁵. However, *in vitro* studies showed that the antiangiogenic activity of resveratrol was dose-dependent, so that it could completely inhibit tube formation and cell migration of HUVECs at concentrations of 50, 100 and 500 μM, while it showed pro-angiogenic activity at lower concentrations (e.g., 5 μM) ¹⁸⁶. The molecular mechanisms involved in the pro- and anti-angiogenic activities of resveratrol have explored in several experimental studies. These include altering endothelial morphology and subsequently causing cytoskeletal rearrangements in both β-catenin and VE-cadherin; activating PI3-K/Akt and

MAPK/ERK signaling pathways followed by upregulation of endothelial NOS and increased levels of NO, leading to over-expression of VEGF and MMPs ¹⁸⁷. On the other hand, resveratrol at high doses can bind to VEGF thus interfering with its binding to VEGF receptors, resulting in a decrease in VEGF receptor-2 phosphorylation and JNK phosphorylation as well as inhibiting the VEGF-mediated phosphorylations of eNOS, Akt and Erk ^{188, 189}. Like other anti-angiogenic substances, targeted delivery of resveratrol to tumor sites could be conducted using a variety of nano-based DDSs, including solid lipid nanoparticles (**Fig. 6**) ^{190, 191}. For example, Pund et al. successfully used a lipid-based nanoemulsion delivery system of resveratrol, and showed its good anti-angiogenic activity *in vivo* using a CAM assay ¹⁹². The nanoemulsification included Acrysol K 150 as a lipid and a mixture of Labrasol and Transcutol HP as a surfactant system to form emulsion particles with a size of 85 nm to 120 nm. A few studies have explored resveratrol nanoparticles; Kim et al. reported the successful preparation of trans-resveratrol (t-RVT) in nanoparticles via temperature-controlled anti-solvent precipitation with hydroxypropyl methylcellulose as the stabilizer ¹⁹³.

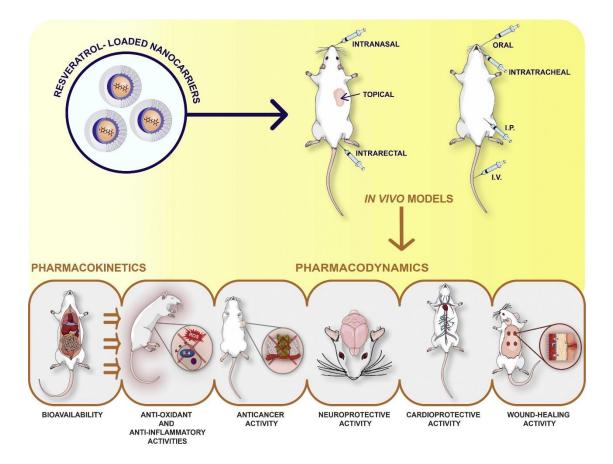


Fig. 6 Pharmacokinetics and pharmacodynamics of resveratrol including bioavailability, anti-oxidant and inflammatory, anticancer, as well as healing properties, are enhanced when administered by nanocarriers *in vivo*. Reproduced with permission from ref ¹⁹⁰.

7.5 Paclitaxel

Paclitaxel (Taxol^{*}) is a naturally occurring diterpene alkaloid, which was firstly isolated from the bark of Pacific Yew, *Taxus brevifolia* Nutt. (Taxaceae) in the 1960s, and since then has been commonly used clinically as first-line chemotherapy for many different cancers (e.g., lung and breast) ¹⁹⁴. As a member of the taxane family, paclitaxel binds to the beta-subunit of polymerized tubulin in the cytoskelton and prevents the dissociation of the tubulin subunits from the tubule, leading to the formation of microtubule bundles, and subsequent cell cycle arrest

inhibiting mitosis ^{195, 196}. The first reports on the anti-angiogenic activity of paclitaxel were published by Dordunoo et al. in 1995¹⁹⁷ and Belotti et al. in 1996¹⁹⁸. Paclitaxel can inhibit angiogenesis at a broad dose range, from ultra-low to high concentrations. For instance, Wang et al. reported that paclitaxel inhibited the proliferation of human ECs at ultra-low concentrations of 0.1–100 pM, with an IC₅₀ (the half maximal inhibitory concentration) of 0.1 pM ¹⁹⁹. The antiangiogenic activity of paclitaxel at low concentrations was also observed using in vivo models of neovascularization (CAM model), in which paclitaxel inhibited angiogenesis at doses of 4, 8, and 12 nM ²⁰⁰. Several studies (in vitro, ex vivo, and in vivo) showed that paclitaxel hinders proliferation, motility, and migration of ECs by interfering with a series of molecular cellular signaling pathways involved in angiogenesis (Fig. 7) ²⁰¹. Two of the most important and welldefined target proteins of paclitaxel are VEGF and FGF-2 in HUVECs, as reported by several studies ^{202, 203}. Moreover, paclitaxel can down-regulate the expression of Ang-1, a potent provasculogenic and angiogenic factor, in vitro ²⁰⁴. The induced expression of TSP-1, a potent endogenous inhibitor of angiogenesis, is another route by which paclitaxel could elicit its antiangiogenic activity ²⁰⁵.

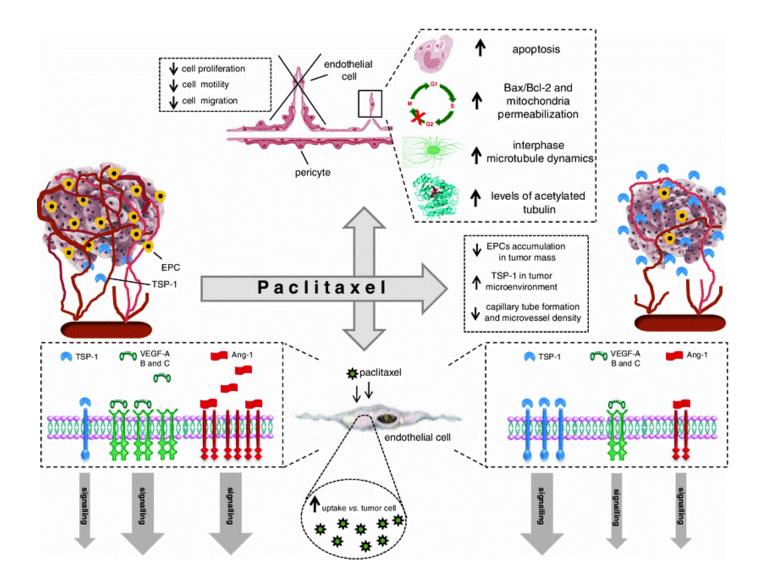


Fig. 7 Different molecular and cellular mechanisms of the antiangiogenic activity of paclitaxel. Reproduced with permission from ref ²⁰¹.

Nano-based systems designed for paclitaxel delivery have shown an enhanced transvascular permeability and increased accumulation in tumors causing increased cancer cell death ²⁰⁵. Moreover, the use of nano-based DDSs could also be effective in the treatment of multidrug-resistant cancers ²⁰⁶. Up to date, several carrier systems have been developed and tested, including liposomes, solid lipid nanoparticles, lipid nanocapsules, and nano-emulsions ²⁰⁷.

Banerjee et al. prepared Tyr-3-octreotide (TOC)-modified solid lipid nanoparticles (SLN) containing paclitaxel to improve anti-cancer efficacy via the inhibition of angiogenesis in glioblastoma-bearing rats ²⁰⁸. The anti-angiogenic potential of this system was confirmed via analysis of tube formation and CD31 staining, and its anti-glioma efficacy was proven by histopathological assessment of the treated animals. Furthermore, the use of pure paclitaxel nanoparticles for treating cancer has also been reported. As one illustration, Wu et al. prepared pure paclitaxel nanoparticles using an electrostatic spraying method and showed their anti-cancer effect on human liver cancer SMMC-7721 cells ²⁰⁹.

7.6 Camptothecin

20-(*S*)-Camptothecin (CPT) is a natural pentacyclic alkaloid first isolated by Wall et al. from the bark of the Chinese tree *Camptotheca acuminata* ²¹⁰. This compound is a topoisomerase-I (Top1) inhibitor with the ability to inhibit DNA replication, thus subsequently killing tumor cells as well as inhibiting EC proliferation ²¹¹. Over the years, medicinal chemists have succeeded in synthesizing several CPT derivatives, including topotecan (TPT, 3), irinotecan (CPT-11, 4), and belotecan (CKD-602, 5) which have received FDA-approval for various cancers such as ovarian and small-cell lung cancer ²¹². Furthermore, a series of water-soluble and nonwater-soluble analogs are being tested in preclinical and clinical trials ²¹³⁻²¹⁷. One of the first reports on the anti-angiogenic activity of CPT was published by Clements et al. ²¹⁸. They aimed to determine the inhibitory effects of sub-cytotoxic doses of CPT and TPT on angiogenesis both *in vitro* and *in vivo*, in comparison to other anti-angiogenic compounds (i.e., TNP-470 and cisplatin). Their results showed that treatment with 50 nM CPT or TPT led to growth inhibition in HUVECs without any cytotoxicity. Furthermore, CPT or TPT effectively inhibited angiogenes in an in vivo disc model comparable to TNP-470. Similar results have been reported in other experimental studies, clarifying the anti-angiogenic potential of CPT at various doses and formulations against different cancers ²¹⁹⁻²²¹. The use of nanotechnology for targeted delivery of CPT is a promising approach to overcome its limitations (e.g., low bioavailability and poor water solubility); therefore, various CPT-based nanodrug platforms (e.g., liposomes and nanosponges) have been tested in cancer therapy. It is worth mentioning that, although nano-structured delivery systems developed for CPT have been extensively studied, their widespread use is limited due to the side effects of the nanomaterials used. Therefore, the application of CPT nanodrugs prepared by selfassembled drug molecules is preferred to delivery systems based on nanocarriers ²²². From an anti-angiogenic point of view, targeted delivery of CPT can be achieved by nano-structured platforms; Gigliotti et al. used CPT-containing nanosponges to enhance the cytotoxic effect against anaplastic thyroid cancer cells in vitro, and suppress angiogenesis in orthotopic xenograft tumors in vivo ²²³. CRLX101 is a nanoparticle preparation containing a cyclodextrin-based polymer and camptothecin, and is in phase II clinical trials for treating metastatic castrationresistant prostate cancer and small cell lung cancer. Preclinical studies have revealed that this nanoformulation could improve cancer (e.g., gastric and breast) chemoradiotherapy via inhibiting DNA repair (apoptosis) and HIF1 α (anti-angiogenesis) ²²⁴⁻²²⁷.

7.7 Combretastatin

Combretastatin A-4 (CA4) is a dihydrostilbenoid used as a chemotherapy drug for the treatment of a variety of solid tumors, such as ovarian, and colon cancer ^{228, 229}. This compound

is extracted from the bark of the South African bush willow tree, i.e., *Combretum caffrum*²³⁰. CA4 exerts its anti-cancer activity via inhibiting polymerization of tubulin via attachment to the colchicine-binding site of the B-tubulin subunit in mammalian cells ^{231, 232} ^{231, 232}. It has been shown that CA4 exhibits cytotoxicity (doses below 4 nM) against bladder cancer cells through inducing G2-M phase arrest with sub-G1 formation ²³³. CA4 can induce apoptosis in cancer cells by activating caspase-3 and decreasing BubR1/Bub3²³³. CA4 could cause the disruption of tubular organization inside HUVECs followed by inhibition of the branching outgrowth ²³⁴. Therefore, the CA4 acts as a vascular disrupting agent, which is considered to be a new class of anti-angiogenic drugs. Recent studies have demonstrated that the anti-angiogenic activity of CA4 could suppress microvessel formation at a dose of 5 nM and completely block microvessel sprouting at a dose of 20 nM in the aortic ring model embedded in Matrigel ²³⁵. The attenuation of the VEGF/VEGFR-2 signaling pathway is considered to explain the anti-angiogenic activity of CA4 ²³⁵. Ren et al. had previously proposed the Raf-MEK-ERK and Rho/Rho-kinase signaling pathways for the anti-angiogenic activity of CA4²³⁶. Poor water-solubility, low bioavailability, and rapid metabolism are the main limitations that could be overcome by nano-formulations of CA4. Up to now, a series of nano-based systems (nanoliposomes and oil nanodroplets) have been developed to enhance the bioavailability of CA4 ^{237, 238}. Co-delivery of CA4 with other chemotherapy agents (e.g., DOX) using iRGD-grafted mesoporous silica nanoparticles was studied to destroy and kill tumor cells and vasculature ²³⁹. Recently, Wang et al. tested coadministration of CA4 nanoparticles and sorafenib to treat hepatocellular carcinoma ²⁴⁰. The authors developed nanoparticles of poly(L-glutamic acid)-graft-methoxy poly(ethylene glycol)/CA4 sodium salt (CA4-NPs) combined with sorafenib. The rationale was that the CA4-NPs

could disrupt established tumor blood vessels and result in extensive tumor necrosis, while sorafenib could reduce VEGF-A-induced angiogenesis (induced by CA4-NP) and lead to the inhibition of tumor proliferation (see **Fig. 8**). The results showed that the combination therapy with sorafenib 30 mg/kg + CA4-NPs 30 mg/kg (on the CA4 basis) could lead to a significant tumor suppression (over 90%) in an orthotopic hepatic H22 xenograft mouse model; and 5 out of 7 mice receiving the combination therapy survived tumor-free for 96 days.

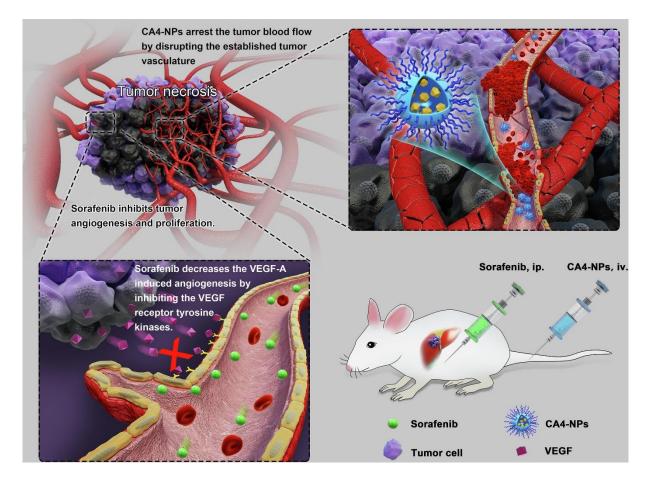


Fig. 8 Schematic representation of the combined mechanism of CA4-NPs and sorafenib to treat hepatocellular carcinoma (HCC). As shown, although the disruption of established tumor blood vessels and extensive tumor necrosis are achieved by systemic administration of CA4-NPs, the overexpression of VEGF-A and thereby angiogenesis occurs in response to hypoxia. On the other hand, sorafenib can decrease the expression of VEGF-A and hence subsequently inhibit angiogenesis and tumor proliferation. This strategy could be considered as a potential approach to completely eradicate the whole tumor. Reproduced with permission from ref ²⁴⁰.

8. Lipid-based nanosystems

Liposomes, solid-lipid nanoparticles (SLNs), self-emulsifying drug delivery systems (SEDDSs), and micelles are the major types of lipid nanoparticles with the ability to load and deliver various chemicals, drugs, and genes used in cancer diagnosis and therapy ²⁴¹. They exhibit some attractive properties as DDS, such as biocompatibility, biodegradability, capacity to selfassemble, as well as the ability to entrap both hydrophobic and hydrophilic drugs ²⁴². In addition, it is easy to tailor their size, functionality, and surface charge via simple approaches ^{243, 244}. Liposomes are FDA-approved self-assembled phospholipid vesicles composed of lipid bilayers surrounding an aqueous core, and they can be produced in a size range of 30 nm to 3000 nm 245 . The loading of bioactive substances with various chemical structures into liposomes can include: (1) hydrophilic drugs in the aqueous core; (2) lipophilic drugs inside the lipid bilayer; and (3) amphiphilic drugs partitioned at the surface of the inner or outer bilayer ^{246, 247}. Active targeting using liposomes is achievable using surface modification with target-specific ligands or antibodies ²⁴⁸. In addition, stimuli-responsive liposomal DDSs are under investigation. ThermoDox is a temperature-responsive nano-liposome used for un-resectable hepatocellular carcinoma in Phase III clinical trials ²⁴⁹. Doxil[®], the first FDA-approved nano-drug, is a liposomal doxorubicin formulation used for the treatment of various cancers, like Kaposi's sarcoma²⁵⁰. Moreover, there are additional FDA-approved liposomal drug formulations for cancer therapy on the market, including MyocetTM, Lipo-dox[®], DaunoXome[®], and Marqibo^{® 251-254}. With respect to anti-angiogenic applications, several research groups have shown the ability of liposomes. For example, Pont et al. showed the effectiveness of Fumagillin (an anti-angiogenic drug)-loaded liposomal nanoparticles to treat early atherosclerotic lesions in mice ²⁵⁵.

SLNs were firstly introduced in 1991 with the goal to create a carrier system as an alternative to traditional colloidal carriers (e.g., emulsions and liposomes) ²⁵⁶⁻²⁵⁸. However, there are some limitations to use of the SLNs as DDSs, including their rapid clearance, serum instability, as well as nonspecific uptake by the mononuclear phagocytic system ²⁵⁸. In this regard, functionalizing the SLNs using a variety of bioactive molecules, including ligands and antibodies, has been suggested to improve their potential in targeted drug delivery ²⁵⁹⁻²⁶¹. Recently, Bayón-Cordero et al. reviewed the application of SLNs in anti-cancer drug delivery, with the advantages of biocompatibility, high bioavailability of encapsulated drugs, possible loading of many hydrophilic and lipophilic molecules, and relatively easy large-scale production ²⁶². The use of SLNs for the loading and delivery of anti-angiogenic agents has been confirmed by several research groups. As one example, VEGF antisense oligonucleotides were successfully loaded into SLNs, and tested *in vitro* and *in vivo* rat glioma models showing down-regulation of VEGF expression levels ²⁶³.

SEDDSs are multi-component systems composed of an oil phase, surfactants, cosurfactants, emulsifying agents, and co-solvents ²⁶⁴. Based on their size, two types of these systems are self-nano-emulsifying agents (SNEDDS) and self-micro-emulsifying agents (SMEDDS) (**Fig. 9**) ^{264, 265}. Up to now, various chemicals and drugs have been successfully loaded into SEDDSs, including anti-cancer agents, and there are more than four such commercialized drug products on the market ^{264, 266, 267}. In 2015, Valicherla et al. prepared docetaxel (DCT) loaded SEDDSs (D-SEDDS) to improve the oral bioavailability and therapeutic efficacy of the drug. The results showed a 3.19-fold increase in bioavailability of the D-SEDDS in rats and a 25-fold increase *in vitro* cytotoxic activity compared to free DCT ²⁶⁸. In order to obtain more effective

anti-angiogenic formulations, several groups have incorporated anti-angiogenic substances (e.g. curcumin) into SEDDSs, and the results have been promising ²⁶⁹⁻²⁷¹.

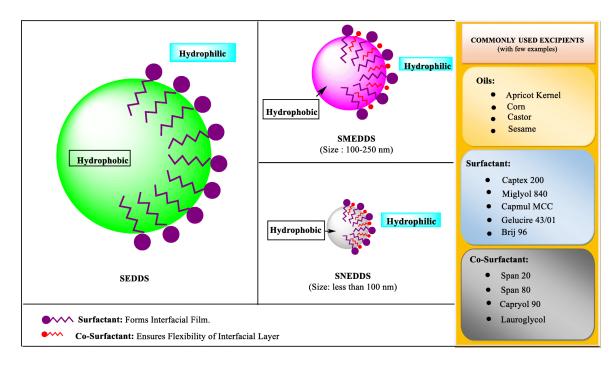


Fig. 9 Illustration of some lipid-based nanosystems, including self-emulsifying drug delivery system (SEDDS), self-micro-emulsifying drug delivery system (SMEDDS), and self-nano-emulsifying drug delivery system (SNEDDS). Reproduced with permission from ref ²⁶⁴.

9. Polymeric nanofibers

Polymeric nanofibers are among the most widely-applied constructs in biomedicine, from anti-tumor strategies to tissue healing. Nanofibers exhibit some attractive properties, including large specific surface area, controllable pore size, and tunable drug release profiles, making them highly-promising candidates for anti-cancer applications ²⁷². Recently, Abid et al. reviewed the anti-cancer applications of electrospun polymeric nanofibers loaded with various chemicals and drugs including, doxorubicin, paclitaxel, and curcumin ²⁷³. Apart from electrospun nanofibers, several studies showed the utility of synthetic nanofibrous peptide scaffolds to mimic the pro-

angiogenic and anti-angiogenic activity of small molecules, including heparin, and maspin ²⁷⁴⁻²⁷⁶. For instance, Fan et al. investigated docetaxel- and curcumin-loaded nanofibrous microspheres made of PLA-PEO-PPO-PEO-PLA polymers as an injectable and sustained-release system for enhancing anti-colon cancer activity ²⁷⁷. The results of the combined nanofibrous microsphere treatment showed a significant increase in the inhibition of angiogenesis and subsequent inhibition of colon cancer in mice. On the contrary, there are a number of publications in which pro-angiogenic cargos were delivered using polymeric nanofibrous scaffolds produced by both electrospinning and self-assembly procedures ^{136, 278-281}. Most of the pro-angiogenic nanofibers have been applied to accelerate tissue repair and regeneration, especially to promote wound healing ²⁸²⁻²⁸⁴.

10. Other carbon-based nanomaterials and nano-systems

Nano-sized carbon-based materials are among the most promising DDSs and include several members, including carbon nanotubes, nanodiamonds, nanohorns, graphene, fullerenes, and nanofibers ²⁸⁵. These nanomaterials show attractive properties; for example, they typically possess high mechanical strength and large specific surface area, and thus provide numerous sites for chemical or physical conjugation; moreover, they are relatively easy to manufacture on a large scale ^{286, 287}. These nanomaterials in either pristine or functionalized formats can be suitable platforms for conjugation, loading and release of a wide range of bioactive molecules ^{288-²⁹⁰. Additionally, some carbon nanomaterials especially carbon nanotubes and graphene are being studied in laser-induced hyperthermia of different types of solid tumors ²⁹¹.} The use of carbon-based nanomaterials in anti-angiogenic cancer therapy is growing. One of the first reports was published by Muruges et al. who showed that 100 µg of graphite, multi-walled carbon nanotubes (MWCNT), and fullerenes could significantly inhibit angiogenesis induced by FGF2 or VEGF *in vivo* in a CAM model assay ²⁹². In a comprehensive study, Wierzbicki et al. evaluated the anti-angiogenic properties of diamond nanoparticles, graphite nanoparticles, graphene nanosheets, MWCNT, and C60 fullerenes at a concentration of 500 mg/L in a CAM assay, ²⁹³. Their results revealed the anti-angiogenic effects of diamond nanoparticles and MWCNTs. However, graphite nanoparticles and graphene showed no anti-angiogenesis activity, and interestingly fullerenes exhibited pro-angiogenic activity.

With respect to the interactions of single-wall carbon nanotubes (SWCNTs) with endothelial cells, Albini et al. concluded that these nano-sized carbon materials could be useful vehicles for targeting the vasculature and potential carriers of anti-angiogenic agents ²⁹⁴. Masotti et al. in 2016 reported that polyethyleneimine (PEI) and polyamidoamine dendrimer (PAMAM)-coated carbon nanotubes (CNTs) were appropriate delivery systems for microRNAs (miR-503 oligonucleotides) for angiogenesis regulation ²⁹⁵. More recently, Su et al. designed and developed a dual-targeted co-delivery system based on iRGD-modified MWCNTs for use in anti-angiogenic therapy of lung cancer ²⁹⁶. For this aim, polyethyleneimine (PEI) and cystamine (SS) were used to attach iRGD and the chemotherapy drug candesartan (CD) to MWCNTs, respectively. Then, the authors assembled functionalized MWCNTs with the plasmid AT2 (pAT2) and prepared iRGD-PEI-MWNT-SS-CD/pAT2 complexes. The results obtained from *in vivo* experiments in nude mice demonstrated that co-delivery of CD and pAT2 synergistically increased anti-angiogenic effects through down-regulation of VEGF (see **Fig. 10**). However, some

reports showed that SWCNTs could promote angiogenesis through an indirect pathway in which SWCNTs enhanced fibrogenesis in mammalian cells (e.g., CRL-1490) via reactive oxygen species (ROS)-mediated phosphorylation of p38MAPK and, thereby, overexpression of pro-angiogenic molecules TGF-β1 and VEGF ²⁹⁷. However, other researchers have reported conflicting results; for example, Roman et al. observed that SWCNTs inhibited angiogenesis *in vivo* and was harmful to the normal embryonic development due to deregulation of important genes involved in cell proliferation, apoptosis, survival, and angiogenesis in brain and liver tissues ²⁹⁸.

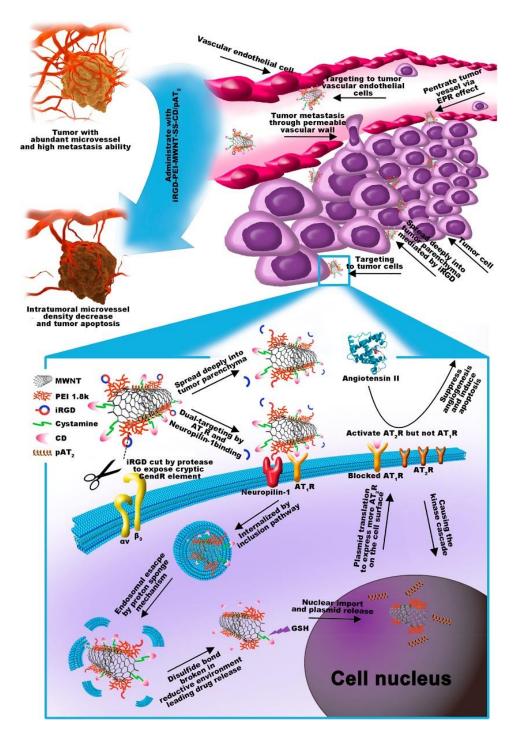


Fig. 10 Schematic representation of the use of iRGD-PEI-MWNT-SS-CD/pAT2 for the inhibition of tumor angiogenesis. Intravenous administration of iRGD-PEI-MWNT-SS-CD/pAT2 complexes results in specific accumulation at tumor tissues via EPR effect; angiotensin II type 1 receptor (AT1R) and integrin receptor-mediated binding. Reproduced with permission from ref ²⁹⁶.

There are several reports in the literature on the use of modified graphene oxide (GO) in anti-angiogenic strategies; for example, Lai et al. prepared bovine serum albumin-capped GO (BSA-GO) which was able to entrap and block VEGF-A₁₆₅ (a potent pro-angiogenic molecule), and thereby inhibit angiogenesis ²⁹⁹. Another example was provided by a study conducted by Shi et al. who conjugated reduced GO (rGO) with ⁶⁴Cu, 1,4,7-triazacyclononane-1,4,7-triacetic acid (NOTA), and the anti-CD105 antibody TRC105 to produce an appropriate system for theranostics ³⁰⁰. On the other hand, there have been several reports reporting the use of GO nanoparticles to promote angiogenesis. As one example, Mukherjee et al. showed that low amounts of GO (10 ng/mL) and rGO (50 ng/mL) could improve angiogenesis via the formation and activation of ROS and reactive nitrogen species (RNS) and consequent activation of Akt and eNOS signaling pathways ³⁰¹. Moreover, Chen et al. showed that SrTiO₃ CNTs could be used as a delivery system for Ag₂O nanoparticles to exert antibacterial, osteogenic, and pro-angiogenic activities simultaneously ³⁰².

Nanodiamonds are another type of carbon-based nanomaterials that can act as platforms in cancer nanomedicine, both for therapy, and imaging. Nanodiamonds are biocompatible, and show efficacy as carriers for various cancer therapeutic drugs, and possess tunable surface structures ³⁰³⁻³⁰⁵. For example, Setyawati et al. used surface-modified nanodiamonds to induce endothelial permeability. They functionalized the samples with –COOH and –NH₂ groups and showed that these derivatives could induce endothelial leakiness in a surface-dependent manner, resulting in increased delivery of doxorubicin to tumors. The mechanism proposed for this phenomenon (i.e., leakiness of the vascular barrier) was based on an increase of intracellular ROS and Ca²⁺, which facilitated the loss of cell-cell interconnections in the vascular barrier caused hy cytoskeletal remodeling (see **Fig. 11**). Zhang et al. used lipid-coated nanodiamonds to enhance the bioavailability and efficacy of an anti-angiogenic drug, sorafenib, to combat metastasis of gastric cancer ³⁰⁶. The authors successfully prepared sorafenib-loaded nanodiamonds with a size of 127.6 ±12.9 nm. The drug-loaded nanodiamonds led to increased bioavailability (up to 7.64 fold) and a higher concentration of sorafenib in the tumor (up to 14.95 fold) in vivo compared to control groups. These improvements showed a significant suppression of the metastasis of gastric cancer to distant organs (liver and kidney). Furthermore, other research groups have studied nanodiamonds in pro-angiogenic strategies, for the loading and delivery of a broad range of pro-angiogenic molecules ³⁰⁷⁻³⁰⁹.

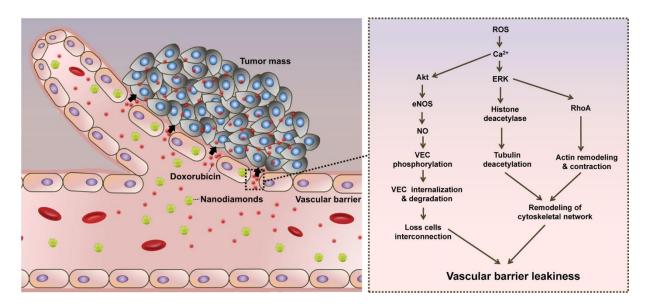


Fig. 11 Schematic representation of nanodiamond (ND)-induced vascular barrier leakiness. ND-induced vascular barrier leakiness leads to higher accumulation of doxorubicin in the tumor site. The increase of intracellular ROS and Ca²⁺ account for the ND-induced vascular barrier leakiness through the loss of cell-cell interconnection in the vascular barrier and cytoskeletal remodeling. Reproduced with permission from ref ³¹⁰

Carbon nanohorns have a conical structure, and are used in drug delivery strategies, both in pristine and functionalized formats ^{311, 312}. The main member of nanohorn family is the single-walled carbon nanohorn (SWNH), which is a tubular unit with a size of 2–5 nm in diameter and

40–50 nm in length ^{313, 314}. Although SWNHs have some properties in common with the CNTs, they exhibit possess more uniform and controllable morphology, and easier large-scale production without metal contamination, making them preferable in the clinical setting ^{315, 316}. Different morphologies of SWNHs have been identified, including "dahlia-like" type, "bud-like" type or "seed-like" type. The dahlia-like SWNHs are most commonly-used type for cancer theranostic applications ³¹⁴. Several reports have shown the applicability of modified SWNHs as DDSs for the delivery of anti-cancer drugs *in vitro* and *in vivo* ³¹⁷⁻³¹⁹. For example, Li et al. reported the use of oxidized SWNHs (oxSWNHs) as an effective DDS for transporting higher doses of vincristine to tumors ³²⁰.

Fullerenes is the first symmetric closed-cage type of the carbon nanomaterial family and have been extensively used in a variety of forms (number of C-atoms, pristine, surface-modified, and hybrid compounds) in different industrial and biomedical areas, including cancer imaging and therapy $^{321-324}$. It has been reported that fullerenes can act as anti-cancer agents on their own; for example, Prylutska et al. reported that water-soluble C₆₀ fullerenes were effective in the treatment of transplanted malignant tumors. They believed that the anti-cancer activity of C₆₀ fullerenes might be related to their high antioxidant activity, and their ability to block some specific cell receptorssuch as EGFRs. The anti-tumor activity of other fullerene derivatives has also been verified in other studies. Jiao et al. studied the anti-tumor and anti-metastatic potential of fullerenol in a mouse breast cancer model 325 . They injected 0.1 mL saline solution containing fullerenol C₆₀(OH)₂₀ (0.08 and 0.4 mg/ml) daily for a period of 16 days and histopathologically evaluated the anti-tumor and anti-metastatic activities of the samples. The results showed that injection of fullerenol modulated oxidative stress and down-regulated the

expression of multiple angiogenic factors (e.g., CD31) in tumors, leading to inhibition of tumor growth and metastasis *in vivo* (Fig. 12).

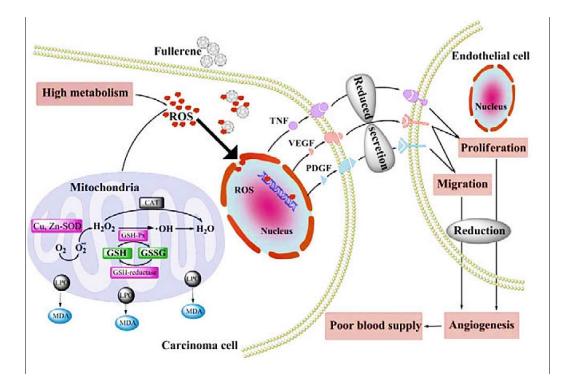


Fig. 12 Schematic representation of the molecular mechanisms involved in anti-cancer effects of C60(OH)20 *in vivo*. Reproduced with permission from ref ³²⁵.

Various surface-functionalized fullerenes (e.g., $Gd@C_{82}(OH)_{22}$, $C_{60}(OH)_{22}$ and $C_{60}(C(COOH)_2)_2$) also showed ROS scavenging properties and hence were potentially applicable in cancer therapy ³²⁶. Moreover, fullerene derivatives have exhibited potent anti-angiogenic activity; Meng et al. reported that the multiple hydroxyl group-functionalized surface of $Gd@C_{82}(OH)_{22}$ fullerene-based nanoparticles (f-NPs) exhibited the ability to simultaneously down-regulate more than 10 pro-angiogenic factors ar both the mRNA and protein levels ³²⁷. These researchers evaluated the *in vivo* efficacy of the functionalized NPs, and found that the surface-modified samples could reduce tumor microvessel density by > 40% as well as efficiently

decrease the speed of blood flow to tumors by up to 40% at 2 weeks post-injection compared to the effect of paclitaxel alone. Moreover, the functionalized NPs had no pronounced toxic sideeffects in nude mice. Based on these results, the authors concluded that this nano-sized compound holds great promise for use in cancer treatment.
 Table. 3 Summary of polymeric and carbon-based nano-sized drug delivery systems for pro- and anti-angiogenic therapeutic strategies.

Nanotechnology platform	Chemical structure	Modification (s)	Therapeutic agent	Effects*	Remarks	Ref (s)
PEG		Cell adhesive (RGDS) and MMP-degradable (GGGPQGflIWGQGK) peptides conjugated PEG	-	PA	- PEG hydrogels could support the lumen formation, expression of ECs proteins (e.g., eNOS), and perivascular investment of PDGFR- β and α -SMA positive cells by 2 weeks of co-culture	80
PLA		APT _{EDB} peptide functionalized pegylated PLA NPs	PTX	AA	 Significantly elevated cellular accumulation of PTX loaded NPs via energy-dependent, caveolae and lipid raft- involved endocytosis. In vitro tube formation assay and in vivo matrigel angiogenesis analysis confirmed a significant improvement in the antiangiogenic ability of PTX. 	84
Pegylated PCL		CGKRK peptide- functionalized pegylated PCL NPs	ΡΤΧ	AA	 An enhanced accumulation via an energy-dependent, lipid raft/caveolae-mediated endocytosis with the involvement of microtubules in HUVECs. An energy-dependent, lipid raft/caveolae-mediated endocytosis with the participation of Golgi apparatus in human U87MG cells. 	92
PEGylated PLGA	$\begin{bmatrix} 0 \\ -1 \\ -1 \\ -1 \end{bmatrix}_{x} \begin{bmatrix} -1 \\ 0 \\ -1 \end{bmatrix}_{y}$	RGD peptide and RGD- peptidomimetic functionalized pEGylated PLGA NPs	ΡΤΧ	AA	 Higher affinity to HUVECs by binding to αvβ3 integrin was observed in the functionalized NPs containing PTX Successful in vivo targeting to transplantable liver tumors was obtained in the case of the functionalized NPs containing PTX, leading to prolonged survival times of mice 	95
Chitosan		Hyaluronic acid coated chitosan NPs	PLXDC1 small interfering siRNA	AA	 Significant inhibition of tumor growth in A2780 tumor- bearing mice Significant decrease in microvessel density 	110
		-	Ursolic acid (UA)	АА	 Inhibition of the angiogenesis in CAM model and H22 xenograft model Controlled release of UA and thereby its reduced side effects 	111

Heparin	$HO_{S}O_{O}OH_{O}OH_{S}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OH_{O}OHOHOOHOOHOOHOOHOOHOOHOOHOOHOOHOOHOOHO$	Cyclic RGD-modified heparin-lithocholic acid (HL)	-	AA	 Significantly inhibition of adhesion and migration of ECs Prohibition of the formation of tubular structures of ECs 	125
		Heparin-surface modified polyurethane (PU) macroporous discs	VEGF ₁₆₅	PA	- Accelerated neovascularization and tissue repair in tread animals with PU containing high (6.6 mg/g) heparin content immobilized VEGF ₁₆₅	126
Albumin		Abituzumab (DI17E6)- coupled NPs	DOX	AA	- DI17E6 coupled NPs specifically targeted αvβ3 integrin positive melanoma cells - DI17E6 coupled NPs containing DOX Inhibited angiogenesis by targeting of endothelial cells	151
Gelatin	² -₩qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-₩-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%-qu-2-%- ₩-4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+4\fmatex+	Electrospun gelatin nanofibers	bFGF	PA	- Capillary formation was improved as a function of bFGF loaded aligned or random nanofibers	136
	ан, н () (н, 50, е н () мн мн мн мн	PEG-modified thiolated gelatin NPs	sFlt-1 (VEGF-R1) plasmid DNA	AA	- Successful suppression of tumor growth and microvessel density	134
PAMAM dendrimer		RGD-4C peptide conjugate	-	AA	- Taken up by cells expressing $\alpha V\beta 3$ receptors, providing suitable imaging agents and/or chemotherapeutics to angiogenic tumor vasculature	328
Multi-walled carbon nanotubes (MWCNT)		Polyethyleneimine (PEI) or polyamidoamine dendrimer (PAMAM) functionalized SWCNTs	miR-503 oligonucleotides	AA	 Reduced toxicity for both polymer-coated SWCNTs in comparison to the pristine counterparts Efficiently delivery of miR-503 oligonucleotides to ECs Providing the possibility to regulate ECs proliferation and <i>in vitro</i> and <i>in vivo</i> angiogenesis 	295

Single-wall carbon nanotubes (SWCNTs)		Polyethylenimine (PEI)- SWCNTs conjugations linked with candesartan	VEGF-targeted siRNA (siVEGF)	AA	 Highly inhibited tube formation of HUVECs Inhibition of tumor growth and tumor-associated angiogenesis repression 	329
Graphene oxide		Bovine serum albumin- capped graphene oxide (BSA-GO)	VEGF165	AA	 Showing high stability in physiological saline solution and having ultrastrong binding affinity to VEGF-A165 Inhibiting the proliferation, migration and tube formation of HUVECs Ability to strongly disturb the physiological process of angiogenesis in CAM model The capability of blocking VEGF-A₁₆₅-induced blood vessel formation in rabbit corneal neovascularization 	299
		Gelatin methacrylate (GelMA) containing GO	-	PA	 Increasing the proliferation and migration of keratinocytes Improved wound healing via promoted angiogenesis 	330
Nanodiamonds		Arg-Gly-Asp-Ser (RGDS) conjugated anodiamonds	VEGF-siRNA	AA	 Prolonged the release time of VEGF-siRNA by 6 folds Reducing the formation of the tubes and without any testable cytotoxicity 	307
Fullerene		Polyhydroxylated fullerenes	Doxorubicin (Dox)	AA	- Inhibiting ECs proliferation in vitro - Exhibiting antiangiogenic activity in zebrafish and murine tumor angiogenesis models	331

11. Inorganic ions, nanoparticles, and nano-systems for anti-angiogenic and pro-angiogenic applications

Many inorganic metallic elements are delivered to humans via normal nutrition or by therapeutic diets since they are known to have specific effects on cell metabolism and biological functions. Some of these elements have also been embedded in implantable/injectable nanomaterials, nano-systems for advanced nanotechnology-based therapies to control angiogenesis. This section deals with the chemical and biological functions produced by inorganic elements with regard to promoting or suppressing angiogenesis; furthermore, a description of the various biomaterials used (e.g., nanoparticles, nanotextured surfaces, hierarchical systems) is provided. Inorganic elements usually perform their angiogenesis-related functions after being released as solble ions; however, direct interaction between the surface of metallic nanoparticles and cells/biomolecules has also been reported in some cases (e.g. gold and silver nanoparticles) (see **Tables. 4** and **5**). Elements having an effect on angiogenesis, but exhibiting severe toxicity to animals and humans (e.g. arsenic, lead and mercury contained in industrial waste nano-particulates) have not been included in this section due to the lack of therapeutic significance.

11.1 Boron

Boron is a trace element playing diverse and vitally-important roles in many biological functions ranging from bone metabolism to anti-inflammatory activity, as comprehensively discussed by Pizzorno in a valuable review ³³². The first evidence of the role of boron in the context of angiogenesis was reported in 2002 by Dzondo-Gadet et al. ³³³, who examined the action of boric

acid at the molecular level using cell-free transcription systems (isolated placenta nuclei) and translation systems (wheat germ extracts). It was found that 10 mM of H_3BO_3 greatly increased mRNA synthesis associated with the translation of pro-angiogenic proteins like VEGF and TGF- β 1.

Based on these early results, boron-releasing bioactive glasses have been intensively investigated over the last two decades, and have been proposed as therapeutic implantable (nano)biomaterials for accelerating wound healing in tissue engineering applications ³³⁴. Bioactive melt-derived B₂O₃-CaO-based glasses are more reactive than silicate glasses upon contact with aqueous solutions and were found to rapidly release a large amount of Ca²⁺ ions into biological fluids, which is beneficial for skin regeneration because calcium promotes the migration of epidermal cells to the wound site ³³⁵. Melt-derived borate glasses with the composition 1605 (6Na₂O-12K₂O-5MgO-20CaO-4P₂O₅-51.6B₂O₃-0.4CuO-1ZnO wt.%) and 13-93B3 (53B₂O₃-6Na₂O-12K₂O- 5MgO-20CaO-4P₂O₅ wt.%) were shown to stimulate VEGF secretion in vitro ³³⁶. Furthermore, Durand et al. ³³⁷ doped 45S5 Bioglass[®] with 2 wt% of B₂O₃ and reported that the presence of boron in the ionic dissolution products stimulated the proliferation and migration of HUVECs, in vitro tubule formation, and the secretion of IL-6 and bFGF to a greater extent compared to the B-free control glass, thereby demonstrating the proangiogenic potential of borate ions. These in vitro results were confirmed in vivo by comparing the vascularization induced by the same materials in an embryonic quail chorioallantoic membrane (CAM) model ³³⁸. Higher expression of integrin $\alpha v\beta 3$ and greater blood vessel density were observed in response to implanted B-doped 45S5 glass. Researchers from Missouri University developed 13-93B3 nano-fibers (diameter in the range of 300 nm to 2 μ m) which, after being organized in a "cotton-candy" morphology, could be used as a dressing material to

treat full-thickness cutaneous wounds ³³⁹. An interesting mechanism was observed to explain the promotion of *in vivo* angiogenesis by this nanomaterial ³⁴⁰, i.e. the newly-formed blood vessels were attached to the hydroxyapatite micro-clusters that originated during the nanofiber degradation due to the glass bioactivity ³⁴¹. After implantation in rats for 22 days, significant regeneration of dermal, epidermal and subcutaneous tissues was reported. Seven out of 12 diabetic patients involved in a clinical study experienced complete healing of their chronic wounds with less scarring and equal or faster-wound closure rate (from 0.3 to 0.8 mm/day depending on the type of injury) compared to other more expensive wound treatments, such as vacuum-assisted systems ³⁴⁰. These 13-93B3 borate glass nano-fibers, trade-named DermaFuse[®], received Food and Drug Administration (FDA) approval for medical applications in 2016 and are currently marketed for treating wound injuries in animals ("RediHeal" veterinary product) as well as acute/chronic wounds in humans (Mirragen[®] Advanced Wound Matrix). At present, these commercial products are the only ones based on nano-bioactive glasses for use in soft tissue engineering and also as stimulators of angiogenesis. Further research is needed to fully elucidate all the biomolecular and biochemical aspects behind the pro-angiogenic effect of boron as well as its synergistic action with other relevant ions (e.g., Ca²⁺) released from these glasses on the complex process of wound healing.

11.2 Calcium

Calcium is one of the most important elements involved in the biological functions of mammals, such as participation in building the mineral phase of hard tissues (bone and teeth) and regulating bone homeostasis via various cell signaling pathways ³⁴². Some proteins, e.g.,

parvalbumin and calbindin-D, can bind to Ca²⁺ ions and store them, thus acting as calcium stores or buffers, and limiting free calcium diffusion in the intracellular environment ³⁴³. Pro-angiogenic factors like platelet-derived growth factor (PDGF), EGF, IGF-I, bFGF and VEGF are known to trigger a significant increase in the level of Ca^{2+} ions in different cell types ³⁴⁴⁻³⁴⁶. In this regard, it was shown that bFGF and VEGF (the most potent pro-angiogenic endogenous factors) could bind to different families of receptor tyrosine kinases (RTKs) which trigger intracellular calcium increases in endothelial cells ³⁴⁷. Fang et al. studied the role of calcium stored in fibroblasts isolated from pterygium, and reported that calcium-related signaling pathways were associated with persistent fibroblast proliferation and angiogenesis, as shown by the high density of blood vessels ³⁴⁸. Ca²⁺ ions can be typically released from all forms of calcium phosphate implants as well as melt-derived and sol-gel bioactive glasses (e.g., micro- and nanoparticles, scaffolds, coatings, fibers) upon contact with biological fluids in vitro and in vivo. The angiogenic properties of calcium phosphates in the context of bone regeneration have been recently discussed by Malhotra and Habibovic ³⁴⁹. The pro-angiogenic effect of bioactive silicate glasses in contact with both hard and soft tissues is well-known, but it is typically considered to be due to all the ionic dissolution products released from the glass, including silicate ions ³⁵⁰.

The bioactive borate glass 13-93B3, with a high CaO content, was recently used to fabricate nanofibrous scaffolds that significantly accelerated wound healing when implanted in both animals and humans ³⁴⁰. A possible explanation for this beneficial effect relied on the release of Ca²⁺ ions which stimulate angiogenesis and accelerate the migration of keratinocytes, thereby promoting skin regeneration. These cotton-candy borate glass nanofibres were also found to

impressively help the healing of long-term venous stasis ulcers in diabetic patients, who were unresponsive to conventional pharmacological treatment ³⁵¹.

11.3 Cerium

Cerium is a rare-earth metal that usually does not participate in biological functions; however, cerium oxide nanoparticles (nanoceria) have recently attracted interest in biomedicine due to their antioxidant properties and ability to act as a free radical scavenger in cells and tissues. Many chemical processes can be used to produce ceria nanoparticles, including hydrothermal methods, sol-gel, and polymer-assisted synthesis; these routes have been comprehensively reviewed by Kargozar et al. ³⁵². Researchers have also found interesting dual properties (stimulatory or antagonistic effect) of nanoceria in the context of angiogenesis. The oxygen-buffering capacity of nanoceria can be exploited to stabilize HIF-1 α , thereby promoting angiogenesis in vitro and in vivo ³⁵³. It was shown that the pro-angiogenic potential of ceria nanoparticles is markedly dependent on the surface valence states of cerium: specifically, high surface area and a high Ce³⁺/Ce⁴⁺ ratio make nanoceria more catalytically active for regulating the intracellular oxygen content, which leads to a stronger pro-angiogenic effect (see Fig. 13)³⁵⁴. Ceria nanoparticles were also found capable of stimulating the migration and proliferation of endothelial cells in vitro ³⁵⁵. Functionalization strategies have been carried out to further enhance the pro-angiogenic properties of nanoceria. Nethi et al. ³⁵⁶ synthesized nanoconjugates of organosilane-functionalized cerium oxide nanoparticles by using an ammonia-catalysed ethylene glycol-assisted precipitation method in an aqueous suspension of samarium-doped nanoceria conjugated with hydrophilic triethoxysilane (6-{2-[2-(2-methoxy-ethoxy)-ethoxy]-

ethoxy}-hexyl) moieties. The ceria/polymer nanoconjugates promoted endothelial cell viability and proliferation without eliciting any significant cytotoxicity and induced *in vivo* blood vessel formation in a chick embryo chorioallantoic membrane model. The p38-MAPK/HIF-1 α signaling pathway was proposed to be the mechanism governing the pro-angiogenic effect induced by these functionalized nanoparticles, which was greater compared to that associated with "conventional" nanoceria.

Ceria nanoparticles, synthesized by using gelatin as a stabilizing agent, retained their proangiogenic properties when embedded in electrospun poly(3-hydroxybutyrate-co-3hydroxyvalerate) membranes ³⁵⁷ and PCL scaffolds ³⁵⁸, as demonstrated by accelerated wound healing in rat models.

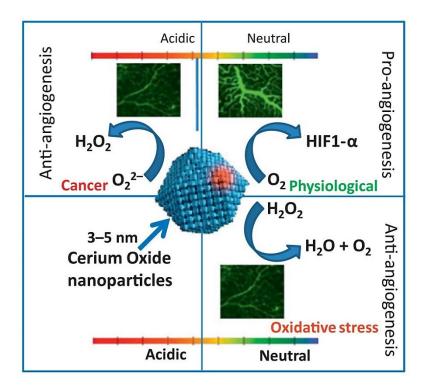


Fig. 13 Schematic illustration of the effect of environment on the pro-angiogenesis and antiangiogenesis properties of nanoceria. The pH, reactive oxygen species (ROSs) generation, and

intracellular oxygen concentration are identified as the main determinants of angiogenic behavior of nanoceria. From ref ³⁵⁴.

Conversely however, cerium oxide nanoparticles can exhibit an anti-angiogenic effect depending on the surrounding environmental conditions. There are some parameters that influence this dual behavior including pH, ROS generation, intracellular oxygen concentration and concentration of the nanoparticles ³⁵⁴. In fact, high concentrations of nanoceria exhibit anti-angiogenic effects ³⁵⁹. For example, the proliferation of HUVECs is reduced if the nanoceria concentration exceeds 8.6 mg/mL ³⁶⁰. It has also been shown that the anti-angiogenic effect is more pronounced when nanoceria is functionalized by heparin ³⁶¹: hence, heparin-functionalized and pristine nanoceria at high concentrations have been proposed as therapeutic agents for reducing endothelial cell growth and vascularization in tumors, thereby acting as an adjuvant in anticancer approaches.

Interestingly, there are some "shape and size effects" associated with ceria nanostructures. Das et al. ³⁵³ showed that exposure to ceria nanorods led to a slight reduction in endothelial cell proliferation, whereas spherical ceria nanoparticles or nanostars elicited no toxic effect in HUVECs. Furthermore, only ceria nanoparticles with size < 15 nm showed the potential to induce tubule formation, whereas micrometer-sized ceria particles even inhibited tube formation in HUVECs. This different behavior is probably due to the higher reactivity of smaller particles, due to a higher specific surface area. Cerium oxide nanoparticles are non-absorbable, but a controlled release of Ce³⁺/Ce⁴⁺ ions from soluble (nano)biomaterials should be studied in future research as a potential means to promote angiogenesis. At present, Ce₂O₃ has already

been incorporated in gel-derived silicate MBGs, but only its physico-chemical role in modulating glass dissolution kinetics and its biological effect in improving bone cell activity have been studied so far ³⁶²⁻³⁶⁴.

11.4 Cobalt

Controlled release of cobalt ions (Co²⁺) has been shown to promote angiogenesis *in vitro* and *in vivo*, via the creation of hypoxia-mimicking conditions. Specifically, Co²⁺ ions can activate the hypoxia-inducible factor-1 (HIF-1) pathway independently of the overall cellular oxygen level 365 . The HIF-1 pathway is the main regulator of cell response to variations in oxygen tension by triggering the expression of about 100 hypoxia-targeted genes ³⁶⁶. HIF-1 is a heterodimeric transcription factor comprising two subunits, i.e. the oxygen tension-regulated HIF-1 α and the constitutively-expressed HIF-1 β subunits ³⁶⁷. Activation of the HIF-1 pathway is strongly related to the concentration of HIF-1 α in the cytoplasm. Specifically, two scenarios are possible: (i) under normoxic conditions, HIF-1 α is continuously produced and then degraded through the ubiquitin proteasome pathway, or (ii) under hypoxic conditions, HIF-1 α is stabilized and can accumulate, translocate to the cell nucleus and then dimerize with HIF-1 β to induce the expression of its target genes. The role of Co^{2+} ions is to "artificially" stabilize HIF-1 α concentration by blocking the protein degradation regardless of the oxygen levels (see Fig. 14). As a result, broad transcriptional responses occur, including the upregulation of pro-angiogenic factors (e.g. VEGF) that subsequently lead to angiogenesis and improvement of the oxygen supply ³⁶⁸.

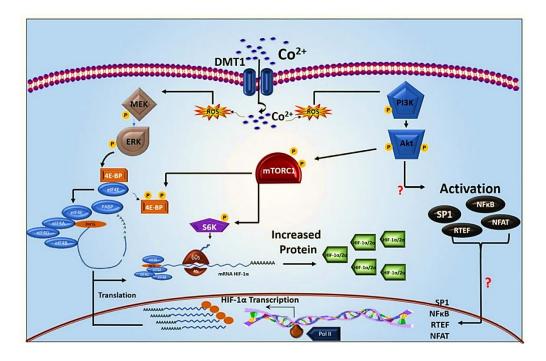


Fig. 14 Representative schematic of cobalt roles in activating two signaling pathways involved in angiogenesis progress, i.e., the PI3K/Akt/mTORC1 and MEK/ERK pathway. It is assumed that cobalt via Akt activation could trigger transcription factors including SP1, NF- κ B, RTEF and NFAT and thereby result in enhanced the transcription of HIF-1/2. Moreover, cobalt through activation of the MEK/ERK pathway could leads to the phosphorylation of 4E-BP and subsequent enhanced HIF-1 α translation. With permission from ref ³⁶⁹.

Altrhough it might have potential for inducing angiogenesis, the therapeutic use of cobalt is still under debate among researchers. Caution is suggested by the occurrence of systemic (and lethal in some cases) toxicity of cobalt ions released from Co-Cr alloys used in hip joint replacement prostheses. Systemic cobalt toxicity was reported to lead to neurotoxicity and heart failure; furthermore, local accumulation of Co²⁺ ions at the implant site could contribute to tumor formation ³⁷⁰⁻³⁷².

At present, there have been some reports about the incorporation of cobalt as a biological modifier in bioactive glasses. Most of these studies have been concerned with improving bone tissue repair and regeneration. Wu et al. ³⁷³ reported the fabrication of Co-

doped multiscale macro-mesoporous scaffolds by a co-templating sol-gel-like procedure where a surfactant (Pluronic P123) was used as a surface-directing agent to produce a mesoporous texture (average diameter 4.1 nm) and an open-cell polyurethane sponge as a template for the macropores (300-500 μm). These hierarchical MBG scaffolds showed promise as multifunctional systems for the synergistic delivery of antibiotics (ampicillin) and Co²⁺ ions, which stimulated VEGF over-expression in bone marrow mesenchymal stem cells (BMSCs). The relatively low amount (<5 mol.%) of cobalt incorporated in the MBGs was non-toxic to the BMSCs, but no *in vivo* studies have been reported on these materials so far.

An international research team led by Stevens and Hill synthesized a series of meltderived Co-doped silicate bioactive glasses with up to 4 mol.% of cobalt and demonstrated their hypoxia-mimicking function on human mesenchymal stem cells (hMSCs) ^{374, 375}. The ionic dissolution products released from these Co-doped glasses (particle size <38 μ m) successfully increased HIF-1 α activity after 8 h of incubation with hMSCs and promoted VEGF expression.

Given that hypoxia plays a key signaling role during cartilage formation, the same research group investigated the influence of Co-doped glasses on inducing hMSC chondrogenesis, in an attempt to develop a novel approach for cartilage tissue engineering ³⁷⁶. It was shown that reduced oxygen tension could aid chondrogenic differentiation ^{377, 378}. Because cartilage is a non-vascular tissue, oxygen must diffuse from the surface of the joint facing the synovial fluid into the cartilage due to the gradient of oxygen tension from the surface of the articular cartilage (partial pressure of oxygen 5%) to the subchondral bone (partial pressure of oxygen 0.1%) ³⁷⁹. Enhanced chondrogenesis in low-oxygen conditions is mainly mediated through HIF-1 α by inducing the expression of pro-chondrogenic genes (e.g. Sox9) ^{380, 381}.

Interestingly, the ionic dissolution products released from melt-derived Co-doped bioactive glass particles were reported to increase the level of HIF-1 α in hMSCs in a cobalt concentration-dependent manner but on the other hand, prolonged exposure to Co-containing solutions reduced cell proliferation and metabolic activity, as well as inhibited chondrogenic differentiation ³⁷⁶. This study suggested that the exposure time to cobalt needs to be taken into careful account and tailored to the specific application, since it can markedly influence the biological and biochemical processes of cells and tissues.

Kargozar et al. showed that incorporation of low amounts of CoO (up to 0.5 mol.%) in melt-derived silicate bioactive glasses increased the expression of angiogenesis-related genes during *in vitro* tests with HUVECs and Saos-2 cells, while causing minimal cytotoxicity after 21 days of culture ³⁸², and the overall bone healing was improved in rabbits at 4 and 12 weeks post-implantation compared to Co-free glass particles . The same research group also showed the importance of the size effect, i.e., fine Co-doped glass particles (9 µm) were more effective in promoting angiogenesis compared to large particles (725 µm), but were also associated with higher cytotoxicity due to more release of Co²⁺ ions ³⁸³. This effect must be taken into account if the use of nano-sized Co-doped glass particles is envisaged, because the higher specific surface area of the nanoparticles would be expected to make them more toxic.

The toxicity of Co-containing nanoparticles has been reported in many studies. It has been demonstrated that the inhalation of tungsten carbide (WC)/Co nano-powder, consisting of 80 to 90% of WC and 5 to 10% of metallic cobalt, could cause interstitial pulmonary disease and lung cancer, the mechanism involving the generation of ROS and DNA damage ³⁸⁴⁻³⁸⁶. In a recent study using normal human bronchial epithelial cells (BEAS-2B) and human lung adenocarcinoma

cells (A549), Liu et al. ³⁸⁷ reported that WC/Co nanoparticles at a concentration of 5 μ g/cm² induced ROS production which activated the Akt and ERK1/2 signaling pathways, and greatly increased the transcriptional activation of AP-1, NF- κ B, and VEGF, thereby promoting pathological angiogenesis. This suggests further caution in proposing cobalt nanomaterials for controlling angiogenesis due to the risks associated with their toxicity and carcinogenicity.

11.5 Copper

Copper is an essential cofactor in various enzymatic activities in animals and humans. It was demonstrated that Cu⁺/Cu²⁺ ions modulate the activity of several proteins and factors involved in angiogenesis (see **Fig. 15**), such as VEGF, fibronectin, angiogenin, collagenase, prostaglandin E-1, ceruloplasmin, FGF-1 and -2, which play important roles in the initiation (vasodilation and vascular permeabilization), maturation (endothelial cell proliferation, migration and morphogenesis), and regulation of blood vessel formation (ECM remodelling) ³⁸⁸. It was also reported that endothelial cells are stimulated to proliferate *in vitro* upon exposure to copper ions regardless of the VEGF levels, which demonstrates the inherent pro-angiogenic effect of copper ³⁸⁹.

There are two main signaling pathways involved in Cu-induced angiogenesis, i.e. (i) the hypoxia-inducible HIF-1 pathway (similar to cobalt); and (ii) the MAPK signaling pathway. The first mechanism is involved in the initiation of the angiogenesis process ³⁹⁰, while the latter plays a role in the endothelial cell proliferative phase ³⁹¹.

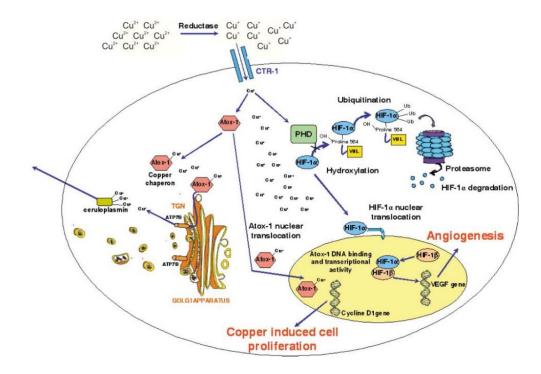


Fig. 15 Schematic representation of the angiogenesis regulation by copper ions. As seen, the entrance of copper into the cells is mediated by the copper transporter Ctr-1 protein and its delivery to intracellular proteins is regulated by copper transport proteins (chaperones) such as Atox-1. By inhibiting PHD-mediated hydroxylation of HIF-1 α , copper facilitates the translocation of the factor into the nucleus, leading to its dimerization with HIF-1 β and subsequent interactions to hypoxia-responsive elements and VEGF gene over-expression. Reproduced with permission from ref. ³⁹².

Since copper is normally involved in physiological vascularization processes ³⁹³, its reduction or removal in patients suffering from cancer is being studied to combat malignancies as an antiangiogenic treatment ³⁹⁴. In fact, the reduction of copper levels by following a Cu-deficient diet or administering a Cu-chelating drug (e.g., penicillamine) can inhibit angiogenesis by "switching" endothelial cells back into the G0 phase or triggering apoptosis ³⁹⁵. Furthermore, copper reduction attenuates the pro-angiogenic activity of VEGF, bFGF, TNF, IL-1β, IL-6 and IL-8 ^{396, 397}

The use of copper ions as pro-angiogenic dopants in implantable (nano)biomaterials has recently been proposed to promote better vascularization of tissues in regenerative medicine. Since the goal is to achieve a sustained release of copper ions over time, biocompatible matrices with a relatively low dissolution rate, such as bioactive glasses, are often selected to deliver the copper ions. Bührer et al. ³⁹⁸ showed that melt-derived 45S5 Bioglass[®] doped with 1 wt.% of Cu increased angiogenesis in a rat arteriovenous loop model when compared to Cu-free 45S5 parent glass. Zhao et al. ³⁹⁹ synthesized Cu-doped borate glass microfibers and showed that their ionic dissolution products stimulated the migration of HUVECs, tubule formation, and VEGF secretion, along with the fibroblast expression of angiogenic genes, to a greater extent as compared to the Cu-free parent glass. This pro-angiogenic effect was proportional to the amount of copper in the glass composition, and the Cu-doped fibers were found to accelerate the healing of full-thickness skin wounds in rats. The pro-angiogenic effect of Cu-doped borate materials was also studied by Bi et al. ⁴⁰⁰ in a rat calvarial defect model. Specifically, meltderived bioactive borate glass (13-93B3 composition) was doped with 0.4 wt.% of copper and used to fabricate porous scaffolds with trabecular, unidirectional or fibrous microstructures. It was reported that the percentage of new blood vessels at 12 weeks post-implantation was higher for Cu-doped 13-93B3 scaffolds compared to Cu-free 13-93B3 control implants with the same porous architecture. The glass fibrous scaffolds exhibited the best pro-angiogenic effect of all these porous microstructures. Similar results were obtained by the same research group in a dorsal skin window model in mice implanted with Cu-doped or Cu-free 13-93B3 glasses ⁴⁰¹.

Wu et al. 402 prepared Cu-doped MBG scaffolds with hierarchical porosity (interconnected large pores within 100-500 μ m and well-ordered mesoporous channels around

5 nm) and reported that their ionic dissolution products (mainly Cu^{2+} ions) stimulated HIF-1 α and VEGF expression in human BMSCs, thus further supporting the suitability of mesoporous materials as platforms for the controlled release of pro-angiogenic ions.

Although most applications of copper for promoting angiogenesis are in the field of bone regeneration, Baino first suggested in 2015⁴⁰³ that Cu-doped MBGs could also be used to accelerate the vascularization of porous orbital implants for ophthalmic socket surgery. This hypothesis was actually confirmed *in vivo* in 2018 by Wang et al. ⁴⁰⁴, who performed primary angiogenic tests in a panniculus carnosus muscle model in rabbits and reported that Cu-doped MBG coating significantly promoted the vascularization of porous hydroxyapatite orbital implants compared to Cu-free materials. Incorporation of copper in glass-ceramic orbital implants was also reported using a nearly-inert alumino-silicate glass-ceramic as a base material ⁴⁰⁵. In a first approach, melt-derived Cu-doped macroporous scaffolds were produced by sponge replication, but the release of copper ions was insufficient to elicit any pro-angiogenic effect. The second strategy, involving the deposition of a thin Cu-doped MBG nano-textured layer on the struts of silicate glass-ceramic foams, permitted a more sustained release of copper to be achieved. At present, none of these Cu-releasing (nano)systems have been tested in preclinical studies, but the early results achieved so far are promising and require further research.

A few studies have addressed the therapeutic properties of metallic copper nanoparticles. Chen et al. ⁴⁰⁶ reported that the lethal dose (LD₅₀) of copper nanoparticles (size 23.5 nm), micro-sized particles (17 μ m) and Cu ions in mice were 413, over 5000 and 110 mg/kg body weight, respectively. Since copper nanoparticles showed higher *in vivo* biocompatibility than copper ions, Mroczek-Sosnowska et al. ⁴⁰⁷ extended the investigation to understand if this

also applied to pro-angiogenic properties (**Fig. 16**). It was found that the pro-angiogenic effect of commercial colloidal copper nanoparticles in a chick embryo chorioallantoic membrane model was actually more potent than that elicited by CuSO₄ salt, thus confirming the hypothesis.

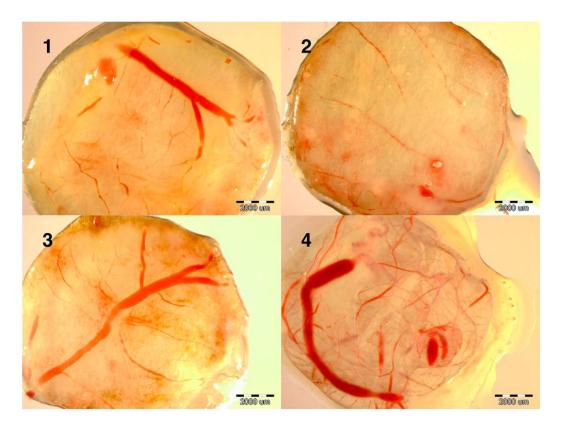


Fig. 16 Pro-angiogenic effect of copper nanoparticles: images of implants maintained in the chicken embryo chorioallantoic membrane (CAM) for 2 days, soaked with (1) control (non-soaked); (2) control (PBS); (3) $CuSO_4$; (4) nano-copper, evaluated at day 12 of incubation. Scale bars, 2000 μ m. Reproduced with permission from ref ⁴⁰⁷.

11.6 Europium

Europium is a lanthanides element, with applications in the context of bioimaging due to its long-life fluorescence properties. In fact, Eu³⁺ ions were found to possess good luminescence with suitable brightness and prolonged signal intensity for use as biolabelling agents ⁴⁰⁸. Most

Eu-containing (nano)materials have therefore been used for luminescent imaging of cells and tissues ⁴⁰⁹, as well as for tracking and monitoring the kinetics of drug release from mesoporous biomaterials ⁴¹⁰. There are few studies about the role of europium in regenerative medicine, but it was reported that Eu³⁺ ions could functionally mimic Ca²⁺ ions, thus influencing bone remodelling and being potentially useful to treat bone density disorders (e.g. osteoporosis) ^{411, 412}. Patra et al. ⁴¹³ first showed that Eu(OH)₃ nanorods, synthesized via a microwave-assisted method, could enhance the proliferation of HUVECs *in vitro* and stimulate vascular sprouting *in vivo* in a chick CAM model. Similar results were also achieved when Eu(OH)₃ nanorods were embedded in electrospun nanofibrous PCL scaffolds ⁴¹⁴, which could be applied in tissue engineering (e.g., soft patches). The pro-angiogenic properties of these nanorods at low concentrations were associated to the production of ROS (especially H₂O₂) both in vitro (HUVECs) and in vivo (zebrafish model) (**Fig. 17**) ⁴¹⁵.

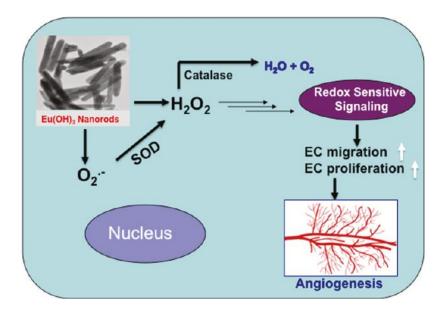


Fig. 17 Redox signaling mechanism proposed for the pro-angiogenic effect induced by $Eu(OH_3)$ nanorods in endothelial cells (EC). ROS (especially H_2O_2) are generated by the nanorods in the cytosolic part of the ECs, thus functioning as signaling molecules. Reproduced from ⁴¹⁵.

This signaling mechanism was confirmed, also using a zebrafish model, in another study dealing with $Eu(OH)_3$ nanorods and nanoparticles produced by hydrothermal treatment ⁴¹⁶.

 $Eu(OH)_3$ nanorods show great promise for therapeutic applications also considering their safety, as suggested by the absence of genotoxicty in a mouse model ⁴¹⁷.

Ma et al. ⁴¹⁸ prepared Eu-doped hydroxyapatite nanorods (length 40-60 nm, width 20-40 nm) by a precipitation method followed by annealing at 600 °C and found that the nanoparticles had a dose- and time-dependent inhibitory effect on HUVECs *in vitro*. However, this effect could be related to the needle-like morphology of the nano-hydroxyapatite (inherent "shape effect") rather than to a toxic release of europium ions.

Other Eu-doped crystalline nanomaterials (e.g., NaYF₄) exhibit poor biodegradability and can evoke adverse responses in cells and tissues (e.g., necrosis of bone cells) ⁴¹⁹, thus requiring the need for removal once they have performed their function in the body.

Clear evidence of the safety and pro-angiogenic effect of Eu³⁺ ions *in vitro* and *in vivo* was reported by Shi et al. ⁵⁸, who synthesized Eu-doped mesoporous silica nanoparticles (Eu-MSNs, size 280-300 nm) by adding TEOS and Eu(NO₃)₃·GH₂O) to a water/ethanol mixture, followed by a sol-gel process where cetyltrimethylammonium bromide (CTAB) was used as a structuredirecting agent. It was found that some angiogenesis-related genes (i.e., CD31, MMP9, VEGFR1/2, and PDGFR α/β) were significantly upregulated in HUVECs by Eu-MSNs. Results from *in vivo* experiments carried out in diabetic rats revealed that the Eu-MSNs could increase formation of blood vessels and capillary network in chronic skin wounds, showing superior proangiogenic ability compared to Eu-free MSNs (**Fig. 18**). As a result of enhanced neovascularization at the wound site, collagen deposition and re-epithelialization were also promoted. The same study also reported that Eu-MSNs underwent partial degradation in cell culture media – which could help to overcome the need for removal in the long term – and the release of Eu³⁺ ions stimulated new bone formation at critical-sized cranial defects in rats.

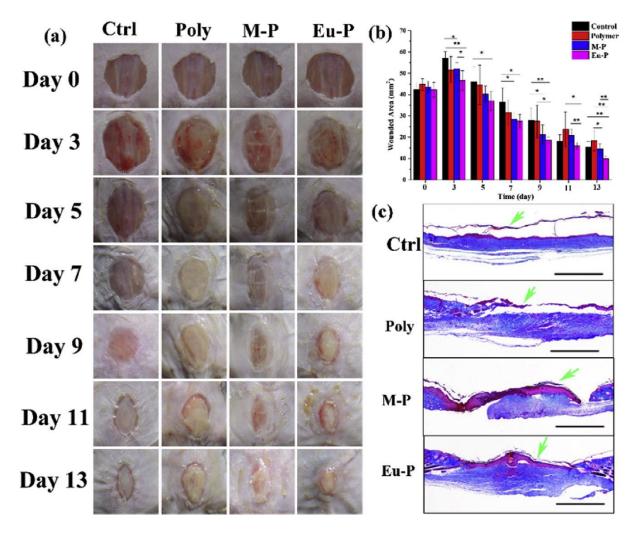


Fig. 18 Overview of the size change of the large excision wounds made in the dorsal skin of diabetic mice with different periods (a) and relevant statistical analysis (b). Eu-MSNs-polymer film (Eu-P) significantly accelerated the wound healing compared to other groups. Masson's Trichrome staining images (c) of wounds treated with different groups of films (blank control indicated as Ctrl, pure polymer film as Poly, MSNs-Polymer composite films as M-P and Eu-MSNs-Polymer composite films with as Eu-P). Green arrows indicates the newly formed epithelium at the wound site (scale bar 500 μ m, *p < 0.05; **p < 0.01). Reproduced with permission from ref ⁵⁸.

In summary, europium is a highly-promising metallic element to be incorporated into implantable nanospheres with multifunctional properties (angiogenesis and osteogenesis) for potential use in both bone and skin tissue engineering.

11.7 Gold

Gold nanoparticles, collectively called "nano-gold," are known to elicit an anti-angiogenic effect, and therefore have recently received attention to combat cancer. In fact, neovascularization outgrowth from preexisting blood vessels is well known to be the key to allowing the growth and progression of tumors ³.

The anti-angiogenic property of nano-gold was first reported by Mukherjee and coworkers who investigated the effects on HUVECs *in vitro* ⁴²⁰ and in mice ⁴²¹. Gold nanoparticles (size <220 nm) were prepared by mixing sodium borohydride with an aqueous solution of tetrachloroauric acid under vigorous agitation ⁴²². It was shown that nano-gold could interact with heparin-binding growth factors (HB-GFs) and inhibit their activity. These nanoparticles bound to vascular permeability factor (VPF)/VEGF-165 and bFGF , which resulted in inhibition of: (i) endothelial/fibroblast cell proliferation *in vitro;* and (ii) VEGF-induced permeability and angiogenesis *in vivo*. Gold nanoparticles also significantly inhibited VEGF receptor-2 phosphorylation, intracellular Ca release and RhoA activation *in vitro* but did not reduce the expression of VEGF-121 and epidermal growth factor, which is not a HB-GF. The ability of gold nanoparticles to inhibit the function of VEGF-165 (the most potent of the VEGF isoforms) ⁴²³ and placental growth factor (PIGF) was confirmed in another study by the same

research team ⁴²⁴. **Fig. 19** provides molecular mechanisms by which nano-gold affect angiogenesis process ⁴²⁵.

The inhibitory effect on pro-angiogenic factors VEGF-165, bFGF, and PIGF is related to the strong affinity of nano-gold for thiols, phosphines, disulfides and amines which are groups commonly present in HB-GFs ^{420, 421}.

An *in vitro* study with HUVECs revealed that the ability of gold nanoparticles to selectively disrupt the functions of pro-angiogenic HB-GFs was dependent on the particle size (inhibitory effect: 20 nm > 10 nm > 5 nm size of nano-gold) ⁴²⁶. Surface charge and chemistry of the nanoparticles were also reported to play an important role; naked nano-gold exhibited the maximum inhibitory effect towards HB-GFs as compared to the same nanoparticles functionalized with various charged ligands. This effect was mediated through direct binding of nano-gold to cysteine residues in HB-GFs. The resulting ionic/pseudo-covalent chemical bonds between the gold surface and HB-GFs induced a conformational change in HB-GFs mediating the inhibition of their function. On the contrary, no alternation was observed in the conformation of non-HB-GFs.

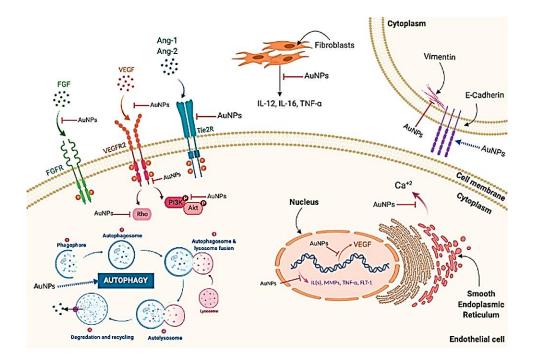


Fig. 19 Schematic illustration of molecular pathways affected by the anti-angiogenic effects of nano-gold (AuNPs). The main angiogenic pathways suppressed by nano-gold include VEGFR2, Tie2R, FGFR, and their downstream signaling pathways. As depicted, VEGF-165-mediated intracellular calcium release is suppressed by AuNPs. Moreover, AuNPs upregulate E-cadherin and downregulate vimentin, which results in reduced epithelial-to-mesenchymal transition (EMT) attenuating angiogenesis. Another anti-angiogenenic mechanism proposed for AuNPs is related to its ability to reduce ILs, MMPs, and TNF- α expression and inhibit neovascularization via induction of autophagy. From ref ⁴²⁵.

The literature strongly supports the suitability and potential of gold nanoparticles for cancer treatment, due to their biocompatibility, and ability to selectively interact with the biomolecular and biochemical processes of cancer cells and not healthy cells ⁴²⁷. Moreover, another useful property of gold (nano)compounds is the strong inhibitory effect on the enzyme thioredoxin reductase (anti-mitochondrial activity), which is involved in cancer cell proliferation ⁴²⁸.

Preclinical studies have been highly promising. Naked gold nanoparticles were reported to inhibit tumor growth and metastasis in a mouse model of ovarian cancer ⁴²⁹. This effect was due to the inhibition of the MAPK pathway (a key pathway for cancer cell proliferation) because

nano-gold disrupts the function of HB-GFs secreted by cancer cells. HB-GFs are also involved in the epithelial–mesenchymal transition (EMT), which is one of the main mechanisms behind cancer metastasis ^{430, 431}. A study carried out using colorectal cancer cells confirmed that treatment with 20-nm gold nanoparticles could actually reverse EMT, thus inhibiting tumor metastasis ⁴³². This was possible as nano-gold reduced the expression of EMT-associated proteins, and up-regulated E-cadherin and down-regulated Snail,.

Furthermore, nano-gold can be used for the treatment of B-cell lymphoblastic leukemia and prostate cancer. B-cell leukemia is a generally incurable disease characterized by resistance against apoptosis, partly because the leukemic cells continuously secrete VEGF and express VEGF receptors. Mukherjee et al. ⁴³³ reported that gold nanoparticles coated with anti-VEGF antibodies (Avastin or bevacizumab) increased the level of apoptosis in leukemic cells, and naked nano-gold was also able to induce the same effect to a more limited extent. An application of nano-gold for the theranostic treatment of prostate cancer was reported. Prostate-specific membrane antigen-targeted gold nanoparticles were loaded with a fluorescent photodynamic therapy drug and successfully tested *in vitro* and *in vivo* ⁴³⁴. This system aimed to provide surgical guidance for accurate resection of prostate cancer and additional photodynamic therapy when surgery was insufficient.

11.8 Iron

Iron plays a key roles as an enzymatic cofactor and as the central metal in heme in many physiological functions, the most important of which is oxygen transport. In fact, about 70% of the iron available in the body is stored in the blood in the form of hemoglobin, a metalloprotein 435 . Iron deficiency can induce the stabilization of HIF and the increase in VEGF secretion, thus stimulating angiogenesis 436 . In this regard, induction of iron deficiency was proposed to be an anticancer strategy to reduce the resistance of tumor cells to antiangiogenic therapies. It was reported that Fe²⁺ ions at concentrations within 10-100 μ M exhibit a relatively weak proangiogenic activity 437 , which is less pronounced than that of other transition metallic ions (e.g. Cu²⁺) 438 . On the contrary, Fe³⁺ ions have a stronger pro-angiogenic effect at the same concentrations, but are highly toxic to the cells, due to the oxidative stress and ROS generation associated with ferric ions 437 . This is probably the main reason why iron-releasing (nano)biomaterials have not been much investigated as implantable systems to promote local angiogenesis. However, some Fe-doped phosphate glass compositions have been suggested for use in tissue engineering as the incorporation of iron allows tailoring the phosphate glass dissolution rate to match that of tissue regeneration $^{439, 440}$.

At present, iron is mainly used in the form of magnetite (Fe₃O₄) nanoparticles that are employed as a diagnostic contrast agent in magnetic resonance imaging, and for cell labeling and tracking ⁴⁴¹. Superparamagnetic iron oxide nanoparticles (SPIONs) have also recently emerged as a promising clinical option to treat some types of tumors via magnetic hyperthermia after being injected into the patient's bloodstream ⁴⁴². Furthermore, there is interest in the development of multifunctional SPION-based systems with: (i) a magnetite core functioning as a contrast agent; (ii) a biocompatible coating; and (iii) a therapeutic outer layer conjugated to targeting ligands such as, nucleic acids, small molecules, or antibodies ^{443, 444}. However, possible toxicity associated with the non-degradable magnetite core of SPIONs is still a matter of debate,

including problems related to biodistribution, local accumulation and long-term fate of Fe₃O₄ nanoparticles *in vivo* ⁴⁴⁵.

11.9 Lithium

Lithium supplementation is a commonly-used clinical approach for the treatment of several psychiatric diseases including bipolar disorder, unipolar depression, and schizophrenia ⁴⁴⁶. In this context, Li⁺ ions act on the regulation of neurotransmitters and mitochondrial function, attenuating the expression of genes associated with signaling pathways such as protein kinases A and C (pKA/pKC) in hyperexcitable neurons, thereby favoring mood stabilization ^{203, 447}.

Lithium is also known to activate the Wnt/ β -catenin pathway to inhibit the glycogen synthase kinase (GSK)-3 β signaling pathway ⁴⁴⁸. The latter is involved in the suppression of nuclear factor-kB (NF- κ B) and activation of I κ B α kinase, c-Jun-N-terminal kinase, p44/p42 MAPK, and Akt via tumor necrosis factor (TNF) ⁴⁴⁹. In fact, it was observed that NF- κ B-regulated gene products such as, cyclooxygenase (COX)-2, cyclin D1, matrix metalloproteinase (MMP)-9, survivin, inhibitor-of-apoptosis protein (IAP) 1 and 2, Bcl-xL, Bfl-1/A1, and TNF receptor-associated factor 1 were generally increased upon exposure to Li⁺ ions.

While lithium was once thought only to be involved in vasculogenesis, but not in angiogenesis ⁴⁵⁰, recent studies have clearly demonstrated its pro-angiogenic effect as well. Lithium was reported to increase VEGF secretion in rat brain endothelial cells by a mechanism involving the PI3K and GSK-3β signaling pathways ⁴⁵¹. The proliferation, migration, and viability of endothelial cells were also shown to be stimulated by Li⁺ ions *in vitro* and *in vivo* through the activation of the Wnt/β-catenin canonical pathway ⁴⁵².

To the best of our knowledge, incorporation of lithium in implantable biomaterials in order to promote angiogenesis has been carried out in only one study. Haro Durand et al. ⁴⁵³ replaced up to 5 wt.% of Na₂O with Li₂O in melt-derived 45S5 Bioglass[®] microparticles, and reported that HUVECs showed a greater migratory/proliferative response and ability to form tubules *in vitro* upon exposure to Li-doped glasses compared to the Li-free parent material. In agreement with previous studies, they also observed the activation of the Wnt/ β -catenin signaling pathway with an increase in expression of the pro-angiogenic cytokines, insulin-like growth factor-1 (IGF-1) and transforming growth factor- β (TGF- β).

Most studies have been devoted to evaluating the physico-chemical and structural properties of Li-doped biomaterials and the biological effect of Li⁺ ions for stimulating osteoblast activity and, hence, osteogenesis ⁴⁵⁴⁻⁴⁵⁶. Han et al. ⁴⁵⁷ also demonstrated that Li⁺ ions released from silicate MBGs could promote the differentiation of periodontal ligament cells into cementoblasts and increase cementogenesis. The multifunctional properties of lithium are promising for new advanced therapies and indeed deserve further investigation in coming years.

11.10 Magnesium

There is no study dealing directly with the use of magnesium nanostructures to stimulate angiogenesis. However, Mg²⁺ ions are known to play a direct role in modulating inflammatory responses and microvascular functions. In this regard, Bernardini et al. ⁴⁵⁸ reported that Mg²⁺ deficiency inhibited the growth and migration of microvascular 1G11 cells while increasing some inflammatory markers such as interleukins 1a and 6, nitric oxide, which is a mediator of inflammatory responses, and VCAM which mediates monocyte/endothelial interactions. On the

contrary, high levels of Mg²⁺ ions stimulate the proliferation and migration of microvascular cells, and hence stimulate angiogenesis.

A couple of studies apparently support the stimulatory effect of Mg-releasing bioceramics and bioactive glasses on angiogenesis. Zhai et al. ⁴⁵⁹ reported that ionic extracts from akermanite, a Si-, Ca- and Mg-containing biocompatible ceramic, up-regulated the expression of genes encoding the receptors for pro-angiogenic cytokines and increased the expression level of genes encoding the pro-angiogenic downstream cytokines, as well as nitric oxide synthase and increased nitric oxide synthesis. It was also shown that akermanite implants promoted neovascularization in a rabbit femoral condyle model at both 8 and 16 weeks post-implantation. Spontaneous angiogenesis and tubule formation in human endometrial stem cells cultured with sol-gel SiO₂-CaO-MgO-P₂O₅ bioactive glass extracts were also reported by Shamosi et al. ⁴⁶⁰. However, in both studies the specific role of Mg²⁺ ions could not be isolated from that of other ionic dissolution products due to the lack of control experiments using Mg-free bioceramic/bioactive glass; therefore, the overall pro-angiogenic effect could be due to the synergistic effects of silicate, Ca²⁺ and Mg²⁺ ions that were collectively released by both materials.

11.11 Niobium

Niobium is used as an alternative to vanadium in Ti-based alloys for metallic orthopedic implants (Ti-6Al-7Nb vs. Ti-6Al-4V). Niobium was found to be less toxic to cells than vanadium, with lower inhibitory effects on the proliferation and viability of human osteoblasts, fibroblasts, and lymphocytes ⁴⁶¹. No reports were found on the use of niobium nanostructures in the context

of angiogenesis; however, there is one study dealing with the pro-angiogenic properties of Nb⁵⁺ ions released from bioactive melt-derived silicate glass-ceramic granules. Miguez-Pacheco et al. ⁴⁶² reported the results of cell culture tests using bone marrow stromal cells (ST-2) in contact with Nb-doped glass-ceramic extracts, which revealed enhanced VEGF secretion induced by the ionic dissolution products released from the Nb-doped material as compared to the parent glass. A complete picture of the biomolecular and biochemical mechanisms behind the pro-angiogenic action of Nb⁵⁺ ions is still to be obtained; nevertheless, incorporation of niobium in MBG platforms allowing a controlled ion release and modulation of angiogenesis deserves investigation in the future.

11.12 Phosphorus

Phosphorus is known to be important, along with calcium, in constructing the mineral phase (mainly hydroxyapatite) of bones and teeth ⁴⁶³. Moreover, the pro-angiogenic role exerted by phosphate ions has been shown in some studies. In this regard, phosphate ions were reported to increase the expression of MMP-2 and bFGF in the lungs of developing mice, thereby stimulating angiogenesis ⁴⁶⁴. It was also observed that high levels of phosphate ions could increase the expression of key pro-angiogenic genes such as forkhead box protein C2 (FOXC2, a regulator of vascular formation and remodeling ⁴⁶⁵, osteopontin (OPN, a cytokine-like factor associated with tumor angiogenesis ⁴⁶⁶ and VEGF in pre-osteoblastic cells ^{467, 468}. On the other hand, hyperphosphatemia can induce human endothelial cell apoptosis resulting from increased ROS generation and mitochondrial dysfunction ⁴⁶⁹.

Phosphate ions can be released from calcium phosphate implants, the effects of which on angiogenesis have been recently reviewed in the context of bone tissue engineering ³⁴⁹. Furthermore, most silicate bioactive glasses, such as 45S5 Bioglass[®], contain a moderate amount of P_2O_5 ; therefore, phosphate ions released from these materials could indeed contribute to their well-established pro-angiogenic potential ⁴⁷⁰.

In vitro studies using Cu- or Co-doped P₂O₅-based glasses revealed the pro-angiogenic properties of phosphate glasses ^{471, 472}. However, it cannot be ignored that both glass formulations incorporated metallic dopants (cobalt and copper) with potent pro-angiogenic effects, so the phosphate ions could have just supplied an adjuvant effect.

11.13 Selenium

Selenium is needed in normal nutrition or can be supplied as a dietary supplement in either elemental form or in inorganic salts or water-soluble organic compounds ⁴⁷³. Elemental selenium is contained in a number of different enzymes and proteins with wide distribution and broad physiological functions throughout the body ⁴⁷⁴. Selenium was found to elicit an anti-angiogenic effect *in vitro* and *in vivo*, which suggested its suitability as an adjuvant in anticancer therapeutic strategies. In this regard, the beneficial activity of selenium against cancer was first reported in the 1980s in a couple of animal studies in rodents ^{475, 476}. Since then, different metabolites of selenium have been shown to play a key role to protect cells against free radicals (glutathione peroxidase), regulate energy use (tri-iodothyronine deiodinase) and modulating the intracellular redox potential (thioredoxin reductase) ⁴⁷⁷.

Jiang et al. ⁴⁷⁸ reported that monomethyl selenium could inhibit the transformation of healthy prostatic epithelial cells into cancerous cells due to different effects including, decreased cell proliferation, apoptosis, and inhibition of angiogenesis. Selenium, administered in the form of inorganic salt (sodium selenite) or organic compound (methyl selenocysteine), was also shown to be effective to decrease the density of blood capillaries in mammary cancer in rats ⁴⁷⁹. However, the biomolecular and biochemical mechanisms behind this experimental evidence still remain unclear. In general, selenium is thought to be associated with: (i) decreased VEGF secretion by cancer cells; and (ii) direct apoptosis of the endothelial cells via inhibition of matrix metalloproteinase (MMP) activity ⁴⁷⁹. It is reasonable to hypothesize that both effects may be due to Se-induced redox regulation of the activity of transcription factors, or redox modification of the functional state/activity of redox-sensitive enzymes and proteins. In fact, VEGF secretion is stimulated by hypoxic conditions through HIF-1 and activator protein-1 (AP-1)⁴⁸⁰, the activity of which are both redox regulated ⁴⁸¹. Therefore, it is possible that selenium inhibits hypoxiainduced VEGF expression by modulating the redox state of thioredoxin, a critical redox mediator for HIF-1 and AP-1, via its effect on the selenoprotein, thioredoxin reductase ⁴⁸². Cell culture studies revealed that AP-1 was significantly inhibited by exposure to high levels of selenium, which binds to cysteine residues forming Se–S mixed disulfides or selenotrisulfides, thereby causing a conformational change in the protein ^{483, 484}.

At present, there are few studies dealing with Se-containing nanomaterials for biomedical applications related to angiogenesis. Aksakal and Boccaccini ⁴⁸⁵ demonstrated the feasibility of depositing a selenium coating on metallic implants using electrophoretic deposition (EPD). EPD was used in that study to produce selenium coatings with thickness in the range of tens of micrometers, although the method is potentially suitable to obtain thin nanoscale coatings. The difficulty of obtaining stable selenium powder suspensions using a mixture of sulfuric acid, ethanol, and distilled water for EPD procedures needs to be overcome.

Selenium/PLGA composite nanoparticles were incorporated in 45S5 Bioglass[®] foams by Stevanovic et al. ⁴⁸⁶ who were interested in the antibacterial properties of the scaffolds, rather than investigating the effects on angiogenesis. Furthermore, selenium was incorporated in mesoporous bioactive glasses (MBGs) to be used as a carrier for the controlled release of doxorubicin, an anticancer drug ⁴⁸⁷. However in this case, the anti-angiogenic potential of selenium was not investigated, and thus the possible multifunctional properties of Se-doped MBGs (Se-induced antiangiogenic effect + anticancer drug effect) still remain to be fully determined.

11.14 Silicon

The available literature suggests that the effects of pure silica and silicate nanomaterials on angiogenesis strongly depend on the form/embodiment in which they are used. Duan et al. ⁴⁸⁸ reported that silicon oxide (silica) nanoparticles could induce dose- and time-dependent cytotoxicity through the production of ROS and generation of oxidative stress, thereby inhibiting angiogenesis and inducing apoptosis of endothelial cells. Specifically, the presence of silica nanoparticles interferes with the formation and development of the heart in zebrafish embryos by inhibiting the activation of ERK1/2 and VEGFR-2, and by down-regulating the expression of homeoboxprotein NKX-2.5 and myocyte-specific enhancer factor 2C. The same research group studied the role of ultrafine silica particulates (size <100 nm) in inducing heart ischemia and

cardiovascular disease in mice ⁴⁸⁹. They synthesized pure-silica nanoparticles (average particle size 62 nm) by the Stober method, and observed that the cardiovascular toxicity triggered by the injected nanoparticles occurred mainly in the vascular endothelium rather than cardiomyocytes. The SiO₂ nanoparticles were able to disrupt the cell cytoskeletal organization, activate endothelial cell autophagy, cause mitochondrial damage, and partially inhibit the expression of cell-adhesion biomolecules. As a result, the endothelial cell homeostasis was disturbed and angiogenesis was eventually impaired. Cardiovascular toxicity was associated with SiO₂ nanoparticle-induced VEGFR2/PI3K/Akt/mTOR and VEGFR2/MAPK/Erk1/2/mTOR signaling pathways; crosstalk was also observed between the VEGFR2-mediated autophagy signaling pathway and the angiogenesis signaling pathway (**Fig. 20**).

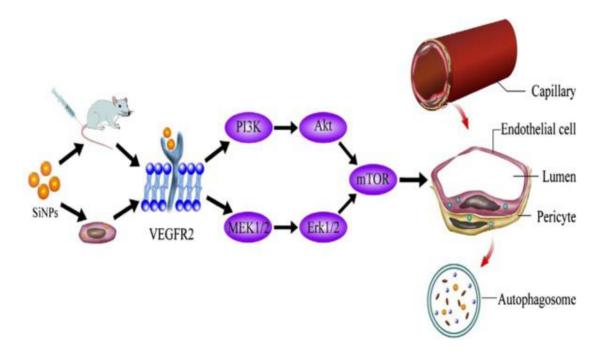


Fig. 20 Schematic model of the molecular mechanisms on VEGFR2-mediated crosstalk between autophagy and angiogenesis signaling pathways triggered by silica nanoparticles. Reproduced with permission from ref ⁴⁸⁹.

The toxicity was related to the persistence of the insoluble (or poorly soluble) pure silica nanoparticles in contact with cells and tissues, and could thus be supposed to be related to "nano-shape aspects." The situation is markedly different when SiO₂-based materials are biodegradable and thus, undergo progressive dissolution in contact with biological fluids, thereby releasing silicate ions with biological and biochemical significance. Silicate ions delivered from calcium silicate bioceramics (concentrations within 0.7-1.8 μ g/mL) were shown to play a key role in stimulating angiogenesis in co-cultures of human dermal fibroblasts (HDFs) and HUVECs ⁴⁹⁰. Specifically, calcium silicate extracts stimulated VEGF expression in HDFs, and enhanced the expression of VEGF receptor 2 in HUVECs. Angiogenesis was initiated by the activation of endothelial nitric oxide synthase and nitric oxide production in these co-cultures. The expression of vascular endothelial cadherin in co-cultured HUVECs was up-regulated and was concentrated at the cell junctions to facilitate endothelial tubule formation. A similar proangiogenic mechanism was observed in vitro by the same research group ⁴⁵⁹ when human aortic endothelial cells were cultured with dissolved extracts from akermanite, containing silicate, Ca^{2+,} and Mg²⁺ ions. Furthermore, akermanite promoted neo-vascularization after 2 and 4 months in *vivo* after being implanted in a rabbit femoral condyle model.

Silica can also be used as a network-forming oxide in the production of silicate bioactive glasses, which are highly versatile candidates for producing tissue-engineering implants. The proangiogenic potential of bioactive glasses has been comprehensively discussed by Kargozar et al. in a recent review ⁴⁷⁰.

In general, the size of the glass particles plays a role in evoking the angiogenic response, as the higher the specific surface area, the more reactive the material, and hence more ion

release. In order to address this issue, an international research team led by Boccaccini investigated the pro-angiogenic potential of poly(D, L-lactic acid) (PDLLA) scaffolds embedding 45S5 Bioglass^{*} (45SiO₂-24.5CaO-24.5Na₂O-6P₂O₅ wt.%) particles produced either by conventional melt-quenching (0.1-25 μm) or by flame synthesis (35-40 nm) ⁴⁹¹. The *in vitro* experiments using human colon (CD-18CO) fibroblasts showed that composite scaffolds containing 20 wt.% of micro- or nano-sized glass increased the expression level of VEGF up to 5 times in comparison to pure PDLLA. These results were confirmed by *in vivo* studies (rats) that revealed higher vascularization and blood vessel-to-tissue percentage in glass-filled PDLLA. The percentage of newly-formed blood vessels was 37% and 78% higher in the scaffolds with micrometric and nanometric glass particles, respectively, as compared to the controls at 8 weeks postoperative.

The same particle size-dependent trend for glasses on angiogenesis was observed at larger size scales. Detsch et al. ⁴⁹² reported that smaller particles (1-2 mm) of melt-derived S53P4 glass (53SiO₂-23Na₂O-20CaO-4P₂O₅ mol.%) stimulated a higher secretion of VEGF in human CD-18CO fibroblasts when compared to larger particles (2-3.15 mm). The pro-angiogenic potential of nano-sized sol-gel bioactive glasses 58S (58.2SiO₂-32.6CaO-9.2P₂O₅ wt.%) and 80S (80SiO₂-15CaO-5P₂O₅ wt.%) was investigated *in vitro* by Mao et al. ⁴⁹³, who reported that both materials could accelerate endothelial cell migration and up-regulate the expression of VEGF and bFGF), which resulted in enhanced tubule formation.

The dependence of the pro-angiogenic effect on glass concentration was reported by Day et al. ⁴⁹⁴, who seeded fibroblasts on 45S5 Bioglass[®]-coated polymeric implants and observed that VEGF secretion was suppressed at high glass concentrations (>0.1 wt.%). This early evidence was further confirmed by other experimental studies ⁴⁹⁵ and the concentration is now considered to

be a key factor for designing therapeutic bioactive glasses, because the same glass formulation can have different effects (angiogenic at relatively low dosage or osteogenenic at relatively high dosage) depending on the concentration of glass particles at the implantation site.

High doses of silicate material can also induce cytotoxicity due to increased concentration of ionic dissolution products, which may also increase the pH of the culture medium, producing excessive alkalinity to allow cell survival ⁴⁹⁶. This trend was also observed *in vivo*, where it was shown that the volume of blood vessels formed at the defect site around silica/collagen composite implants was inversely proportional to the biomaterial volume ⁴⁹⁷. It cannot be ignored that, when multicomponent SiO₂-based bioactive glasses are used, it is impossible to separate the biological effects of silicate ions from those of the other ions released from the material: therefore, the beneficial or adverse effects observed could be the result of a synergistic combination of the various ions. It is also worth underlining that, although silicate ions can indeed elicit a pro-angiogenic effect on their own, incorporation of metallic dopants with more potent angiogenic effects into SiO₂-based glass matrices, especially mesoporous silica nanostructures ⁴⁹⁸ may be a better strategy to promote vascular sprouting.

11.15 Silver

Silver is well known for its antimicrobial properties both in the form of free ions and nanoparticles ⁴⁹⁹. Some recent studies have also demonstrated the activity of silver nanoparticles to modulate angiogenesis; however, the picture is still incomplete, and the mechanisms involved are yet to be fully understood.

On one hand, it was reported that silver nanoparticles (average size 500 nm) biosynthesized in *Bacillus licheniformis* biomass can inhibit the proliferation and migration of bovine retinal endothelial cells supplemented with exogenous VEGF, as well as micro-vessel formation in mice ^{500, 501} (**Fig. 21**). The inhibitory effect on angiogenesis was attributed to the inactivation of the PI3K/Akt signaling pathway by the metallic nanoparticles (see **Fig. 22**). This was confirmed using biosynthesized silver nanoparticles with a smaller size (average diameter 16.5 nm) in a chick CAM model ⁵⁰². These experimental findings suggest the potential of silver nanoparticles for treating diseases where suppressing pathological angiogenesis is a goal, such as age-related macular degeneration.

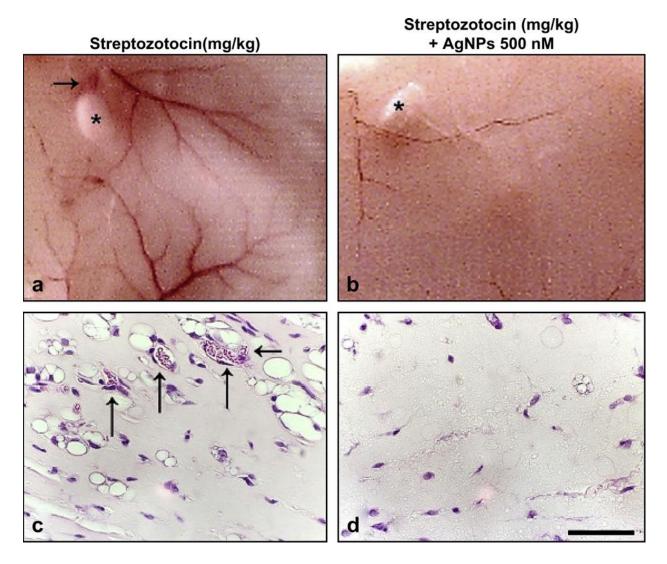


Fig. 21 Anti-angiogenic activity of Ag nanoparticles *in vivo* (rat model). Top panel: Gross photographs of Day 7 Matrigel implants with skin vessel background. Representative figures show (a) Streptozotocin without Ag nanoparticles, (b) Streptozotocin + Ag nanoparticles. Bottom panel: Histologic sections and hematoxylin and eosinstained cross-sections showing representative photographs obtained from the sections of retina stained by hematoxylin and eosin in rats (c,d). Significant differences from control group were observed (p < 0.05). Reproduced with permission from ref ⁵⁰⁰.

On the other hand, silver nanoparticles (diameter below 100 nm) produced using plant extracts from *Azadirachta indica* were suggested as potential agents for stimulating *in vivo* angiogenesis, since they could stimulate the closure of thermally-induced wounds in rats ⁵⁰³. Specifically, the wounds decreased in size over time, achieving closure at 2 weeks in healthy mice

and at 3 weeks in diabetic animals. However, the pro-angiogenic effect proposed to be elicited by silver nanoparticles was questionable in this study, as no clear evidence was provided. On the contrary, it could be suggested that wound healing was actually favored by the antimicrobial properties of Ag⁺ ions, which may have played a predominant role that overcame the antiangiogenic (i.e., anti-healing) effect of the nanoparticles.

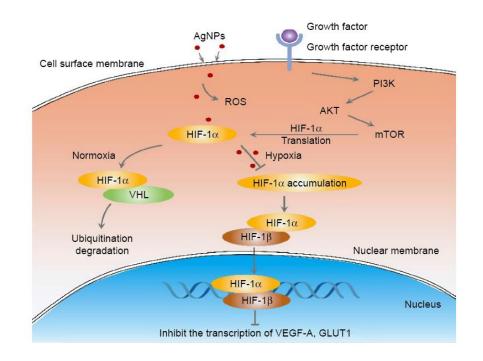


Fig. 22 Schematic representation of the effect of AgNPs on angiogenesis process in cancer cells. AgNPs could enter the cells and prevent HIF-1 α accumulation in the cytoplasm, followed by the suppression of HIF-1 target gene expression such as VEGF. from ref ⁵⁰⁴

A recent study showed that poly(vinyl pyrrolidone)-coated silver nanoparticles (average size 2.3 nm) exhibited pro-angiogenic properties both *in vitro* and *in vivo* (mouse model) ⁵⁰⁵. In fact, polymer-coated nanoparticles induced tube formation by endothelial cells, generation of ROS, and production of angiogenic factors like VEGF and nitric oxide (NO). From a biomolecular

viewpoint, the silver nanoparticles promoted the activation of FAK, Akt, ERK1/2, and p38, which are all involved in the VEGFR-mediated signaling pathway. Silver-induced angiogenesis has also been observed to occur *in vivo* around melanomas in mice. The pro-angiogenic effect observed in this study could be attributed to the presence of the surface polymer coating; a size effect could also play a role as the silver nanoparticles were significantly smaller than those used in other studies reporting an anti-angiogenic effect, but this issue remains to be elucidated.

11.16 Sulfur

Sulfur is delivered to humans via normal nutrition, for example by allium vegetables which are known to possess antioxidant, anti-inflammatory and even anticancer effects ⁵⁰⁶. S-containing compounds may be used as drugs in medical applications as they have pro- or anti-angiogenic properties depending on the type and concentration of the specific molecules. H₂S was reported to directly increase endothelial cell migration and growth as well as the formation of tubular structures *in vitro* ^{507, 508}. NaHS and Na₂S promote significant angiogenesis *in vitro* via the release of H₂S that activates the KATP channel/MAPK pathway ⁵⁰⁹, and NaHS was also found able to improve regional blood flow in mice ⁵¹⁰. Heparan sulfate proteoglycans facilitate cell signaling by acting as co-receptors for key pro-angiogenic factors like bFGF and VEGF ⁵¹¹. On the contrary, high-molecular-weight sulfonated polysaccharides show promise as anti-angiogenic agents in anti-cancer treatment as they can carry out metal chelation, or a competitive process (which is still unclear) with heparan sulfate proteoglycans, thereby inhibiting tubule formation ⁵¹².

A novel nanotechnology approach involving sulfur was recently published by Cacciotti et al. who functionalized electrospun poly(lactic acid) nanofibrous patches (average diameter of the fibers around 700 nm) with organosulphur compounds extracted from garlic. These low-cost H₂S-releasing membranes could find potential applications in various biomedical sectors, primarily as wound dressings, to combat oxidative stress in cells and improve tissue regeneration, although their specific pro-angiogenic properties still remain to be confirmed.

11.17 Terbium

Apart from europium, terbium is another element of the class of lanthanides which has recently received attention in the context of therapeutic angiogenesis. Zhao et al. reported the synthesis of $Tb(OH)_3$ nanorods and spherical nanoparticles via hydrothermal method and observed that their pro-angiogenic properties, as well as their ROS-mediated mechanism of action, are analogous to those of $Eu(OH)_3$ nanomaterials in a zebrafish model ⁴¹⁶.

Following its own previous studies on Eu(OH)₃ nanorods, Patra's group ⁵¹³ reported the synthesis of Tb(OH)₃ nanorods assisted by microwave irradiation. These nanorods exhibited proangiogenic properties in vitro towards endothelial cells (HUVECs and EA.hy926) as well as the ability to stimulate blood vessel growth in the chick CAM in vivo assay. The authors tried elucidating the biochemical mechanism behind angiogenesis and assessed that Tb(OH)₃ nanorods stimulated NO_X-mediated generation of ROS, which then activated the PI3K/Akt/MAPK signaling cascade: this resulted in the formation of intracellular NO, which is a key signaling molecule for angiogenesis ⁵¹⁴. Enhanced wound healing induced by Tb(OH)₃ nanorods was also observed in a punch biopsy mouse model.

11.18 Titanium

Titanium has been widely used for the production of load-bearing orthopedic and dental implants for decades due to its high biocompatibility and good mechanical properties ^{515, 516}. Despite may studies published on this topic, the biophysical, biochemical and biomolecular mechanisms behind the activity of titanium-based materials on angiogenesis are still to be fully understood ⁵¹⁷. In general, it was shown that titanium implants could be favorable to angiogenesis under certain circumstances (high hydrophilicity and various micro-/nanostructures), whereas titanium oxide (TiO₂, titania) nanoparticles typically exert an anti-angiogenic effect.

It is well known that the surface characteristics (e.g., ionic charge, chemistry, topography) of implantable biomaterials are key to determining the biological response of cells and tissues. The effect of the surface properties of titanium and titanium alloys in the context of bone tissue engineering has been recently reviewed by Spriano et al. ⁵¹⁸. As regards angiogenesis, it was shown that titanium surfaces with high surface energy and micro-roughness can promote the secretion of pro-angiogenic growth factors (primarily VEGF) by osteoblasts, as well as the migration and differentiation of human aortic endothelial cells cultured in contact with titanium implant extracts ⁵¹⁹. These results are consistent with other *in vitro* studies showing that hydrophilic titanium surfaces increased the absorption of plasma fibronectin ⁵²⁰, improved osteoblast differentiation and up-regulated osteoblast-related growth factors ⁵²¹. The micro-/nano-roughened surface of titanium dental screws was also found to promote osteointegration and neovascularization *in vivo* ⁵²².

The role played by hydrophilicity and surface topography is more complex regarding endothelial cells. Shi et al. ⁵²³ reported that the expression levels of some angiogenesis-related proteins (e.g., EPCR and E-selectin) were higher when osteoblast/endothelial cells were cocultured on a hydrophobic smooth titanium surface compared to a rougher one. The favorable effect of the smooth surface was explained by the tendency of endothelial cells to spread and attach onto smooth biological surfaces typical of blood vessels. However, these findings were not consistent with the results reported by Au et al. ⁵²⁴ using only HUVECs, which revealed a higher expression of pro-angiogenic genes when cultured on hydrophilic rough titanium surfaces. This apparent inconsistency can be explained taking into account that the cell behavior might be affected by cell-cell interactions and cross-talk in the co-culture experiment ⁵²³. In fact, many other authors have observed an increase of VEGF secretion by endothelial progenitor cells in contact with micro-rough titanium surfaces, which were also capable of accelerating vascularization in human patients ⁵²⁵⁻⁵²⁷. Endothelial progenitor cells can affect neovascularization by secreting paracrine factors (e.g. cytokines and VEGF) and forming a primary cell network after differentiating into endothelial cells or perivascular supporting cells. Ziebart et al. ⁵²⁷ showed that rough titanium surfaces promoted an undifferentiated rounded phenotype with low proliferation rate and lower endothelial nitric oxide (NO) synthase and/or inducible NO synthase activities; however, these cells showed a high rate of VEGF secretion, thus eventually promoting angiogenesis.

Comparison between different studies is difficult due to the differing experimental conditions used, cell types and titanium topographies, which prevent firm conclusions from being drawn at this stage. Functionalization strategies have also been recently carried out on

titanium implants to impart them with clear pro-angiogenic properties. For example, Chen et al. ⁵²⁸ modified the surface of titanium substrates by depositing a composite coating (nanofibers of chitosan-catechol, gelatin, and hydroxyapatite) that improved angiogenesis *in vitro* and *in vivo*; of course, this effect cannot solely be attributed to titanium in itself.

Doping titanium-based surfaces with other metals having a potent pro-angiogenic (e.g., copper) is another interesting approach recently reported by Zong et al. ⁵²⁹, who applied an anodization treatment through magnetron sputtering to TiCu layers previously deposited on pure titanium foils. The resulting Cu-doped titania-based nanotubular surfaces were capable of up-regulating VEGF secretion by endothelial cells as compared to Cu-free titania nanotubes due to the release of Cu²⁺ ions. On the contrary, pure titania nanoparticles were found to exert an anti-angiogenic effect apparently associated with their specific ability to inhibit the angiogenic processes, rather than to a "general" nanosize-dependent cytotoxicity. Interestingly, Jo et al. ⁵³⁰ observed that titania nanoparticles inhibited VEGF-induced tube formation and migration of human retinal microvascular endothelial cells via the suppression of VEGFR2 and MAPK. This property potentially suggests therapeutic applications in which the suppression of angiogenesis is a goal, including age-related macular disease and tumor treatment.

A very recent study by Augustine et al. ⁵³¹ showed that, interestingly, titania nanorods produced via hydrothermal treatment exhibit a pro-angiogenic effect. This finding was assessed both in a CAM model and in rats, where electrospun PCL meshes loaded with titania nanorods provided a faster wound healing compared to bare PCL fibres.

Hence, the comparison between the above-mentioned studies suggests a "shape effect" related to nano-titania (pro-angiogenic nanorods vs. anti-angiogenic spherical nanoparticles), which deserves to be more comprehensively investigated in the future.

11.19 Yttrium

Yttrium oxide (Y₂O₃) has been used for decades to stabilize the tetragonal phase of zirconia in ceramic components for joint prostheses ⁵³². More recently, Y₂O₃ nanoparticles have been proved to have antioxidant and radical scavenging ability ⁵³³, which are of great interest in advanced tissue engineering applications. Investigation of the potential of Y₂O₃ nanoparticles in the specific context of angiogenesis is in its very beginning, but the existing evidence shows promise. Following the approach reported in their previous studies on nanoceria, Augustine et al. ⁵³⁴ produced Y₂O₃ nanoparticles using gelatin as a stabilizer and incorporated them in electrospun PCL scaffolds. An amount of 1 wt.% Y₂O₃ nanoparticles was found to be the most effective to promote the proliferation of fibroblasts (L-929) and osteoblast-like cells (UMR-106), as well as to support the highest blood vessel formation in a chick CAM model. Gene expression study following subcutaneous implantation in rats demonstrated that the presence of Y₂O₃ nanoparticles in the polymeric scaffolds could upregulate the expression of cell proliferation and angiogenesis-related biomolecules, such as VEGF and EGFR.

11.20 Zinc

Zinc is an essential element in human metabolism because Zn²⁺ ions are included in a number of proteins and are involved in many biological processes ⁵³⁵. Zinc can have anti- or pro-

angiogenic effects depending on the form under which it is available, i.e., Zn^{2+} cations or zinc oxide (ZnO) nanoparticles, respectively. It was observed that the ability of Zn^{2+} ions to bind to endostatin is essential for the potent anti-angiogenic activity of this angiogenesis inhibitor that can convert malignant cancer cells into a "quiescent" tumor phenotype unable to induce angiogenesis, and thus unable to grow ⁵³⁶. The effect of zinc on cancer cells was also related to a conformational change in the p53 protein, resulting in halting the progression of cancer cell mitosis and promoting cell death. As regards the effect of Zn^{2+} ions on healthy cells, it was observed that zinc concentrations in the range of 10 to 500 μ M neither inhibited nor stimulated the growth of HUVECs ⁵³⁵; however, zinc could enhance the proliferation of bovine aortic endothelial cells in the presence of exogenous bFGF, suggesting a role of zinc ions in amplifying the bFGF-dependent proliferation of the cells ⁵³⁷.

An *in vitro* study published by Shiah et al. ⁵³⁸ reported that the drug bis(diethylthiocarbamoyl) disulfide (disulfiram), which is used for treating alcoholism, could directly interact with MMP-2 and MMP-9, inhibiting their proteolytic activity through a zinc-chelating mechanism. As a result, reduction of angiogenesis was predicted using disulfuram *in vivo*, with therapeutic benefits in cancer treatment. Zinc is also capable of reversing the expression of many genes modulated by hypoxia, thus reducing the activity of HIF-1 with an associated decrease in VEGF secretion and reduction of angiogenesis ^{539, 540}.

On the contrary, pro-angiogenic effects were observed *in vitro* and *in vivo* when zinc was used in the form of ZnO nano-sized structures like nanoparticles ⁵⁴¹ and nanoflowers ⁵⁴² (**Fig. 23**), alone or embedded in a polymeric scaffold (e.g. electrospun poly(vinylidene fluoride-trifluoroethylene)

fibres ⁵⁴³). In both cases, generation of ROS by the ZnO nanomaterials resulted in upregulation of bFGF and VEGF, which ultimately led to improved vascularization.

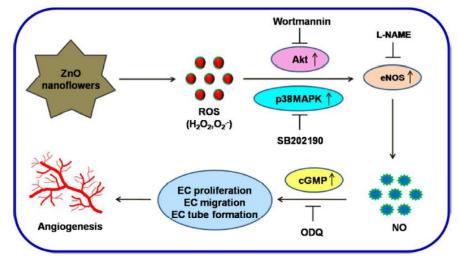


Fig. 23 The possible molecular mechanisms and signaling pathways involved in zinc nanoflower-induced angiogenesis. Reproduced with permission from ref. ⁵⁴².

 Table. 4 Pro-angiogenic biochemical and biological functions elicited by inorganic elements and nanomaterials (the elements are listed in alphabetical order).

Element	Effect*	Notes/Biochemical and biological functions	Ref (s)
В	PA	Borate ((BO ₃) ³⁻) ions can induce: - Stimulation of endothelial cell migration and proliferation - Increased secretion of VEGF and other pro-angiogenic factors - Tubule formation	336, 401, 544
Са	PA	Ca ²⁺ ions induce: - Endothelial cell proliferation - Overexpression of PDGF, EGF, IGF-I, bFGF, VEGF	342, 343, 346
Ce	PA	 Nanoceria could stabilize HIF-1α in ECs and up-regulate VEGF expression, resulting in induced pro-angiogenesis High surface area and increased Ce^{3+/}Ce⁴⁺ ratio make nanoceria a robust inducer of angiogenesis 	353
Со	РА	Co ²⁺ ions induce: - Activation of the HIF-1 pathway - Overexpression of angiogenic factors VEGF and bFGF - Enhanced tubule formation	365, 366
Cu	ΡΑ	Cu ⁺ /Cu ²⁺ ions and copper nanoparticles induce: - Activation of the HIF-1 pathway - MAPK signaling pathway - Activation of VEGF, bFGF, TNF, IL-1β, IL-6, and IL-8 - Endothelial cell proliferation	390, 393, 394, 407

Eu	ΡΑ	Eu ³⁺ ions induce: - Overexpression of angiogenic genes CD31, MMP9, VEGFR1/2 and PDGFR α/β of HUVECs - Promotion of endothelial cell proliferation <i>in vitro</i> and vascular sprouting <i>in vivo</i> (CAM model and mice) Eu(OH) ₃ nanorods/nanoparticles promote angiogenesis mediated by ROS production (especially H ₂ O ₂).	417, 419
Fe	PA	Fe ²⁺ /Fe ³⁺ ions induce: - ROS generation - Stabilization of HIF - VEGF increase	435, 436, 445
Li	PA	Li ⁺ ions promote: - VEGF secretion - Vasculogenesis	448, 451, 452
Mg	PA	Mg ²⁺ ions released from silicate bioceramics and glasses induce: - Stimulation of proliferation and migration of microvascular cells - Enhancement of the mitogenic response to angiogenic factors	458
Nb	PA	Nb ⁵⁺ ions released from bioactive glasses promote angiogenesis <i>in vitro</i> through enhancing VEGF secretion	462
Ρ	РА	Phosphate ((PO ₄) ²⁻) ions induce: - Stimulation of pro-angiogenic bFGF, VEGF, FOXC2, and osteopontin - Stimulation of migration and tube formation in the HUVEC model	349, 468
S	PA	Different sulphur compounds can exert pro-angiogenic (H ₂ S, NaHS, Na ₂ S)	508, 509
Si	PA	- Pro-angiogenic effect elicited by silicate ((SiO ₄) ⁴⁻) ions (induction of endothelial cell homing, polarization and migration; induction of angiogenic differentiation and new blood vessel sprouting)	459, 490
Tb	PA	Tb(OH) ₃ nanorods stimulate NO _x -mediated generation of ROS, with activation of the PI3K/Akt/MAPK signaling cascade and formation of intracellular NO, which is a key signaling molecule for angiogenesis	513
Ti	ΡΑ	- Pro-angiogenic effect elicited by hydrophilic and relatively smooth titanium surfaces	519-524
Y	PA	$Y_2 O_3$ nanoparticles stimulate VEGF and EGFR secretion	534
Zn	PA	- Pro-angiogenic effect elicited by ZnO nanoparticles through ROS generation and upregulation of bFGF and VEGF	537

 Table. 5 Anti-angiogenic biochemical and biological functions elicited by inorganic elements and nanomaterials (the elements are listed in alphabetical order).

Element	Effect*	Notes/Biochemical and biological functions	Ref (s)
Ag	AA	Silver nanoparticles act on the PI3K/Akt signaling pathway	310
Au	AA	Gold nanoparticles induce: - Inhibition of the MAPK pathway - Inhibition of pro-angiogenic factors (e.g., VEGF, bFGF, PIGF)	421, 427
Ce	АА	The antiangiogenic effect was found to occur at high concentrations and in the presence of rod-shaped nanoceria.	361, 545
S	АА	Sulphur compounds such as heparan sulfonate can exert antiangiogenic effects	512
Se	АА	 Inhibition of VEGF secretion Apoptosis of endothelial cells 	476, 477, 479
Si	АА	- Anti-angiogenic effect elicited by pure silica nanoparticles or at a high dosage of silicate materials (cytotoxicity)	530
ті	АА	- Anti-angiogenic effect elicited by titania nanoparticles via suppression of VEGF/MAPK pathways	546
Zn	АА	- Anti-angiogenic effect elicited by Zn ²⁺ ions (e.g., activation of endostatin, reverse effect on hypoxia-modulated genes)	538, 539

12. Nanoparticles for imaging of angiogenesis

Visualization of tumor angiogenesis provides invaluable information to assess the biologic aggressiveness and allow monitoring of tumor response to anti-angiogenic therapies. Scientists have taken advantage of nanotechnology for imaging of tumor angiogenesis since nanoparticles show several advantages, such as the ability to carry high payloads of diagnostic or imaging agents, with an increased signal-to-noise ratio, longer circulation times, and enhanced image contrast ⁶¹. Successful imaging of angiogenesis leads to gaining valuable information, which can

be useful in determining the optimal dose and schedule of an anti-angiogenic therapeutic, as well as measuring early signs of tumor relapse/recurrence⁵⁴⁷. Magnetic (e.g. Gd), fluorescent and radiolabeled nanoparticles (e.g., Tc-99m and I-123) have commonly been used to detect the angiogenesis process. Positron-emission tomography (PET), single-photon emission computed tomography (SPECT), computerized tomography (CT), magnetic resonance imaging (MRI), optical imaging, and ultrasounds imaging have all been investigated as techniques to measure the progress of angiogenesis in vivo ⁵⁴⁸⁻⁵⁵³. Targeting $\alpha_{v}\beta_{3}$ –integrin and VEGFR2 has been proposed for targeted imaging of tumor angiogenesis ⁵⁵⁴⁻⁵⁵⁷. The important parameters measured are microvessel density (the so-called "hot-spots") and circulating markers of angiogenesis ⁵⁴⁷. The latter are comprised of soluble circulating protein markers such as angiogenic GFs and their receptors (e.g., VEGF), cell adhesion and ECM molecules (VCAM-1), circulating EPCs and their precursors ⁵⁴⁷. However, there are other biomarkers that are over-expressed on the tumor cell surface, which could be potential targets for detection of tumor angiogenesis. As an illustration, Wu et al. functionalized gold nanoparticles with a tumor-homing cyclized asparagine-glycinearginine peptide (SH-cNGR) and carboxylpoly(ethylene glycol)thiol (SH-PEG-COOH) via Au-S bonds to target the aminopeptidase-N (APN/CD13) over-expressed on the endothelium of tumor angiogenesis ⁵⁵⁸. CT imaging and immunohistochemistry results showed that the surfacefunctionalized nanoparticles showed significantly higher and faster tumor uptake postintravenous injection in comparison to unmodified samples. The authors suggested that this nano-based imaging system could be a promising contrast agent for targeted angiogenesis imaging using CT.

Organic based nanoparticles have also been used to detect angiogenesis *in vivo*. Ryu et al. developed a HSA-based nanoprobe for non-invasive optical imaging of MMP activity in a small rodent hindlimb ischemia model ⁵⁵⁹. For this aim, the authors covalently conjugated MMPspecific fluorogenic peptide probes to HSA for preparation of self-quenched MMP eHSA nanoprobes 36 nm in diameter. It has been proven that MMP-2 and MMP-9 are two essential mediators in the angiogenesis process. The nanoprobes showed enhanced fluorescence emission in the presence of MMP-2 and MMP-9 without any cytotoxicity. Moreover, the authors showed longer blood circulation half-life for this system compared to control groups after intravenous injection in a mouse hindlimb ischemia model, as well as successful optical imaging of MMP activity during angiogenesis. It should be mentioned that the use of viral nanoparticles (e.g., cowpea mosaic virus nanoparticles) as next-generation imaging agents may be a new approach to non-invasive monitoring of angiogenesis, and could serve as suitable therapy and imaging probes ⁵⁶⁰⁻⁵⁶².

Multimodality (e.g., bimodal, trimodal, and four-modal) imaging has emerged as an approach for imaging of the angiogenesis process due to its potential to provide complementary information that can be co-registered ⁵⁶³⁻⁵⁶⁵. For instance, lanthanide-based nanoprobes have been used for imaging of tumor angiogenesis thanks to their favorable optical, magnetic, radioactive, and X-ray attenuation properties. In 2013, Sun et al. optimized core-shell lanthanide upconversion nanophosphors with an enhanced imaging ability for tumor angiogenesis in small animals (mice) ⁵⁶⁶. NaLuF₄: Yb,Tm nanocrystals and ¹⁵³Sm³⁺ doped NaGdF₄ were the core and the shell of the nanophosphors, respectively. The recorded lifetime for upconversion luminescence (UCL) was 1044 μ s at 800 nm, and its relaxation rate (1/T₁) was 1044 μ s and 18.15 s⁻¹ .mM⁻¹. The

detailed information obtained by four-modal imaging techniques, i.e., X-ray, CT, MRI, and SPECT (see **Fig. 24**), showed the effectiveness and applicability of this system for monitoring tumor angiogenesis *in vivo*.

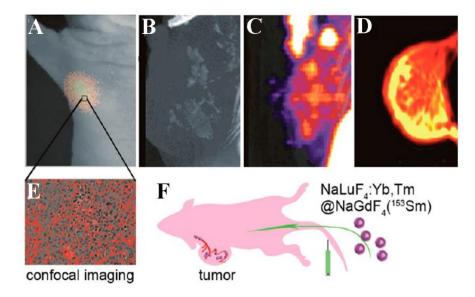


Fig. 24 The use of NaLuF4: Yb,Tm@NaGdF4(153Sm) for four-modal imaging of tumor-bearing nude mice at 60 min post intravenous injection. A-D represent images obtained from upconversion luminescence (UCL), X-ray CT, SPECT, and MR of tumor, respectively. E exhibits UCL confocal image of the paraffin section of tumor tissue, and F is actually a schematic illustration of tumor angiogenesis imaging by applying the nanoparticles as the probe. ⁵⁶⁶.

Carbon-based nanoparticles (e.g., nanographene oxide and quantum dots) are other promising candidates for targeted angiogenesis imaging *in vivo* ^{567, 568}. The use of quantum dots for *in vivo* imaging purposes was first reported by Dubertret and colleagues in 2002 ⁵⁶⁹. Today, paramagnetic QD-micelles have been used for MR and optical-based molecular imaging *in vivo* ⁵⁷⁰⁻⁵⁷². QDs in pristine and modified forms were suggested to be tools in multimodal molecular imaging of tumor angiogenesis ^{573, 574}.

13. Summary and future perspectives

Because angiogenesis is indispensible for tumor growth (beyond a certain relatively small size) and also for cancer metastasis which is the leading cause of cancer death, therefore, inhibiting angiogenesis has been an important part of cancer therapy in the clinical setting for some decades. VEGF signaling pathway plays a central role in both physiological and pathological processes of angiogenesis; therefore, its regulation has been considered to be one of the main targets in cancer therapy ⁵⁷⁵. Recent studies have revealed the critical role of angiogenesis in the failure of initially successful immunotherapy approached to solid tumors. Therefore combining antiangiogenic therapy with modern immunotherapy has been proposed to improve the effectiveness of immunotherapy and also to diminish the risk of autoimmune-related adverse effects .

Many cancers could be resistant to anti-angiogenic approaches targeting the VEGF signaling pathway either inherently or by acquired resistance. In the case of acquired resistance, cancer cells upregulate different pro-angiogenic molecules which might be connected with the genetic instability of cancers ⁵⁷⁶. Also, this resistance could be related to the ability of cancer cells to receive nutrients from existing adjacent blood vessels ⁵⁷⁷. The invention of nanotechnology has opened up new horizons in many areas of biomedical science, and inhibiting angiogenesis using different types of nanoparticles is now a promising approach especially in the case of resistant cancers. Up to now, a large number of studies have been published dealing with nanoparticles that inhibit angiogenesis. Better outcomes may be obtained when organic and inorganic nanoparticles are combined. There has been a series of anti-angiogenic nano-sized drugs (e.g., humanized anti-VEGF monoclonal antibody AvastinTM) gaining marketing approval to

treat various types of cancer ^{21, 32}. In addition, it should be mentioned that many attempts are being made to develop nano-sized particles incorporating previously used anti-angiogenic commercial drugs, as well as substances originating from herbs and phytochemicals. The high efficacy of Taxol[®] (generic name paclitaxel) nanoparticles has been reported in several studies ^{209, 578}. The use of nano-based DDSs (e.g., lipid-based and carbon-based nanosystems) provide another possibility for researchers to take advantage of nanotechnology to inhibit angiogenesis in cancers both *in vitro* and *in vivo*. Doxil[®], MyocetTM, Lipo-dox[®], DaunoXome[®], and Marqibo[®] are well-known examples of FDA-approved nanosystems containing drugs with additional antiangiogenic properties, which are currently used in cancer therapy. Targeted therapy of angiogenesis by surface-modified nanoparticles and nanosystems is currently under investigation to improve the efficacy of anti-angiogenic based cancer therapy. The anti-angiogenic effect of nanoparticles depends on many factors, including their size and shape ⁵⁷⁹. Although there are several reported in vitro and in vivo experiments concerning the size effects of nanoparticles on angiogenesis, some conflicting results make it difficult to draw firm conclusions about this parameter ^{546, 580, 581}. However, the use of smaller particles is generally suggested to improve the results of anti-angiogenesis based cancer therapy. It is worth underlining that metallic and metal oxide nanoparticles can exhibit either pro-angiogenic or anti-angiogenic properties depending on various parameters, including not only the size but also the surface properties (e.g. wettability, charge) and effective concentration. All these factors were shown to be key determinants for the production of ROS, which have a deep impact on the angiogenic properties of nanomaterials ⁵⁸². In the last decade, redox signalling-based nanomedicine has indeed emerged as a fascinating approach for the treatment of angiogenesis-related diseases, where nano-sized materials may

promote angiogenesis via the controlled production of ROS or antiangiogenesis by triggering excessive ROS formation ⁵⁸³⁻⁵⁸⁵. A generally-valid "set of rules" on how ROS production can be actually controlled does not exist, also considering the high number of factors involved (e.g. type of nanomaterial, shape, size, concentration, specific environment/model etc.) and their interlocking – which may even be unpredictable: hence, the need for detailed and individual studies on every material/system embedding nanoparticles is recommended before exploitation for therapeutic purpose.

Imaging and monitoring of angiogenesis by nanotechnology-based probes may lead to faster and more accurate diagnosis of cancer progression; while multimodality imaging of angiogenesis for instance using lanthanide-based nanoprobes has shown good efficacy.

The issue of toxicity of nanoparticle-based molecules, chemicals, and drugs requires to be carefully evaluated; some nanoparticulate elements with the ability to modulate angiogenesis (e.g., arsenic, lead, and mercury) have severe toxicity to mammalian cells and are never used in therapeutic applications. Moreover, the selection of solvents and reagents used for preparation of nanoparticles is of great importance when aiming to use them in the human body. Green chemistry offers better approaches to reduce toxicity and also to improve the stability of nanomaterials ^{586, 587}.

On the other side, improving angiogenesis is critical in wound healing, tissue engineering and reconstructive strategies, since it can facilitate the growth and repair of damaged tissues and organs ⁴⁷⁰. A large number of studies have investigated the pro-angiogenic potential of various organic and inorganic nanomaterials both *in vitro* and *in vivo*. The results obtained so far can be regarded as quite promising, and much attention has been given to this direction. The critical issues for the use of nanotechnology in promoting angiogenic strategies are similar to the above-mentioned factors affecting anti-angiogenic strategies.

One interesting theme that has emerged in serveral of the approaches covered in this review, is the bimodal effects of many nanostructures, that can stimulate angiogenesis at low doses or concentrations, while the same material can inhibit angiogenesis at higher doses or concentrations. This may even allow the same preparation to be used for opposite goals at different doses depending on the disease or condition to be treated.

Taken together, nanotechnology has had and will continue to have a major impact on the therapeutic tools and imaging approaches targeting the neovascularization process in both proangiogenic and anti-angiogenic strategies. In addition, the quality of recorded images of angiogenesis process is really improved by applying nanotechnology-based methods. Based on the current knowledge of nanotechnology, it can be assumed that novel chemical formulations will be invented and developed into different formats (e.g., small molecules) affecting angiogenesis in a more effective manner, even in the case of cancer resistances.

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References

- 1. R. K. Jain, *Nature medicine*, 2003, **9**, 685.
- 2. E. M. Conway, D. Collen and P. Carmeliet, *Cardiovascular research*, 2001, **49**, 507-521.
- 3. P. Carmeliet and R. K. Jain, *Nature*, 2000, **407**, 249-257.
- 4. R. H. Adams and K. Alitalo, *Nature reviews Molecular cell biology*, 2007, **8**, 464.
- 5. L. Schito and G. L. Semenza, *Trends in cancer*, 2016, **2**, 758-770.
- R. Auerbach, R. Lewis, B. Shinners, L. Kubai and N. Akhtar, *Clinical chemistry*, 2003, 49, 32-40.
- 7. M. W. Irvin, A. Zijlstra, J. P. Wikswo and A. Pozzi, *Experimental Biology and Medicine*, 2014, **239**, 1476-1488.
- 8. A. A. Ucuzian, A. A. Gassman, A. T. East and H. P. Greisler, *Journal of Burn Care & Research*, 2010, **31**, 158-175.
- 9. M. Yunus, P. J. Jansson, Z. Kovacevic, D. S. Kalinowski and D. R. Richardson, *Biochimica et Biophysica Acta (BBA)-General Subjects*, 2019.
- 10. J. Rajasekar, M. K. Perumal and B. Vallikannan, *The Journal of Nutritional Biochemistry*, 2019, **71**, 1-15.
- 11. A. Fallah, A. Sadeghinia, H. Kahroba, A. Samadi, H. R. Heidari, B. Bradaran, S. Zeinali and O. Molavi, *Biomedicine & Pharmacotherapy*, 2019, **110**, 775-785.
- 12. A. E. El-Kenawi and A. B. El-Remessy, *British journal of pharmacology*, 2013, **170**, 712-729.
- 13. D. Ribatti, *Leukemia research*, 2009, **33**, 638-644.
- 14. H. P. Eikesdal, H. Sugimoto, G. Birrane, Y. Maeshima, V. G. Cooke, M. Kieran and R. Kalluri, *Proceedings of the National Academy of Sciences*, 2008, **105**, 15040-15045.
- 15. M. Ueda and H. Saji, *The Scientific World Journal*, 2014, **2014**.
- 16. R. K. Jain, *Nature medicine*, 2001, **7**, 987.
- 17. J. Folkman, New england journal of medicine, 1971, 285, 1182-1186.
- 18. R. Ronca, M. Benkheil, S. Mitola, S. Struyf and S. Liekens, *Medicinal research reviews*, 2017, **37**, 1231-1274.
- 19. G. C. Jayson, R. Kerbel, L. M. Ellis and A. L. Harris, *The Lancet*, 2016, **388**, 518-529.

- 20. L. M. Ellis and D. J. Hicklin, *Nature reviews cancer*, 2008, **8**, 579.
- 21. C. Aghajanian, S. V. Blank, B. A. Goff, P. L. Judson, M. G. Teneriello, A. Husain, M. A. Sovak, J. Yi and L. R. Nycum, *Journal of clinical oncology*, 2012, **30**, 2039.
- E. Pujade-Lauraine, F. Hilpert, B. Weber, A. Reuss, A. Poveda, G. Kristensen, R. Sorio, I. Vergote, P. Witteveen and A. Bamias, *Obstetrical & Gynecological Survey*, 2014, 69, 402-404.
- 23. G. Bergers and D. Hanahan, *Nature Reviews Cancer*, 2008, **8**, 592.
- G. Jiménez-Valerio, M. Martínez-Lozano, N. Bassani, A. Vidal, M. Ochoa-de-Olza, C. Suárez, X. García-del-Muro, J. Carles, F. Viñals and M. Graupera, *Cell reports*, 2016, 15, 1134-1143.
- 25. T. Cascone, L. Xu, H. Y. Lin, W. Liu, H. T. Tran, Y. Liu, K. Howells, V. Haddad, E. Hanrahan and M. B. Nilsson, *Clinical Cancer Research*, 2017, **23**, 5489-5501.
- 26. M. Pàez-Ribes, E. Allen, J. Hudock, T. Takeda, H. Okuyama, F. Viñals, M. Inoue, G. Bergers, D. Hanahan and O. Casanovas, *Cancer cell*, 2009, **15**, 220-231.
- K. J. Gotink, H. J. Broxterman, M. Labots, R. R. De Haas, H. Dekker, R. J. Honeywell, M. A. Rudek, L. V. Beerepoot, R. J. Musters and G. Jansen, *Clinical Cancer Research*, 2011, **17**, 7337-7346.
- 28. S. A. Patel and A. J. Minn, *Immunity*, 2018, **48**, 417-433.
- 29. J. Nam, S. Son, L. J. Ochyl, R. Kuai, A. Schwendeman and J. J. Moon, *Nature communications*, 2018, **9**, 1074.
- 30. D.-H. Kong, M. Kim, J. Jang, H.-J. Na and S. J. I. j. o. m. s. Lee, 2017, 18, 1786.
- 31. M. Rajabi and S. J. B. Mousa, 2017, 5, 34.
- 32. N. Ferrara, K. J. Hillan, H.-P. Gerber and W. J. N. r. D. d. Novotny, 2004, **3**, 391.
- 33. M. I. Braghiroli, J. Sabbaga and P. M. J. E. r. o. a. t. Hoff, 2012, **12**, 567-580.
- 34. J. L. Spratlin, K. E. Mulder and J. R. J. F. O. Mackey, 2010, 6, 1085-1094.
- 35. R. M. Poole and A. J. D. Vaidya, 2014, **74**, 1047-1058.
- 36. K. K. Ciombor, J. Berlin and E. J. C. C. R. Chan, 2013, **19**, 1920-1925.
- 37. A. Gaya and V. J. C. t. r. Tse, 2012, **38**, 484-493.
- 38. M. J. D. Shirley, 2017, **77**, 107-112.
- 39. B. J. Andrick and A. J. A. o. P. Gandhi, 2017, **51**, 1090-1098.

- 40. Y. Chen, M. A. Tortorici, M. Garrett, B. Hee, K. J. Klamerus and Y. K. J. C. p. Pithavala, 2013, **52**, 713-725.
- 41. C. Grüllich, in *Small Molecules in Oncology*, Springer, 2014, pp. 207-214.
- 42. S. Zschäbitz and C. Grüllich, in *Small Molecules in Oncology*, Springer, 2018, pp. 187-198.
- 43. S. Wind, U. Schmid, M. Freiwald, K. Marzin, R. Lotz, T. Ebner, P. Stopfer and C. J. C. p. Dallinger, 2019, 1-17.
- 44. R. B. Verheijen, J. H. Beijnen, J. H. Schellens, A. D. Huitema and N. J. C. p. Steeghs, 2017, 56, 987-997.
- 45. C. L. Shamroe and J. M. J. A. o. P. Comeau, 2013, 47, 1540-1546.
- 46. S. M. J. D. Hoy, 2014, **74**, 793-806.
- 47. L. Rimassa, T. Pressiani, N. Personeni and A. J. E. r. o. a. t. Santoro, 2017, **17**, 567-576.
- 48. B. I. J. E. o. o. p. Rini, 2006, **7**, 453-461.
- 49. M. J. N. R. C. O. Debiec-Rychter, 2007, **4**, 342.
- 50. P. Fallahi, S. M. Ferrari, E. Baldini, M. Biricotti, S. Ulisse, G. Materazzi, P. Miccoli and A. J. E. r. o. a. t. Antonelli, 2016, **16**, 1109-1118.
- 51. Y. X. Zhu, K. M. Kortuem, A. K. J. L. Stewart and lymphoma, 2013, **54**, 683-687.
- 52. L. Lu, F. Payvandi, L. Wu, L.-H. Zhang, R. J. Hariri, H.-W. Man, R. S. Chen, G. W. Muller, C. C. Hughes and D. I. J. M. r. Stirling, 2009, **77**, 78-86.
- 53. J.-J. Patard, D. Pouessel, K. Bensalah and S. J. W. j. o. u. Culine, 2008, **26**, 135-140.
- 54. D. A. Krueger, M. M. Care, K. Holland, K. Agricola, C. Tudor, P. Mangeshkar, K. A. Wilson, A. Byars, T. Sahmoud and D. N. J. N. E. J. o. M. Franz, 2010, **363**, 1801-1811.
- 55. Y. Yina and D. Talapin, *Chemical Society Reviews*, 2013, **42**, 2484-2487.
- 56. F. Farjadian, A. Ghasemi, O. Gohari, A. Roointan, M. Karimi and M. R. Hamblin, *Nanomedicine*, 2019, **14**, 93-126.
- 57. S. Kargozar, S. Ramakrishna and M. Mozafari, *Current Opinion in Biomedical Engineering*, 2019, **10**, 181-190.
- 58. M. Shi, L. Xia, Z. Chen, F. Lv, H. Zhu, F. Wei, S. Han, J. Chang, Y. Xiao and C. Wu, *Biomaterials*, 2017, **144**, 176-187.
- 59. M. Zhang and L. Jiang, *Journal of biomedical nanotechnology*, 2016, **12**, 1975-1986.

- 60. S. Kargozar, M. Mozafari, S. Hamzehlou, H.-W. Kim and F. Baino, *Materials Letters*, 2019, **251**, 241-246.
- 61. D. Banerjee, R. Harfouche and S. Sengupta, *Vascular cell*, 2011, **3**, 3.
- 62. J. L. Paris, G. Villaverde, S. Gómez-Graña and M. Vallet-Regí, *Acta biomaterialia*, 2020, **101**, 459-468.
- 63. F. Danhier, A. Le Breton and V. r. Préat, *Molecular pharmaceutics*, 2012, 9, 2961-2973.
- 64. N. Graf, D. R. Bielenberg, N. Kolishetti, C. Muus, J. Banyard, O. C. Farokhzad and S. J. Lippard, *ACS nano*, 2012, **6**, 4530-4539.
- 65. P. Majumder, *Bioengineering*, 2018, 5, 76.
- 66. C. Rüegg and G. C. Alghisi, in *Angiogenesis Inhibition*, Springer, 2010, pp. 83-101.
- 67. A. Clavreul, M. Pourbaghi-Masouleh, E. Roger and P. Menei, *International journal of nanomedicine*, 2019, **14**, 2497.
- 68. G. Taneja, A. Sud, N. Pendse, B. Panigrahi, A. Kumar and A. K. Sharma, *Cardiovascular toxicology*, 2019, **19**, 1-12.
- 69. P. Sanchez-Moreno, J. L. Ortega-Vinuesa, J. M. Peula-Garcia, J. A. Marchal and H. Boulaiz, *Current drug targets*, 2018, **19**, 339-359.
- 70. B. Bahrami, M. Hojjat-Farsangi, H. Mohammadi, E. Anvari, G. Ghalamfarsa, M. Yousefi and F. Jadidi-Niaragh, *Immunology Letters*, 2017, **190**, 64-83.
- G. E. Vargas, L. A. H. Durand, V. Cadena, M. Romero, R. V. Mesones, M. Mačković, S. Spallek, E. Spiecker, A. R. Boccaccini and A. A. Gorustovich, *Journal of Materials Science: Materials in Medicine*, 2013, 24, 1261-1269.
- 72. A. M. Abdalla, L. Xiao, M. W. Ullah, M. Yu, C. Ouyang and G. Yang, *Theranostics*, 2018, **8**, 533.
- 73. I. Ali, K. Salim, M. A Rather, W. A Wani and A. Haque, *Current cancer drug targets*, 2011, **11**, 135-146.
- 74. R. Gaspar and R. Duncan, *Advanced drug delivery reviews*, 2009, **61**, 1220-1231.
- 75. T. Yadavalli, S. Ramasamy, G. Chandrasekaran, I. Michael, H. A. Therese and R. Chennakesavulu, *Journal of magnetism and Magnetic Materials*, 2015, **380**, 315-320.
- 76. G. A. Hughes, in *Nanomedicine in Cancer*, Pan Stanford, 2017, pp. 47-72.
- 77. Y. Li, T. Thambi and D. S. Lee, *Advanced healthcare materials*, 2018, 7, 1700886.

- 78. J. V. Jokerst, T. Lobovkina, R. N. Zare and S. S. Gambhir, *Nanomedicine*, 2011, **6**, 715-728.
- 79. M. Qiu, C. Wang, D. Chen, C. Shen, H. Zhao and Y. He, *Applied Sciences*, 2016, 6, 290.
- 80. E. B. Peters, N. Christoforou, K. W. Leong, G. A. Truskey and J. L. West, *Cellular and molecular bioengineering*, 2016, **9**, 38-54.
- 81. B. K. Lee, Y. Yun and K. Park, Advanced drug delivery reviews, 2016, 107, 176-191.
- 82. Q. Hu, X. Gao, G. Gu, T. Kang, Y. Tu, Z. Liu, Q. Song, L. Yao, Z. Pang and X. Jiang, *Biomaterials*, 2013, **34**, 5640-5650.
- 83. H. M. Burt, J. K. Jackson, S. K. Bains, R. T. Liggins, A. M. C. Oktaba, A. L. Arsenault and W. L. Hunter, *Cancer letters*, 1995, **88**, 73-79.
- 84. G. Gu, Q. Hu, X. Feng, X. Gao, J. Menglin, T. Kang, D. Jiang, Q. Song, H. Chen and J. Chen, *Biomaterials*, 2014, **35**, 8215-8226.
- 85. S. François, N. Chakfé, B. Durand and G. Laroche, *Acta Biomaterialia*, 2009, **5**, 2418-2428.
- 86. G. Gigliobianco, C. K. Chong and S. MacNeil, *Journal of biomaterials applications*, 2015, **30**, 50-60.
- 87. J. Kanczler, J. Barry, P. Ginty, S. Howdle, K. Shakesheff and R. Oreffo, *Biochemical and biophysical research communications*, 2007, **352**, 135-141.
- 88. A. de Jesus Sousa-Batista, C. Cerqueira-Coutinho, F. S. do Carmo, M. de Souza Albernaz and R. Santos-Oliveira, *American journal of therapeutics*, 2019, **26**, e12-e17.
- 89. C. Bao, M. S. Chong, L. Qin, Y. Fan, E. Y. Teo, D. Sandikin, M. Choolani and J. K. Y. Chan, *Materials Technology*, 2019, 1-7.
- E. Niza, J. A. Castro-Osma, I. Posadas, C. Alonso-Moreno, I. Bravo, A. Garzón, J. Canales-Vázquez, V. Ceña, A. Lara-Sánchez and J. Albaladejo, *International journal of pharmaceutics*, 2019, 558, 110-119.
- 91. N. Filipović, L. Veselinović, S. Ražić, S. Jeremić, M. Filipič, B. Žegura, S. Tomić, M. Čolić and M. Stevanović, *Materials Science and Engineering: C*, 2019, **96**, 776-789.
- 92. Q. Hu, X. Gao, T. Kang, X. Feng, D. Jiang, Y. Tu, Q. Song, L. Yao, X. Jiang, H. Chen and J. Chen, *Biomaterials*, 2013, **34**, 9496-9508.
- 93. Y. C. Jiang, X. F. Wang, Y. Y. Xu, Y. H. Qiao, X. Guo, D. F. Wang, Q. Li and L. S. Turng, *Biomacromolecules*, 2018, **19**, 3747-3753.
- 94. J.-M. Lü, X. Wang, C. Marin-Muller, H. Wang, P. H. Lin, Q. Yao and C. Chen, *Expert* review of molecular diagnostics, 2009, **9**, 325-341.

- 95. F. Danhier, B. Vroman, N. Lecouturier, N. Crokart, V. Pourcelle, H. Freichels, C. Jérôme, J. Marchand-Brynaert, O. Feron and V. Préat, *Journal of Controlled Release*, 2009, **140**, 166-173.
- 96. I. d'Angelo, Y. Parajó, A. Horváth, G. Kéri, M. I. La Rotonda and M. J. Alonso, *Journal of microencapsulation*, 2010, **27**, 57-66.
- 97. T. Duan, Z. Xu, F. Sun, Y. Wang, J. Zhang, C. Luo and M. Wang, *Biomedicine & Pharmacotherapy*, 2019, **117**, 109121.
- 98. T. Kang, X. Gao, Q. Hu, D. Jiang, X. Feng, X. Zhang, Q. Song, L. Yao, M. Huang and X. Jiang, *Biomaterials*, 2014, **35**, 4319-4332.
- K. K. Chereddy, C.-H. Her, M. Comune, C. Moia, A. Lopes, P. E. Porporato, J. Vanacker, M. C. Lam, L. Steinstraesser and P. Sonveaux, *Journal of Controlled Release*, 2014, **194**, 138-147.
- 100. J. S. Park, H. N. Yang, S. W. Yi, J.-H. Kim and K.-H. Park, *Biomaterials*, 2016, **76**, 226-237.
- 101. Y. Parajó, I. d'Angelo, A. Horváth, T. Vantus, K. György, A. Welle, M. Garcia-Fuentes and M. J. Alonso, *European Journal of Pharmaceutical Sciences*, 2010, **41**, 644-649.
- 102. S. M. Ahsan, M. Thomas, K. K. Reddy, S. G. Sooraparaju, A. Asthana and I. Bhatnagar, *International journal of biological macromolecules*, 2018, **110**, 97-109.
- 103. A. Ali and S. Ahmed, *International journal of biological macromolecules*, 2018, **109**, 273-286.
- 104. D.-H. Ngo and S.-K. Kim, in *Advances in food and nutrition research*, Elsevier, 2014, vol. 73, pp. 15-31.
- 105. M. Z. Karagozlu and S.-K. Kim, in *Advances in food and nutrition research*, Elsevier, 2014, vol. 72, pp. 215-225.
- 106. L.-Y. Zheng and J.-F. Zhu, Carbohydrate polymers, 2003, 54, 527-530.
- 107. Y. Xu, Z. Wen and Z. Xu, Anticancer Research, 2009, 29, 5103-5109.
- 108. K. H. Prashanth and R. Tharanathan, *Biochimica et Biophysica Acta (BBA)-General* Subjects, 2005, **1722**, 22-29.
- 109. H. Wu, B. B. Aam, W. Wang, A. L. Norberg, M. Sørlie, V. G. Eijsink and Y. Du, *Carbohydrate polymers*, 2012, **89**, 511-518.
- 110. G. H. Kim, J. E. Won, Y. Byeon, M. G. Kim, T. I. Wi, J. M. Lee, Y. Y. Park, J. W. Lee, T. H. Kang, I. D. Jung, B. C. Shin, H. J. Ahn, Y. J. Lee, A. K. Sood, H. D. Han and Y. M. Park, *Drug Delivery*, 2018, **25**, 1394-1402.

- 111. H. Jin, J. Pi, F. Yang, C. Wu, X. Cheng, H. Bai, D. Huang, J. Jiang, J. Cai and Z. W. Chen, *Applied Microbiology and Biotechnology*, 2016, **100**, 6643-6652.
- 112. A. A. Zahid, R. Ahmed, S. R. ur Rehman, R. Augustine, M. Tariq and A. Hasan, *International journal of biological macromolecules*, 2019, **136**, 901-910.
- 113. X. Shen, J. Fang, X. Lv, Z. Pei, Y. Wang, S. Jiang and K. Ding, *Journal of Biological Chemistry*, 2011, **286**, 26616-26627.
- 114. J. Harenberg, G. Leber, C. Dempfle, D. Heene, R. Zimmermann and W. Kübler, *Nouvelle revue francaise d'hematologie*, 1989, **31**, 363-369.
- 115. R. Group, The Lancet, 1990, 336, 827-830.
- 116. J. M. Walenga and R. L. Bick, *Medical Clinics of North America*, 1998, 82, 635-658.
- 117. F. Alam, T. A. Al-Hilal, S. W. Chung, D. Seo, F. Mahmud, H. S. Kim, S. Y. Kim and Y. Byun, *Biomaterials*, 2014, **35**, 6543-6552.
- 118. H. J. Chung, H. K. Kim, J. J. Yoon and T. G. Park, *Pharmaceutical research*, 2006, **23**, 1835-1841.
- 119. M. M. Kemp, A. Kumar, S. Mousa, E. Dyskin, M. Yalcin, P. Ajayan, R. J. Linhardt and S. A. Mousa, *nanotechnology*, 2009, **20**, 455104.
- 120. D. Y. Lee, S. K. Kim, Y. S. Kim, J. H. Nam, I. San Kim, R. W. Park, S. Y. Kim and Y. Byun, *Journal of Controlled Release*, 2007, **118**, 310-317.
- 121. M. d. P. Rodriguez-Torres, L. S. Acosta-Torres and L. A. Diaz-Torres, *Journal of Nanomaterials*, 2018, **2018**.
- 122. I.-K. Park, T. H. Tran, I.-H. Oh, Y.-J. Kim, K. J. Cho, K. M. Huh and Y.-k. Lee, *European Journal of Pharmaceutical Sciences*, 2010, **41**, 148-155.
- 123. Z. Khatun, M. Nurunnabi, K. J. Cho, Y. Byun, Y. H. Bae and Y.-k. Lee, *Journal of controlled release*, 2014, **177**, 64-73.
- 124. Y. Liang and K. L. Kiick, Acta biomaterialia, 2014, 10, 1588-1600.
- 125. K. Park, Y. S. Kim, G. Y. Lee, R. W. Park, I. S. Kim, S. Y. Kim and Y. Byun, *Pharmaceutical Research*, 2008, **25**, 2786-2798.
- 126. M. McLuckie, C. A. Schmidt, A. Oosthuysen, N. Sanchez-Macedo, H. Merker, D. Bezuidenhout, S. P. Hoerstrup and N. Lindenblatt, *Journal of Biomedical Materials Research Part A*, 2017, **105**, 2543-2550.
- 127. K. Su and C. Wang, *Biotechnology letters*, 2015, **37**, 2139-2145.
- 128. M. Foox and M. Zilberman, *Expert opinion on drug delivery*, 2015, **12**, 1547-1563.

- 129. N. Sahoo, R. K. Sahoo, N. Biswas, A. Guha and K. Kuotsu, *International journal of biological macromolecules*, 2015, **81**, 317-331.
- 130. W.-T. Kuo, H.-Y. Huang, M.-J. Chou, M.-C. Wu and Y.-Y. Huang, *Journal of Nanomaterials*, 2011, **2011**, 28.
- 131. S. Balthasar, K. Michaelis, N. Dinauer, H. von Briesen, J. Kreuter and K. Langer, *Biomaterials*, 2005, **26**, 2723-2732.
- 132. S. F. Lin, P. L. Jiang, J. S. Tsai, Y. Y. Huang, S. Y. Lin, J. H. Lin and D. Z. Liu, *Journal of Biomedical Materials Research Part B: Applied Biomaterials*, 2019, **107**, 1228-1237.
- 133. S. R. ElMasry, R. M. Hathout, M. Abdel-Halim and S. Mansour, *Journal of Drug Delivery Science and Technology*, 2018, **48**, 30-39.
- 134. S. Kommareddy and M. Amiji, *Cancer gene therapy*, 2007, **14**, 488.
- 135. J. Xie, H. Wang, Y. Wang, F. Ren, W. Yi, K. Zhao, Z. Li, Q. Zhao, Z. Liu and H. Wu, *Cardiovascular therapeutics*, 2013, **31**, e12-e18.
- 136. R. B. Montero, X. Vial, D. T. Nguyen, S. Farhand, M. Reardon, S. M. Pham, G. Tsechpenakis and F. M. Andreopoulos, *Acta biomaterialia*, 2012, **8**, 1778-1791.
- 137. A. O. Elzoghby, W. M. Samy and N. A. Elgindy, *Journal of controlled release*, 2012, **157**, 168-182.
- 138. W. Lin, A. Coombes, M. Davies, S. Davis and L. Illum, *Journal of Drug Targeting*, 1993, **1**, 237-243.
- 139. J. Y. Jun, H. H. Nguyen, H. S. Chun, B.-C. Kang and S. Ko, *Food chemistry*, 2011, **127**, 1892-1898.
- 140. G. V. Patil, Drug development research, 2003, 58, 219-247.
- 141. S. Sundar, J. Kundu and S. C. Kundu, *Science and Technology of Advanced Materials*, 2010, **11**, 014104.
- 142. H. Hao, Q. Ma, C. Huang, F. He and P. Yao, *International journal of pharmaceutics*, 2013, **444**, 77-84.
- 143. S. H. Lee, D. Heng, W. K. Ng, H.-K. Chan and R. B. Tan, *International journal of pharmaceutics*, 2011, **403**, 192-200.
- 144. S. H. Choi, H. J. Byeon, J. S. Choi, L. Thao, I. Kim, E. S. Lee, B. S. Shin, K. C. Lee and Y. S. Youn, *Journal of controlled release*, 2015, **197**, 199-207.
- 145. E. Miele, G. P. Spinelli, E. Miele, F. Tomao and S. Tomao, *International journal of nanomedicine*, 2009, **4**, 99.

- 146. F. Kratz, Journal of controlled release, 2014, 190, 331-336.
- 147. L. Noorani, M. Stenzel, R. Liang, M. H. Pourgholami and D. L. Morris, *Journal of nanobiotechnology*, 2015, **13**, 25.
- 148. V. Kushwah, S. S. Katiyar, C. P. Dora, A. K. Agrawal, D. A. Lamprou, R. C. Gupta and S. Jain, *Acta biomaterialia*, 2018, **73**, 424-436.
- 149. V. Gonzalez-Villasana, C. Rodriguez-Aguayo, T. Arumugam, Z. Cruz-Monserrate, E. Fuentes-Mattei, D. Deng, R. F. Hwang, H. Wang, C. Ivan and R. J. Garza, *Molecular cancer therapeutics*, 2014, **13**, 2583-2594.
- 150. S. Son, S. Song, S. J. Lee, S. Min, S. A. Kim, J. Y. Yhee, M. S. Huh, I. C. Kwon, S. Y. Jeong and Y. Byun, *Biomaterials*, 2013, **34**, 9475-9485.
- 151. S. Wagner, F. Rothweiler, M. G. Anhorn, D. Sauer, I. Riemann, E. C. Weiss, A. Katsen-Globa, M. Michaelis, J. Cinatl Jr and D. Schwartz, *Biomaterials*, 2010, **31**, 2388-2398.
- 152. T.-P. Fan, J.-C. Yeh, K. W. Leung, P. Y. Yue and R. N. Wong, *Trends in Pharmacological Sciences*, 2006, **27**, 297-309.
- 153. M. B. Majnooni, S. Fakhri, A. Smeriglio, D. Trombetta, C. R. Croley, P. Bhattacharyya, E. Sobarzo-Sánchez, M. H. Farzaei and A. Bishayee, *Molecules*, 2019, **24**.
- 154. A. B. Kunnumakkara, P. Anand and B. B. Aggarwal, Cancer letters, 2008, 269, 199-225.
- 155. B. Shan, C. Schaaf, A. Schmidt, K. Lucia, M. Buchfelder, M. Losa, D. Kuhlen, J. Kreutzer, M. Perone and E. Arzt, *Journal of Endocrinology*, 2012, **214**, 389.
- 156. J. L. Arbiser, N. Klauber, R. Rohan, R. Van Leeuwen, M. T. Huang, C. Fisher, E. Flynn and H. R. Byers, *Molecular Medicine*, 1998, **4**, 376-383.
- 157. S. S. Bhandarkar and J. L. Arbiser, in *The molecular targets and therapeutic uses of curcumin in health and disease*, Springer, 2007, pp. 185-195.
- 158. R. K. Basniwal, H. S. Buttar, V. Jain and N. Jain, *Journal of agricultural and food chemistry*, 2011, **59**, 2056-2061.
- M. M. Yallapu, P. K. B. Nagesh, M. Jaggi and S. C. Chauhan, *The AAPS journal*, 2015, 17, 1341-1356.
- 160. L. Li, F. S. Braiteh and R. Kurzrock, *Cancer: Interdisciplinary International Journal of the American Cancer Society*, 2005, **104**, 1322-1331.
- 161. C. Gong, S. Deng, Q. Wu, M. Xiang, X. Wei, L. Li, X. Gao, B. Wang, L. Sun and Y. Chen, *Biomaterials*, 2013, **34**, 1413-1432.
- 162. X. Yang, Z. Li, N. Wang, L. Li, L. Song, T. He, L. Sun, Z. Wang, Q. Wu and N. Luo, *Scientific reports*, 2015, **5**, 10322.

- 163. Q. Ding, T. Niu, Y. Yang, Q. Guo, F. Luo and Z. Qian, *Journal of biomedical nanotechnology*, 2014, **10**, 632-641.
- 164. A. Mukerjee, A. P. Ranjan and J. K. Vishwanatha, *Journal of biomedical nanotechnology*, 2016, **12**, 1374-1392.
- 165. A. P. Ranjan, A. Mukerjee, L. Helson, R. Gupta and J. K. Vishwanatha, *Anticancer research*, 2013, **33**, 3603-3609.
- 166. H.-L. Tan, K.-G. Chan, P. Pusparajah, S. Saokaew, A. Duangjai, L.-H. Lee and B.-H. Goh, *Frontiers in pharmacology*, 2016, **7**, 191.
- 167. X. Yu, Y. Tong, X. Q. Han, H. F. Kwok, G. G. L. Yue, C. Bik-San Lau and W. Ge, *Phytotherapy Research*, 2013, **27**, 1368-1375.
- 168. Z. Da, Journal of International Translational Medicine, 2014, 2, 385-388.
- 169. J. Hong, Z. Zhang, W. Lv, M. Zhang, C. Chen, S. Yang, S. Li, L. Zhang, D. Han and W. Zhang, *PLoS One*, 2013, **8**, e71347.
- 170. Y.-h. YE, F.-h. HU, J.-p. Zou, Y. Zhan, S.-q. Wang and J.-y. Liu, *Chinese Academy of Medical Sciences*, 2015, **37**, 264-268.
- 171. J.-X. Yang, I. Fichtner, M. Becker, M. Lemm and X.-M. Wang, *The American journal of Chinese medicine*, 2009, **37**, 1153-1165.
- 172. S. Li, S. J. Priceman, H. Xin, W. Zhang, J. Deng, Y. Liu, J. Huang, W. Zhu, M. Chen and W. Hu, *PLoS One*, 2013, **8**, e81657.
- 173. H. J. Choi, J.-S. Eun, D. K. Kim, R. H. Li, T.-Y. Shin, H. Park, N.-P. Cho and Y. Soh, *European journal of pharmacology*, 2008, **579**, 58-65.
- 174. X. Jing, W. Yin, H. Tian, M. Chen, X. Yao, W. Zhu, F. Guo and Y. Ye, *Life sciences*, 2018, **202**, 52-60.
- 175. Y. Wu, L. Xia, Y. Zhou, W. Ma, N. Zhang, J. Chang, K. Lin, Y. Xu and X. Jiang, *Journal of Materials Chemistry B*, 2015, **3**, 4871-4883.
- 176. Y. Tang, A. Jacobi, C. Vater, L. Zou, X. Zou and M. Stiehler, *Stem cells*, 2015, **33**, 1863-1877.
- 177. Y. Zheng, L. Lu, Z. Yan, S. Jiang, S. Yang, Y. Zhang, K. Xu, C. He, X. Tao and Q. Zhang, *Artificial cells, nanomedicine, and biotechnology*, 2019, **47**, 801-811.
- 178. L.-Y. Han, Y.-L. Wu, C.-Y. Zhu, C.-S. Wu and C.-R. Yang, *Pharmaceutics*, 2019, **11**, 51.
- 179. M. Gong, C. Chi, J. Ye, M. Liao, W. Xie, C. Wu, R. Shi and L. Zhang, *Colloids and Surfaces B: Biointerfaces*, 2018, **170**, 201-209.

- 180. M. Jang, L. Cai, G. O. Udeani, K. V. Slowing, C. F. Thomas, C. W. Beecher, H. H. Fong, N. R. Farnsworth, A. D. Kinghorn and R. G. Mehta, *Science*, 1997, 275, 218-220.
- 181. J. A. Baur and D. A. Sinclair, *Nature reviews Drug discovery*, 2006, 5, 493.
- 182. S.-H. Tseng, S.-M. Lin, J.-C. Chen, Y.-H. Su, H.-Y. Huang, C.-K. Chen, P.-Y. Lin and Y. Chen, *Clinical Cancer Research*, 2004, **10**, 2190-2202.
- 183. S. Garvin, K. Öllinger and C. Dabrosin, *Cancer letters*, 2006, 231, 113-122.
- 184. M. Fouad, A. Agha, M. A. Merzabani and S. Shouman, *Human & experimental toxicology*, 2013, **32**, 1067-1080.
- 185. Y. Kimura and H. Okuda, *The Journal of nutrition*, 2001, **131**, 1844-1849.
- 186. J. C. Wong and R. R. Fiscus, Anticancer research, 2015, 35, 273-281.
- 187. F. Simão, A. S. Pagnussat, J. H. Seo, D. Navaratna, W. Leung, J. Lok, S. Guo, C. Waeber, C. G. Salbego and E. H. Lo, *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*, 2012, **32**, 884-895.
- 188. W.-H. Hu, R. Duan, Y.-T. Xia, Q.-P. Xiong, H.-Y. Wang, G. K.-L. Chan, S.-Y. Liu, T. T.-X. Dong, Q.-W. Qin and K. W.-K. Tsim, *Journal of agricultural and food chemistry*, 2018, **67**, 1127-1137.
- 189. P.-L. Chen and A. S Easton, *Current neurovascular research*, 2011, 8, 14-24.
- 190. A. C. Santos, I. Pereira, M. Pereira-Silva, L. Ferreira, M. Caldas, M. Collado-González, M. Magalhães, A. Figueiras, A. J. Ribeiro and F. Veiga, *Colloids and Surfaces B: Biointerfaces*, 2019, **180**, 127-140.
- 191. D. Arora and S. Jaglan, *Environmental chemistry letters*, 2018, 16, 35-41.
- 192. S. Pund, R. Thakur, U. More and A. Joshi, *Colloids and Surfaces B: Biointerfaces*, 2014, **120**, 110-117.
- 193. S. Kim, W. K. Ng, Y. Dong, S. Das and R. B. Tan, *Journal of Food Engineering*, 2012, **108**, 37-42.
- 194. D. G. Kingston, Journal of Natural Products, 2000, 63, 726-734.
- 195. J.-E. Damber, C. Vallbo, P. Albertsson, B. Lennernäs and K. Norrby, *Cancer Chemotherapy and Pharmacology*, 2006, **58**, 354-360.
- 196. Y. Dai, X. Cai, X. Bi, C. Liu, N. Yue, Y. Zhu, J. Zhou, M. Fu, W. Huang and H. Qian, *European Journal of Medicinal Chemistry*, 2019, **171**, 104-115.

- 197. S. K. Dordunoo, J. K. Jackson, L. A. Arsenault, A. M. C. Oktaba, W. L. Hunter and H. M. Burt, *Cancer chemotherapy and pharmacology*, 1995, **36**, 279-282.
- 198. D. Belotti, V. Vergani, T. Drudis, P. Borsotti, M. R. Pitelli, G. Viale, R. Giavazzi and G. Taraboletti, *Clinical Cancer Research*, 1996, **2**, 1843-1849.
- 199. J. Wang, P. Lou, R. Lesniewski and J. Henkin, Anti-cancer drugs, 2003, 14, 13-19.
- 200. A. Vacca, D. Ribatti, M. Iurlaro, F. Merchionne, B. Nico, R. Ria and F. Dammacco, *Journal of hematotherapy & stem cell research*, 2002, **11**, 103-118.
- 201. G. Bocci, A. Di Paolo and R. Danesi, Angiogenesis, 2013, 16, 481-492.
- 202. N. Klauber, S. Parangi, E. Flynn, E. Hamel and R. J. D'Amato, *Cancer research*, 1997, **57**, 81-86.
- 203. J. R. Merchan, D. R. Jayaram, J. G. Supko, X. He, G. J. Bubley and V. P. Sukhatme, *International journal of cancer*, 2005, **113**, 490-498.
- 204. K. Hata, M. Osaki, D. K. Dhar, K. Nakayama, R. Fujiwaki, H. Ito, N. Nagasue and K. Miyazaki, *Cancer Chemotherapy and Pharmacology*, 2004, **53**, 68-74.
- 205. F. Danhier, P. Danhier, C. J. De Saedeleer, A.-C. Fruytier, N. Schleich, A. des Rieux, P. Sonveaux, B. Gallez and V. Préat, *International journal of pharmaceutics*, 2015, 479, 399-407.
- 206. K. W. Kang, M.-K. Chun, O. Kim, R. K. Subedi, S.-G. Ahn, J.-H. Yoon and H.-K. Choi, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2010, **6**, 210-213.
- 207. P. Ma and R. J. Mumper, Journal of nanomedicine & nanotechnology, 2013, 4, 1000164.
- 208. I. Banerjee, K. De, D. Mukherjee, G. Dey, S. Chattopadhyay, M. Mukherjee, M. Mandal, A. K. Bandyopadhyay, A. Gupta and S. Ganguly, *Acta biomaterialia*, 2016, **38**, 69-81.
- 209. C. Wu, Y. Gao, Y. Liu and X. Xu, *International Journal of Nanomedicine*, 2018, **13**, 6189.
- 210. M. E. Wall, M. Wani, C. Cook, K. H. Palmer, A. a. McPhail and G. Sim, *Journal of the American Chemical Society*, 1966, **88**, 3888-3890.
- 211. E. Martino, S. Della Volpe, E. Terribile, E. Benetti, M. Sakaj, A. Centamore, A. Sala and S. Collina, *Bioorganic & medicinal chemistry letters*, 2017, **27**, 701-707.
- 212. Y. Q. Liu, W. Q. Li, S. L. Morris-Natschke, K. Qian, L. Yang, G. X. Zhu, X. B. Wu, A. L. Chen, S. Y. Zhang and X. Nan, *Medicinal research reviews*, 2015, **35**, 753-789.
- 213. N. V. Koshkina, E. S. Kleinerman, C. Waldrep, S.-F. Jia, L. L. Worth, B. E. Gilbert and V. Knight, *Clinical Cancer Research*, 2000, **6**, 2876-2880.

- C. Verschraegen, B. Gilbert, A. Huaringa, R. Newman, N. Harris, F. Leyva, L. Keus, K. Campbell, T. Nelson-Taylor and V. Knight, *Annals of the New York Academy of Sciences*, 2000, **922**, 352-354.
- 215. C. H. Takimoto and R. Thomas, *Annals of the New York Academy of Sciences*, 2000, **922**, 224-236.
- 216. G. Pratesi, G. L. Beretta and F. Zunino, Anti-cancer drugs, 2004, 15, 545-552.
- 217. Y. Pommier, M. Cushman and J. H. Doroshow, Oncotarget, 2018, 9, 37286.
- 218. M. K. Clements, C. B. Jones, M. Cumming and S. S. Daoud, *Cancer Chemotherapy and Pharmacology*, 1999, **44**, 411-416.
- C. L. Gigliotti, R. Minelli, R. Cavalli, S. Occhipinti, G. Barrera, S. Pizzimenti, G. Cappellano, E. Boggio, L. Conti and R. Fantozzi, *Journal of biomedical nanotechnology*, 2016, 12, 114-127.
- 220. M. Dai, X. Xu, J. Song, S. Fu, M. Gou, F. Luo and Z. Qian, *Cancer letters*, 2011, **312**, 189-196.
- 221. R. G. P. T. Jayasooriya, S. R. Park, Y. H. Choi, J.-W. Hyun, W.-Y. Chang and G.-Y. Kim, *Environmental toxicology and pharmacology*, 2015, **39**, 1189-1198.
- 222. Y. Wen, Y. Wang, X. Liu, W. Zhang, X. Xiong, Z. Han and X. Liang, *Cancer biology & medicine*, 2017, **14**, 363.
- C. L. Gigliotti, B. Ferrara, S. Occhipinti, E. Boggio, G. Barrera, S. Pizzimenti, M. Giovarelli, R. Fantozzi, A. Chiocchetti and M. Argenziano, *Drug delivery*, 2017, 24, 670-680.
- 224. C.-J. Lin, Y.-L. Lin, F. Luh, Y. Yen and R.-M. Chen, Oncotarget, 2016, 7, 42408.
- 225. X. Tian, M. Nguyen, H. P. Foote, J. M. Caster, K. C. Roche, C. G. Peters, P. Wu, L. Jayaraman, E. G. Garmey and J. E. Tepper, *Cancer research*, 2017, **77**, 112-122.
- 226. S. Gaur, L. Chen, T. Yen, Y. Wang, B. Zhou, M. Davis and Y. Yen, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2012, **8**, 721-730.
- 227. S. J. Conley, T. L. Baker, J. P. Burnett, R. L. Theisen, D. Lazarus, C. G. Peters, S. G. Clouthier, S. Eliasof and M. S. Wicha, *Breast cancer research and treatment*, 2015, **150**, 559-567.
- 228. T. Doura, K. Takahashi, Y. Ogra and N. Suzuki, *ACS medicinal chemistry letters*, 2017, **8**, 211-214.
- 229. J. Seguin, N. Mignet, H. L. Ossa, M. Tanter and J.-L. Gennisson, *Ultrasound in medicine & biology*, 2017, **43**, 2352-2361.

- 230. G. R. Pettit, S. B. Singh, M. L. Niven, E. Hamel and J. M. Schmidt, *Journal of natural products*, 1987, **50**, 119-131.
- 231. G. M. Tozer, C. Kanthou, C. S. Parkins and S. A. Hill, *International journal of experimental pathology*, 2002, **83**, 21-38.
- 232. C. M. West and P. Price, Anti-cancer drugs, 2004, 15, 179-187.
- 233. C. H. Shen, J. J. Shee, J. Y. Wu, Y. W. Lin, J. D. Wu and Y. W. Liu, *British journal of pharmacology*, 2010, **160**, 2008-2027.
- 234. J. Griggs, J. C. Metcalfe and R. Hesketh, *The lancet oncology*, 2001, **2**, 82-87.
- 235. M. Su, J. Huang, S. Liu, Y. Xiao, X. Qin, J. Liu, C. Pi, T. Luo, J. Li and X. Chen, *Scientific reports*, 2016, **6**, 28139.
- 236. X. Ren, M. Dai, L. P. Lin, P. K. Li and J. Ding, *British journal of pharmacology*, 2009, **156**, 1228-1238.
- M. E. Nik, A. A. Momtazi-Borojeni, P. Zamani, J. G. Navashenaq, M. Iranshahi, M. R. Jaafari and B. Malaekeh-Nikouei, *Journal of Cellular Physiology*, 2019, 234, 14721-14733.
- 238. V. Mico, A. Charalambous, S. A. Peyman, R. H. Abou-Saleh, A. F. Markham, P. L. Coletta and S. D. Evans, *International journal of pharmaceutics*, 2017, **526**, 547-555.
- 239. X. Li, M. Wu, L. Pan and J. Shi, *International journal of nanomedicine*, 2016, **11**, 93.
- 240. Y. Wang, H. Yu, D. Zhang, G. Wang, W. Song, Y. Liu, S. Ma, Z. Tang, Z. Liu and K. Sakurai, *Acta biomaterialia*, 2019, **92**, 229-240.
- 241. R. Kumar, A. Singh, N. Garg and P. F. Siril, *Ultrasonics sonochemistry*, 2018, **40**, 686-696.
- 242. D. Peer, J. M. Karp, S. Hong, O. C. Farokhzad, R. Margalit and R. Langer, *Nature nanotechnology*, 2007, **2**, 751.
- 243. D. Pozzi, V. Colapicchioni, G. Caracciolo, S. Piovesana, A. L. Capriotti, S. Palchetti, S. De Grossi, A. Riccioli, H. Amenitsch and A. Laganà, *Nanoscale*, 2014, **6**, 2782-2792.
- 244. H. Hatakeyama, H. Akita and H. Harashima, *Advanced drug delivery reviews*, 2011, **63**, 152-160.
- 245. A. Akbarzadeh, R. Rezaei-Sadabady, S. Davaran, S. W. Joo, N. Zarghami, Y. Hanifehpour, M. Samiei, M. Kouhi and K. Nejati-Koshki, *Nanoscale research letters*, 2013, **8**, 102.
- 246. V. P. Torchilin, *Nature reviews Drug discovery*, 2005, 4, 145.

- B. S. Pattni, V. V. Chupin and V. P. Torchilin, *Chemical reviews*, 2015, **115**, 10938-10966.
- 248. V. Torchilin, *Critical reviews in therapeutic drug carrier systems*, 1985, **2**, 65-115.
- 249. S. Mura, J. Nicolas and P. Couvreur, *Nature materials*, 2013, **12**, 991.
- 250. Y. C. Barenholz, Journal of controlled release, 2012, 160, 117-134.
- 251. K. Gardikis, C. Tsimplouli, K. Dimas, M. Micha-Screttas and C. Demetzos, *International journal of pharmaceutics*, 2010, **402**, 231-237.
- 252. H.-H. Chou, K.-L. Wang, C.-A. Chen, L.-H. Wei, C.-H. Lai, C.-Y. Hsieh, Y.-C. Yang, N.-F. Twu, T.-C. Chang and M.-S. Yen, *Gynecologic oncology*, 2006, **101**, 423-428.
- 253. A. Fassas and A. Anagnostopoulos, *Leukemia & lymphoma*, 2005, 46, 795-802.
- 254. M. Rahimi-Rad, E. Alizadeh and R. Samarei, *Pneumologia (Bucharest, Romania)*, 2011, **60**, 85-86.
- 255. I. Pont, A. Calatayud-Pascual, A. López-Castellano, E. P. Albelda, E. García-España, L. Martí-Bonmatí, J. C. Frias and M. T. Albelda, *PloS one*, 2018, **13**, e0190540-e0190540.
- 256. M. Üner and G. Yener, *International journal of nanomedicine*, 2007, **2**, 289.
- 257. N. Zhang, Q. Ping, G. Huang, W. Xu, Y. Cheng and X. Han, *International journal of pharmaceutics*, 2006, **327**, 153-159.
- 258. E. Rostami, S. Kashanian, A. H. Azandaryani, H. Faramarzi, J. E. N. Dolatabadi and K. Omidfar, *Chemistry and physics of lipids*, 2014, **181**, 56-61.
- 259. A. Costa, B. Sarmento and V. Seabra, *European Journal of Pharmaceutical Sciences*, 2018, **114**, 103-113.
- 260. J. O. Muga, J. W. Gathirwa, M. Tukulula and W. G. Jura, *Malaria journal*, 2018, 17, 133.
- 261. D. P. Gaspar and A. J. Almeida, in *Surface Modification of Nanoparticles for Targeted Drug Delivery*, Springer, 2019, pp. 73-98.
- 262. L. Bayón-Cordero, I. Alkorta and L. Arana, Nanomaterials, 2019, 9, 474.
- A. M. Brioschi, S. Calderoni, L. G. Pradotto, M. Guido, A. Strada, F. Zenga, C. A. Benech, F. Benech, L. Serpe and G. P. Zara, *Journal of Nanoneuroscience*, 2009, 1, 65-74.
- 264. S. Rani, R. Rana, G. K. Saraogi, V. Kumar and U. Gupta, *AAPS PharmSciTech*, 2019, **20**, 129.

- 265. B. Singh, S. Beg, R. K. Khurana, P. S. Sandhu, R. Kaur and O. P. Katare, *Critical Reviews in Therapeutic Drug Carrier Systems*, 2014, **31**, 121-185.
- 266. H. Y. Cho, J. H. Kang, L. Ngo, P. Tran and Y. B. Lee, *Journal of Nanomaterials*, 2016, 2016.
- 267. W. Huang, H. Su, L. Wen, A. Shao, F. Yang and G. Chen, *Journal of Drug Delivery Science and Technology*, 2018, **48**, 266-273.
- 268. G. R. Valicherla, K. M. Dave, A. A. Syed, M. Riyazuddin, A. P. Gupta, A. Singh, K. Mitra, D. Datta and J. R. Gayen, *Scientific reports*, 2016, **6**, 26895.
- 269. Y. V. Ramshankar, S. Suresh and K. Devi, *Clinical Research and Regulatory Affairs*, 2008, **25**, 213-234.
- 270. R. Nazari-Vanani, N. Azarpira, H. Heli, K. Karimian and N. Sattarahmady, *Colloids and Surfaces B: Biointerfaces*, 2017, **160**, 65-72.
- 271. S. Pund, G. Borade and G. Rasve, *Phytomedicine*, 2014, **21**, 307-314.
- 272. R. Goyal, L. K. Macri, H. M. Kaplan and J. Kohn, *Journal of Controlled Release*, 2016, 240, 77-92.
- 273. S. Abid, T. Hussain, Z. A. Raza and A. Nazir, *Materials Science and Engineering: C*, 2019, **97**, 966-977.
- 274. R. Mammadov, B. Mammadov, S. Toksoz, B. Aydin, R. Yagci, A. B. Tekinay and M. O. Guler, *Biomacromolecules*, 2011, **12**, 3508-3519.
- 275. V. A. Kumar, N. L. Taylor, S. Shi, B. K. Wang, A. A. Jalan, M. K. Kang, N. C. Wickremasinghe and J. D. Hartgerink, *ACS nano*, 2015, **9**, 860-868.
- 276. R. H. Zha, S. Sur, J. Boekhoven, H. Y. Shi, M. Zhang and S. I. Stupp, *Acta biomaterialia*, 2015, **12**, 1-10.
- 277. R. Fan, X. Li, J. Deng, X. Gao, L. Zhou, Y. Zheng, A. Tong, X. Zhang, C. You and G. Guo, *Scientific reports*, 2016, **6**, 28373.
- 278. H. Zigdon-Giladi, A. Khutaba, R. Elimelech, E. E. Machtei and S. Srouji, *Journal of Biomedical Materials Research Part A*, 2017, **105**, 2712-2721.
- 279. H.-J. Lai, C.-H. Kuan, H.-C. Wu, J.-C. Tsai, T.-M. Chen, D.-J. Hsieh and T.-W. Wang, *Acta biomaterialia*, 2014, **10**, 4156-4166.
- 280. W. Li, D. Wu, S. Zhu, Z. Liu, B. Luo, L. Lu and C. Zhou, *Chemical Engineering Journal*, 2019, **365**, 270-281.
- 281. X. Ren, Y. Han, J. Wang, Y. Jiang, Z. Yi, H. Xu and Q. Ke, *Acta biomaterialia*, 2018, **70**, 140-153.

- 282. W. Gao, L. Sun, X. Fu, Z. Lin, W. Xie, W. Zhang, F. Zhao and X. Chen, *Journal of Materials Chemistry B*, 2018, **6**, 277-288.
- 283. A. Satish and P. S. Korrapati, AAPS PharmSciTech, 2019, 20, 110.
- 284. D. Zhang, L. Li, Y. Shan, J. Xiong, Z. Hu, Y. Zhang and J. Gao, *Journal of Drug Delivery Science and Technology*, 2019, **52**, 272-281.
- C. Cha, S. R. Shin, N. Annabi, M. R. Dokmeci and A. Khademhosseini, *ACS nano*, 2013, 7, 2891-2897.
- A. Bianco, K. Kostarelos and M. Prato, *Expert opinion on drug delivery*, 2008, 5, 331-342.
- 287. D.-J. Lim, M. Sim, L. Oh, K. Lim and H. Park, *Archives of pharmacal research*, 2014, **37**, 43-52.
- 288. C. Caoduro, E. Hervouet, C. Girard-Thernier, T. Gharbi, H. Boulahdour, R. Delage-Mourroux and M. Pudlo, *Acta biomaterialia*, 2017, **49**, 36-44.
- 289. X. Hu, S. Liu, G. Zhou, Y. Huang, Z. Xie and X. Jing, *Journal of controlled release*, 2014, **185**, 12-21.
- 290. R. Wang, H. Cui, J. Wang, N. Li, Q. Zhao, Y. Zhou, Z. Lv and W. Zhong, *RSC Advances*, 2016, **6**, 47272-47280.
- 291. R. Singh and S. V. Torti, Advanced drug delivery reviews, 2013, 65, 2045-2060.
- 292. S. Murugesan, S. A. Mousa, L. J. O'Connor, D. W. Lincoln and R. J. Linhardt, *FEBS letters*, 2007, **581**, 1157-1160.
- 293. M. Wierzbicki, E. Sawosz, M. Grodzik, M. Prasek, S. Jaworski and A. Chwalibog, *Nanoscale research letters*, 2013, **8**, 195.
- 294. A. Albini, V. Mussi, A. Parodi, A. Ventura, E. Principi, S. Tegami, M. Rocchia, E. Francheschi, I. Sogno and R. Cammarota, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2010, 6, 277-288.
- 295. A. Masotti, M. R. Miller, A. Celluzzi, L. Rose, F. Micciulla, P. W. Hadoke, S. Bellucci and A. Caporali, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2016, **12**, 1511-1522.
- 296. Y. Su, Y. Hu, Y. Wang, X. Xu, Y. Yuan, Y. Li, Z. Wang, K. Chen, F. Zhang and X. Ding, *Biomaterials*, 2017, **139**, 75-90.
- 297. N. Azad, A. K. V. Iyer, L. Wang, Y. Liu, Y. Lu and Y. Rojanasakul, *Nanotoxicology*, 2013, **7**, 157-168.

- 298. D. Roman, A. Yasmeen, M. Mireuta, I. Stiharu and A.-E. Al Moustafa, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2013, **9**, 945-950.
- 299. P.-X. Lai, C.-W. Chen, S.-C. Wei, T.-Y. Lin, H.-J. Jian, I. P.-J. Lai, J.-Y. Mao, P.-H. Hsu, H.-J. Lin and W.-S. Tzou, *Biomaterials*, 2016, **109**, 12-22.
- 300. S. Shi, K. Yang, H. Hong, H. F. Valdovinos, T. R. Nayak, Y. Zhang, C. P. Theuer, T. E. Barnhart, Z. Liu and W. Cai, *Biomaterials*, 2013, **34**, 3002-3009.
- 301. S. Mukherjee, P. Sriram, A. K. Barui, S. K. Nethi, V. Veeriah, S. Chatterjee, K. I. Suresh and C. R. Patra, *Advanced healthcare materials*, 2015, **4**, 1722-1732.
- 302. Y. Chen, A. Gao, L. Bai, Y. Wang, X. Wang, X. Zhang, X. Huang, R. Hang, B. Tang and P. K. Chu, *Materials Science and Engineering: C*, 2017, **75**, 1049-1058.
- 303. Y. Zhu, J. Li, W. Li, Y. Zhang, X. Yang, N. Chen, Y. Sun, Y. Zhao, C. Fan and Q. Huang, *Theranostics*, 2012, **2**, 302.
- 304. Z.-Y. Lien, T.-C. Hsu, K.-K. Liu, W.-S. Liao, K.-C. Hwang and J.-I. Chao, *Biomaterials*, 2012, **33**, 6172-6185.
- 305. J. Xiao, X. Duan, Q. Yin, Z. Zhang, H. Yu and Y. Li, Biomaterials, 2013, 34, 9648-9656.
- 306. Z. Zhang, B. Niu, J. Chen, X. He, X. Bao, J. Zhu, H. Yu and Y. Li, *Biomaterials*, 2014, **35**, 4565-4572.
- 307. C. Cui, Y. Wang, K. Yang, Y. Wang, J. Yang, J. Xi, M. Zhao, J. Wu and S. Peng, *Journal of biomedical nanotechnology*, 2015, **11**, 70-80.
- 308. S. Pacelli, F. Acosta, A. R. Chakravarti, S. G. Samanta, J. Whitlow, S. Modaresi, R. P. H. Ahmed, J. Rajasingh and A. Paul, *Acta biomaterialia*, 2017, **58**, 479-491.
- 309. M. M. Schimke, R. Stigler, X. Wu, T. Waag, P. Buschmann, J. Kern, G. Untergasser, M. Rasse, D. Steinmüller-Nethl and A. Krueger, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2016, **12**, 823-833.
- 310. M. I. Setyawati, V. N. Mochalin and D. T. Leong, ACS nano, 2015, 10, 1170-1181.
- 311. D. Chen, C. A. Dougherty, K. Zhu and H. Hong, *Journal of controlled release*, 2015, **210**, 230-245.
- 312. J. Guerra, M. A. Herrero, B. Carrion, F. C. Pérez-Martínez, M. Lucío, N. Rubio, M. Meneghetti, M. Prato, V. Ceña and E. Vázquez, *Carbon*, 2012, **50**, 2832-2844.
- 313. K. Murata, K. Kaneko, F. Kokai, K. Takahashi, M. Yudasaka and S. Iijima, *Chemical physics letters*, 2000, **331**, 14-20.
- 314. N. Karousis, I. Suarez-Martinez, C. P. Ewels and N. Tagmatarchis, *Chemical reviews*, 2016, **116**, 4850-4883.

- 315. M. Zhang, T. Yamaguchi, S. Iijima and M. Yudasaka, *Nanomedicine: Nanotechnology*, *Biology and Medicine*, 2013, **9**, 657-664.
- 316. T. Azami, D. Kasuya, R. Yuge, M. Yudasaka, S. Iijima, T. Yoshitake and Y. Kubo, *The Journal of Physical Chemistry C*, 2008, **112**, 1330-1334.
- 317. J. Yang, H. Su, W. Sun, J. Cai, S. Liu, Y. Chai and C. Zhang, *Theranostics*, 2018, **8**, 1966.
- 318. K. Ajima, T. Murakami, Y. Mizoguchi, K. Tsuchida, T. Ichihashi, S. Iijima and M. Yudasaka, *ACS nano*, 2008, **2**, 2057-2064.
- M. Zhang, T. Murakami, K. Ajima, K. Tsuchida, A. S. Sandanayaka, O. Ito, S. Iijima and M. Yudasaka, *Proceedings of the National Academy of Sciences*, 2008, **105**, 14773-14778.
- 320. N. Li, Q. Zhao, C. Shu, X. Ma, R. Li, H. Shen and W. Zhong, *International journal of pharmaceutics*, 2015, **478**, 644-654.
- 321. S. Goodarzi, T. Da Ros, J. Conde, F. Sefat and M. Mozafari, *Materials Today*, 2017, **20**, 460-480.
- 322. J. Xu, H. Wang, Y. Hu, Y. S. Zhang, L. Wen, F. Yin, Z. Wang, Y. Zhang, S. Li and Y. Miao, *Advanced Science*, 2019, 1801233.
- 323. A. Grebinyk, S. Prylutska, S. Grebinyk, Y. Prylutskyy, U. Ritter, O. Matyshevska, T. Dandekar and M. Frohme, *Nanoscale research letters*, 2019, **14**, 61.
- 324. H. Kazemzadeh and M. Mozafari, *Drug Discovery Today*, 2019, 24, 898-905.
- 325. F. Jiao, Y. Liu, Y. Qu, W. Li, G. Zhou, C. Ge, Y. Li, B. Sun and C. Chen, *Carbon*, 2010, **48**, 2231-2243.
- 326. J.-J. Yin, F. Lao, P. P. Fu, W. G. Wamer, Y. Zhao, P. C. Wang, Y. Qiu, B. Sun, G. Xing and J. Dong, *Biomaterials*, 2009, **30**, 611-621.
- 327. H. Meng, G. Xing, B. Sun, F. Zhao, H. Lei, W. Li, Y. Song, Z. Chen, H. Yuan and X. Wang, *ACS nano*, 2010, **4**, 2773-2783.
- 328. R. Shukla, T. P. Thomas, J. Peters, A. Kotlyar, A. Myc and J. R. Baker Jr, *Chemical communications*, 2005, 5739-5741.
- 329. X. Ding, Y. Su, C. Wang, F. Zhang, K. Chen, Y. Wang, M. Li and W. Wang, ACS Applied Materials and Interfaces, 2017, 9, 23353-23369.
- 330. S. R. ur Rehman, R. Augustine, A. A. Zahid, R. Ahmed and A. Hasan, 2019.
- P. Chaudhuri, R. Harfouche, S. Soni, D. M. Hentschel and S. Sengupta, ACS Nano, 2010, 4, 574-582.

- 332. L. Pizzorno, Integrative Medicine: A Clinician's Journal, 2015, 14, 35.
- 333. M. Dzondo-Gadet, R. Mayap-Nzietchueng, K. Hess, P. Nabet, F. Belleville and B. Dousset, *Biological Trace Element Research*, 2002, **85**, 23-33.
- 334. P. Balasubramanian, T. Buettner, V. M. Pacheco and A. R. Boccaccini, *Journal of the European Ceramic Society*, 2018, **38**, 855-869.
- 335. X. Liu, M. N. Rahaman and D. E. Day, *Journal of Materials Science: Materials in Medicine*, 2013, **24**, 583-595.
- 336. S. Chen, Q. Yang, R. K. Brow, K. Liu, K. A. Brow, Y. Ma and H. Shi, *Materials Science and Engineering: C*, 2017, **73**, 447-455.
- 337. L. A. H. Durand, A. Góngora, J. M. P. López, A. R. Boccaccini, M. P. Zago, A. Baldi and A. Gorustovich, *Journal of Materials Chemistry B*, 2014, **2**, 7620-7630.
- L. A. H. Durand, G. E. Vargas, N. M. Romero, R. Vera-Mesones, J. M. Porto-López, A. R. Boccaccini, M. P. Zago, A. Baldi and A. Gorustovich, *Journal of Materials Chemistry* B, 2015, 3, 1142-1148.
- 339. S. B. Jung and D. E. Day, *Journal*, 2012.
- 340. P. Wray, Am. Ceram. Soc. Bull, 2011, 90, 24-31.
- 341. L. L. Hench, Annals of the New York academy of sciences, 1988, 523, 54-71.
- 342. D. E. Clapham, *Cell*, 1995, **80**, 259-268.
- 343. M. J. Berridge, P. Lipp and M. D. Bootman, *Nature reviews Molecular cell biology*, 2000, **1**, 11.
- 344. L. Munaron, C. Distasi, V. Carabelli, F. M. Baccino, G. Bonelli and D. Lovisolo, *The Journal of physiology*, 1995, **484**, 557-566.
- 345. H. m. Wu, Y. Yuan, D. C. Zawieja, J. Tinsley and H. J. Granger, *American Journal of Physiology-Heart and Circulatory Physiology*, 1999, **276**, H535-H542.
- 346. L. Munaron and A. Fiorio Pla, *Journal of cellular physiology*, 2000, **185**, 454-463.
- M. J. Cross and L. Claesson-Welsh, *Trends in pharmacological sciences*, 2001, 22, 201-207.
- 348. K. Liang, Z. Jiang, B. Zhao, J. Shen, D. Huang and L. Tao, *British Journal of Ophthalmology*, 2012, **96**, 1246-1251.
- 349. A. Malhotra and P. Habibovic, *Trends in biotechnology*, 2016, **34**, 983-992.
- 350. L. L. Hench, Journal of the European Ceramic Society, 2009, 29, 1257-1265.

- 351. S. Jung, D. Day, T. Day, W. Stoecker and P. Taylor, *Wound Repair and Regeneration*, 2011, **19**.
- 352. S. Kargozar, F. Baino, S. J. Hoseini, S. Hamzehlou, M. Darroudi, J. Verdi, L. Hasanzadeh, H.-W. Kim and M. Mozafari, *Nanomedicine*, 2018, **13**, 3051-3069.
- 353. S. Das, S. Singh, J. M. Dowding, S. Oommen, A. Kumar, T. X. Sayle, S. Saraf, C. R. Patra, N. E. Vlahakis and D. C. Sayle, *Biomaterials*, 2012, **33**, 7746-7755.
- 354. S. Das, S. Chigurupati, J. Dowding, P. Munusamy, D. R. Baer, J. F. McGinnis, M. P. Mattson, W. Self and S. Seal, *MRS Bulletin*, 2014, **39**, 976-983.
- 355. S. Chigurupati, M. R. Mughal, E. Okun, S. Das, A. Kumar, M. McCaffery, S. Seal and M. P. Mattson, *Biomaterials*, 2013, **34**, 2194-2201.
- 356. S. K. Nethi, H. S. Nanda, T. W. Steele and C. R. Patra, *Journal of Materials Chemistry B*, 2017, **5**, 9371-9383.
- 357. R. Augustine, A. Hasan, N. K. Patan, Y. B. Dalvi, R. Varghese, A. Antony, R. N. Unni, N. Sandhyarani and A.-E. A. Moustafa, *ACS Biomaterials Science & Engineering*, 2019.
- 358. R. Augustine, Y. B. Dalvi, P. Dan, N. George, D. Helle, R. Varghese, S. Thomas, P. Menu and N. Sandhyarani, *ACS Biomaterials Science & Engineering*, 2018, **4**, 4338-4353.
- 359. M. A. Saghiri, J. Orangi, A. Asatourian, C. M. Sorenson and N. Sheibani, *Critical reviews in oncology/hematology*, 2016, **98**, 290-301.
- 360. J. M. Dowding, S. Das, A. Kumar, T. Dosani, R. McCormack, A. Gupta, T. X. Sayle, D. C. Sayle, L. von Kalm and S. Seal, *ACS nano*, 2013, **7**, 4855-4868.
- 361. M. S. Lord, B. Tsoi, C. Gunawan, W. Y. Teoh, R. Amal and J. M. Whitelock, *Biomaterials*, 2013, **34**, 8808-8818.
- 362. A. Salinas, S. Shruti, G. Malavasi, L. Menabue and M. Vallet-Regí, *Acta biomaterialia*, 2011, **7**, 3452-3458.
- 363. S. Shruti, A. J. Salinas, G. Malavasi, G. Lusvardi, L. Menabue, C. Ferrara, P. Mustarelli and M. Vallet-Regì, *Journal of Materials Chemistry*, 2012, **22**, 13698-13706.
- 364. S. Shruti, F. Andreatta, E. Furlani, E. Marin, S. Maschio and L. Fedrizzi, *Applied Surface Science*, 2016, **378**, 216-223.
- 365. G. Chachami, G. Simos, A. Hatziefthimiou, S. Bonanou, P.-A. Molyvdas and E. Paraskeva, *American journal of respiratory cell and molecular biology*, 2004, **31**, 544-551.

- 366. C. Bracken, M. Whitelaw and D. Peet, *Cellular and Molecular Life Sciences CMLS*, 2003, **60**, 1376-1393.
- 367. G. L. Wang, B.-H. Jiang, E. A. Rue and G. L. Semenza, *Proceedings of the national academy of sciences*, 1995, **92**, 5510-5514.
- 368. P. Carmeliet and R. K. Jain, *Nature*, 2011, **473**, 298.
- 369. J. Muñoz-Sánchez and M. E. Chánez-Cárdenas, *Journal of Applied Toxicology*, 2019, **39**, 556-570.
- 370. R. M. Peters, P. Willemse, P. C. Rijk, M. Hoogendoorn and W. P. Zijlstra, *Case reports in orthopedics*, 2017, **2017**.
- 371. B. Green, E. Griffiths and S. Almond, *BMC psychiatry*, 2017, **17**, 33.
- 372. C. Jantzen, H. L. Jørgensen, B. R. Duus, S. L. Sporring and J. B. Lauritzen, *Acta orthopaedica*, 2013, **84**, 229-236.
- 373. C. Wu, Y. Zhou, W. Fan, P. Han, J. Chang, J. Yuen, M. Zhang and Y. Xiao, *Biomaterials*, 2012, **33**, 2076-2085.
- 374. M. M. Azevedo, O. Tsigkou, R. Nair, J. R. Jones, G. Jell and M. M. Stevens, *Tissue Engineering Part A*, 2014, **21**, 382-389.
- 375. M. Azevedo, G. Jell, M. O'donnell, R. Law, R. Hill and M. Stevens, *Journal of Materials Chemistry*, 2010, **20**, 8854-8864.
- 376. E. Littmann, H. Autefage, A. Solanki, C. Kallepitis, J. Jones, M. Alini, M. Peroglio and M. Stevens, *Journal of the European Ceramic Society*, 2018, **38**, 877-886.
- 377. H. Ren, Y. Cao, Q. Zhao, J. Li, C. Zhou, L. Liao, M. Jia, Q. Zhao, H. Cai and Z. C. Han, *Biochemical and biophysical research communications*, 2006, **347**, 12-21.
- 378. H.-H. Lee, C.-C. Chang, M.-J. Shieh, J.-P. Wang, Y.-T. Chen, T.-H. Young and S.-C. Hung, *Scientific reports*, 2013, **3**, 2683.
- 379. S. Zhou, Z. Cui and J. P. Urban, Arthritis & Rheumatism, 2004, 50, 3915-3924.
- 380. R. Amarilio, S. V. Viukov, A. Sharir, I. Eshkar-Oren, R. S. Johnson and E. Zelzer, *Development*, 2007, **134**, 3917-3928.
- C. Zhang, F. Yang, R. Cornelia, W. Tang, S. Swisher and H. Kim, *Bone*, 2011, 48, 507-513.
- 382. S. Kargozar, N. Lotfibakhshaiesh, J. Ai, A. Samadikuchaksaraie, R. G. Hill, P. A. Shah, P. B. Milan, M. Mozafari, M. Fathi and M. T. Joghataei, *Journal of Non-Crystalline Solids*, 2016, 449, 133-140.

- 383. S. Kargozar, F. Baino, N. Lotfibakhshaiesh, R. G. Hill, P. B. Milan, S. Hamzehlou, M. T. Joghataei and M. Mozafari, *Materials Today: Proceedings*, 2018, **5**, 15768-15775.
- 384. D. Lison, P. Carbonnelle, L. Mollo, R. Lauwerys and B. Fubini, *Chemical research in toxicology*, 1995, **8**, 600-606.
- 385. P. Wild, A. Perdrix, S. Romazini, J.-J. Moulin and F. Pellet, *Occupational and environmental medicine*, 2000, **57**, 568-573.
- 386. J. Tanaka, H. Moriyama, M. Terada, T. Takada, E. Suzuki, I. Narita, Y. Kawabata, T. Yamaguchi, A. Hebisawa and F. Sakai, *BMJ open*, 2014, **4**, e004407.
- 387. L.-Z. Liu, M. Ding, J. Z. Zheng, Y. Zhu, B. A. Fenderson, B. Li, J. Y. Jing and B.-H. Jiang, *Biological trace element research*, 2015, **166**, 57-65.
- 388. E. Urso and M. Maffia, Journal of vascular research, 2015, 52, 172-196.
- 389. Q.-f. Li, X.-q. Ding and Y. J. Kang, *The Journal of nutritional biochemistry*, 2014, **25**, 44-49.
- 390. D. C. Rigiracciolo, A. Scarpelli, R. Lappano, A. Pisano, M. F. Santolla, P. De Marco, F. Cirillo, A. R. Cappello, V. Dolce and A. Belfiore, *Oncotarget*, 2015, **6**, 34158.
- 391. G. Mavria, Y. Vercoulen, M. Yeo, H. Paterson, M. Karasarides, R. Marais, D. Bird and C. J. Marshall, *Cancer cell*, 2006, **9**, 33-44.
- 392. L. D. D'Andrea, A. Romanelli, R. Di Stasi and C. Pedone, *Dalton Transactions*, 2010, **39**, 7625-7636.
- 393. L. Finney, S. Vogt, T. Fukai and D. Glesne, *Clinical and Experimental Pharmacology and Physiology*, 2009, **36**, 88-94.
- 394. A. Nasulewicz, A. Mazur and A. Opolski, *Journal of Trace Elements in Medicine and Biology*, 2004, **18**, 1-8.
- 395. V. Goodman, G. Brewer and S. Merajver, *Endocrine-related cancer*, 2004, **11**, 255-263.
- 396. S. Brem, *Cancer Control*, 1999, **6**, 1-18.
- 397. Q. Pan, C. G. Kleer, K. L. Van Golen, J. Irani, K. M. Bottema, C. Bias, M. De Carvalho, E. A. Mesri, D. M. Robins and R. D. Dick, *Cancer research*, 2002, 62, 4854-4859.
- 398. G. Bührer, U. Rottensteiner, A. Hoppe, R. Detsch, D. Dafinova, T. Fey, P. Greil, C. Weis, J. P. Beier and A. R. Boccacini, *Biomedical glasses*, 2016, **2**.
- 399. S. Zhao, L. Li, H. Wang, Y. Zhang, X. Cheng, N. Zhou, M. N. Rahaman, Z. Liu, W. Huang and C. Zhang, *Biomaterials*, 2015, **53**, 379-391.

- 400. L. Bi, M. N. Rahaman, D. E. Day, Z. Brown, C. Samujh, X. Liu, A. Mohammadkhah, V. Dusevich, J. D. Eick and L. F. Bonewald, *Acta biomaterialia*, 2013, **9**, 8015-8026.
- 401. R. J. Watters, R. F. Brown and D. E. Day, *Biomedical glasses*, 2015, 1.
- 402. C. Wu, Y. Zhou, M. Xu, P. Han, L. Chen, J. Chang and Y. Xiao, *Biomaterials*, 2013, **34**, 422-433.
- 403. F. Baino, Journal of Biomedical Materials Research Part A, 2015, 103, 1259-1275.
- 404. C. Wang, K. Jin, J. He, J. Wang, X. Yang, C. Yao, X. Dai, C. Gao, Z. Gou and J. Ye, *Journal of Biomedical Nanotechnology*, 2018, **14**, 688-697.
- 405. F. Baino, I. Potestio and C. Vitale-Brovarone, *Materials*, 2018, 11, 1524.
- 406. Z. Chen, H. Meng, G. Xing, C. Chen, Y. Zhao, G. Jia, T. Wang, H. Yuan, C. Ye and F. Zhao, *Toxicology letters*, 2006, **163**, 109-120.
- 407. N. Mroczek-Sosnowska, E. Sawosz, K. Vadalasetty, M. Łukasiewicz, J. Niemiec, M. Wierzbicki, M. Kutwin, S. Jaworski and A. Chwalibog, *International journal of molecular sciences*, 2015, 16, 4838-4849.
- 408. M. Maloubier, D. K. Shuh, S. G. Minasian, J. I. Pacold, P.-L. Solari, H. Michel, F. o. R. Oberhaensli, Y. Bottein, M. Monfort and C. Moulin, *Environmental science & technology*, 2016, **50**, 10730-10738.
- 409. B. McMahon, P. Mauer, C. P. McCoy, T. C. Lee and T. Gunnlaugsson, *Journal of the American Chemical Society*, 2009, **131**, 17542-17543.
- 410. Y. Fan, P. Yang, S. Huang, J. Jiang, H. Lian and J. Lin, *The Journal of Physical Chemistry C*, 2009, **113**, 7826-7830.
- 411. G. Miao, X. Chen, C. Mao, X. Li, Y. Li and C. Lin, *Journal of sol-gel science and technology*, 2014, **69**, 250-259.
- 412. G. Li, G. Liang, S. Zhao, K. Ma, W. Feng, D. Zhou and X. Liu, *Advances in Applied Ceramics*, 2015, **114**, 164-174.
- C. R. Patra, R. Bhattacharya, S. Patra, N. E. Vlahakis, A. Gabashvili, Y. Koltypin, A. Gedanken, P. Mukherjee and D. Mukhopadhyay, *Advanced Materials*, 2008, 20, 753-756.
- 414. R. Augustine, S. K. Nethi, N. Kalarikkal, S. Thomas and C. R. Patra, *Journal of Materials Chemistry B*, 2017, **5**, 4660-4672.
- 415. C. R. Patra, J.-H. Kim, K. Pramanik, L. V. d'Uscio, S. Patra, K. Pal, R. Ramchandran, M. S. Strano and D. Mukhopadhyay, *Nano letters*, 2011, **11**, 4932-4938.

- 416. H. Zhao, O. J. Osborne, S. Lin, Z. Ji, R. Damoiseux, Y. Wang, A. E. Nel and S. Lin, *Small*, 2016, **12**, 4404-4411.
- 417. V. S. Bollu, S. K. Nethi, R. K. Dasari, S. S. N. Rao, S. Misra and C. R. Patra, *Nanotoxicology*, 2016, **10**, 413-425.
- 418. X. Ma, Y. Liu, X. M. Wu, C. Wang, Q. Li and T. Fu, *Materials Technology*, 2016, **31**, 23-27.
- K. Ge, W. Sun, S. Zhang, S. Wang, G. Jia, C. Zhang and J. Zhang, *RSC Advances*, 2016, 6, 21725-21734.
- 420. R. Bhattacharya, P. Mukherjee, Z. Xiong, A. Atala, S. Soker and D. Mukhopadhyay, *Nano Letters*, 2004, **4**, 2479-2481.
- 421. P. Mukherjee, R. Bhattacharya, P. Wang, L. Wang, S. Basu, J. A. Nagy, A. Atala, D. Mukhopadhyay and S. Soker, *Clinical Cancer Research*, 2005, **11**, 3530.
- 422. D. J. Maxwell, J. R. Taylor and S. Nie, *Journal of the American Chemical Society*, 2002, **124**, 9606-9612.
- 423. N. Ferrara, H.-P. Gerber and J. LeCouter, *Nature Medicine*, 2003, 9, 669-676.
- 424. R. Bhattacharya and P. Mukherjee, *Advanced Drug Delivery Reviews*, 2008, **60**, 1289-1306.
- 425. R. S. Darweesh, N. M. Ayoub and S. Nazzal, *International Journal of Nanomedicine*, 2019, **14**, 7643-7663.
- 426. R. R. Arvizo, S. Rana, O. R. Miranda, R. Bhattacharya, V. M. Rotello and P. Mukherjee, *Nanomedicine : nanotechnology, biology, and medicine*, 2011, **7**, 580-587.
- 427. S. Jain, D. G. Hirst and J. M. O'Sullivan, *The British Journal of Radiology*, 2012, **85**, 101-113.
- 428. P. J. Barnard and S. J. Berners-Price, *Coordination Chemistry Reviews*, 2007, **251**, 1889-1902.
- 429. R. R. Arvizo, S. Saha, E. Wang, J. D. Robertson, R. Bhattacharya and P. Mukherjee, *Proceedings of the National Academy of Sciences*, 2013, **110**, 6700-6705.
- 430. J. P. Thiery, H. Acloque, R. Y. J. Huang and M. A. Nieto, *Cell*, 2009, **139**, 871-890.
- 431. Vesselina G. Cooke, Valerie S. LeBleu, D. Keskin, Z. Khan, Joyce T. O'Connell, Y. Teng, Michael B. Duncan, L. Xie, G. Maeda, S. Vong, H. Sugimoto, Rafael M. Rocha, A. Damascena, Ricardo R. Brentani and R. Kalluri, *Cancer Cell*, 2012, 21, 66-81.
- 432. J. Conde, M. Larguinho, A. Cordeiro, L. R. Raposo, P. M. Costa, S. Santos, M. S. Diniz, A. R. Fernandes and P. V. Baptista, *Nanotoxicology*, 2014, **8**, 521-532.

- 433. P. Mukherjee, R. Bhattacharya, N. Bone, Y. K. Lee, C. R. Patra, S. Wang, L. Lu, C. Secreto, P. C. Banerjee, M. J. Yaszemski, N. E. Kay and D. Mukhopadhyay, *Journal of Nanobiotechnology*, 2007, 5, 4.
- 434. J. D. Mangadlao, X. Wang, C. McCleese, M. Escamilla, G. Ramamurthy, Z. Wang, M. Govande, J. P. Basilion and C. Burda, *ACS Nano*, 2018, **12**, 3714-3725.
- 435. N. Abbaspour, R. Hurrell and R. Kelishadi, *Journal of research in medical sciences: the official journal of Isfahan University of Medical Sciences*, 2014, **19**, 164.
- 436. T. Ohara, K. Noma, S. Urano, S. Watanabe, S. Nishitani, Y. Tomono, F. Kimura, S. Kagawa, Y. Shirakawa and T. Fujiwara, *International journal of cancer*, 2013, **132**, 2705-2713.
- 437. N. T. Le and D. R. Richardson, *Biochimica et Biophysica Acta (BBA)-Reviews on Cancer*, 2002, **1603**, 31-46.
- 438. G. f. Hu, Journal of cellular biochemistry, 1998, **69**, 326-335.
- 439. A. Patel and J. Knowles, *Journal of Materials Science: Materials in Medicine*, 2006, **17**, 937-944.
- 440. M. S. Mohammadi, I. Ahmed, N. Muja, S. Almeida, C. Rudd, M. Bureau and S. Nazhat, *Acta biomaterialia*, 2012, **8**, 1616-1626.
- 441. L. Zhu, C. Staley, D. Kooby, B. El-Rays, H. Mao and L. Yang, *Cancer letters*, 2017, **388**, 139-148.
- 442. K. Maier-Hauff, F. Ulrich, D. Nestler, H. Niehoff, P. Wust, B. Thiesen, H. Orawa, V. Budach and A. Jordan, *Journal of neuro-oncology*, 2011, **103**, 317-324.
- 443. F. M. Kievit and M. Zhang, Accounts of chemical research, 2011, 44, 853-862.
- 444. M. V. Yigit, A. Moore and Z. Medarova, *Pharmaceutical research*, 2012, **29**, 1180-1188.
- 445. N. Dissanayake, K. Current and S. Obare, *International journal of molecular sciences*, 2015, **16**, 23482-23516.
- 446. R. Oruch, M. A. Elderbi, H. A. Khattab, I. F. Pryme and A. Lund, *European journal of pharmacology*, 2014, **740**, 464-473.
- 447. C.-T. Chiu, Z. Wang, J. G. Hunsberger and D.-M. Chuang, *Pharmacological reviews*, 2013, **65**, 105-142.
- 448. C. D. Mao, P. Hoang and P. E. DiCorleto, *Journal of Biological Chemistry*, 2001, **276**, 26180-26188.
- 449. Y. Takada, X. Fang, M. S. Jamaluddin, D. D. Boyd and B. B. Aggarwal, *Journal of Biological Chemistry*, 2004, **279**, 39541-39554.

- 450. J. J. GILES and J. G. BANNIGAN, The Journal of Anatomy, 1999, 194, 197-205.
- 451. S. Guo, K. Arai, M. F. Stins, D.-M. Chuang and E. H. Lo, Stroke, 2009, 40, 652-655.
- 452. L. F. Zeilbeck, B. Müller, V. Knobloch, E. R. Tamm and A. Ohlmann, *PloS one*, 2014, **9**, e95546.
- 453. L. Haro Durand, G. Vargas, R. Vera-Mesones, A. Baldi, M. Zago, M. Fanovich, A. Boccaccini and A. Gorustovich, *Materials*, 2017, **10**, 740.
- 454. P. M. Vilarinho, N. Barroca, S. Zlotnik, P. Félix and M. H. Fernandes, *Materials Science and Engineering: C*, 2014, **39**, 395-402.
- 455. R. Brückner, M. Tylkowski, L. Hupa and D. S. Brauer, *Journal of Materials Chemistry B*, 2016, **4**, 3121-3134.
- 456. V. Miguez-Pacheco, T. Büttner, A. Maçon, J. Jones, T. Fey, D. De Ligny, P. Greil, J. Chevalier, A. Malchere and A. Boccaccini, *Journal of Non-Crystalline Solids*, 2016, **432**, 65-72.
- 457. P. Han, M. Xu, J. Chang, N. Chakravorty, C. Wu and Y. Xiao, *Biomaterials Science*, 2014, **2**, 1230-1243.
- 458. D. Bernardini, A. Nasulewicz, A. Mazur and J. Maier, *Front Biosci*, 2005, **10**, 1177-1182.
- 459. W. Zhai, H. Lu, L. Chen, X. Lin, Y. Huang, K. Dai, K. Naoki, G. Chen and J. Chang, *Acta Biomaterialia*, 2012, **8**, 341-349.
- 460. A. Shamosi, M. Farokhi, J. Ai and E. Sharifi, *Journal of Medical Hypotheses and Ideas*, 2015, **9**, 94-98.
- 461. N. J. Hallab, S. Anderson, M. Caicedo and J. J. Jacobs, *Journal of ASTM International*, 2006, **3**, 1-12.
- 462. V. Miguez-Pacheco, D. De Ligny, J. Schmidt, R. Detsch and A. Boccaccini, *Journal of the European Ceramic Society*, 2018, **38**, 871-876.
- 463. E. Takeda, Y. Taketani, N. Sawada, T. Sato and H. Yamamoto, *Biofactors*, 2004, **21**, 345-355.
- 464. H. Jin, S.-H. Chang, C.-X. Xu, J.-Y. Shin, Y.-S. Chung, S.-J. Park, Y.-S. Lee, G.-H. An, K.-H. Lee and M.-H. Cho, *Toxicological sciences*, 2007, **100**, 215-223.
- 465. T. Kume, Trends in cardiovascular medicine, 2008, 18, 224-228.
- 466. M. Hirama, F. Takahashi, K. Takahashi, S. Akutagawa, K. Shimizu, S. Soma, Y. Shimanuki, K. Nishio and Y. Fukuchi, *Cancer letters*, 2003, **198**, 107-117.

- 467. C. E. Camalier, M. Yi, L. R. Yu, B. L. Hood, K. A. Conrads, Y. J. Lee, Y. Lin, L. M. Garneys, G. F. Bouloux and M. R. Young, *Journal of cellular physiology*, 2013, **228**, 1536-1550.
- 468. Y. Lin, K. E. McKinnon, S. W. Ha and G. R. Beck Jr, *Molecular carcinogenesis*, 2015, **54**, 926-934.
- 469. G. S. Di Marco, M. Hausberg, U. Hillebrand, P. Rustemeyer, W. Wittkowski, D. Lang and H. Pavenstadt, *American Journal of Physiology-Renal Physiology*, 2008, **294**, F1381-F1387.
- 470. S. Kargozar, F. Baino, S. Hamzehlou, R. G. Hill and M. Mozafari, *Trends in biotechnology*, 2018, **36**, 430-444.
- 471. C. Stähli, N. Muja and S. N. Nazhat, *Tissue Engineering Part A*, 2012, **19**, 548-557.
- 472. I.-H. Lee, H.-s. Yu, N. J. Lakhkar, H.-W. Kim, M.-S. Gong, J. C. Knowles and I. B. Wall, *Materials Science and Engineering: C*, 2013, **33**, 2104-2112.
- 473. C. Ip, The Journal of nutrition, 1998, 128, 1845-1854.
- 474. G. Combs Jr, L. Clark and B. Turnbull, *Biofactors*, 2001, 14, 153-159.
- 475. K. El-Bayoumy, *Cancer research*, 1985, **45**, 3631-3635.
- 476. M. M. Jacobs, P. Shubik and R. Feldman, *Cancer letters*, 1980, 9, 353-357.
- 477. N. M. Corcoran, M. Najdovska and A. J. Costello, *The Journal of urology*, 2004, **171**, 907-910.
- 478. C. Jiang, H. Ganther and J. Lu, *Molecular Carcinogenesis: Published in cooperation with* the University of Texas MD Anderson Cancer Center, 2000, **29**, 236-250.
- C. Jiang, W. Jiang, C. Ip, H. Ganther and J. Lu, *Molecular Carcinogenesis: Published in cooperation with the University of Texas MD Anderson Cancer Center*, 1999, 26, 213-225.
- 480. A. P. Levy, N. S. Levy, S. Wegner and M. A. Goldberg, *Journal of Biological Chemistry*, 1995, **270**, 13333-13340.
- 481. K. Hirota, M. Matsui, S. Iwata, A. Nishiyama, K. Mori and J. Yodoi, *Proceedings of the National Academy of Sciences*, 1997, **94**, 3633-3638.
- 482. A. Gallegos, M. Berggren, J. R. Gasdaska and G. Powis, *Cancer research*, 1997, **57**, 4965-4970.
- 483. G. Spyrou, M. Björnstedt, S. Kumar and A. Holmgren, FEBS letters, 1995, 368, 59-63.

- 484. I. Y. Kim and T. C. Stadtman, *Proceedings of the National Academy of Sciences*, 1997, **94**, 12904-12907.
- 485. B. Aksakal and A. Boccaccini, *Materials Letters*, 2012, **76**, 177-180.
- 486. M. Stevanović, N. Filipović, J. Djurdjević, M. Lukić, M. Milenković and A. Boccaccini, *Colloids and Surfaces B: Biointerfaces*, 2015, **132**, 208-215.
- X. Wang, Y. Zhang, Y. Ma, D. Chen, H. Yang and M. Li, *Ceramics International*, 2016, 42, 3609-3617.
- 488. J. Duan, Y. Yu, Y. Li, Y. Yu and Z. Sun, *Biomaterials*, 2013, 34, 5853-5862.
- 489. J. Duan, Y. Yu, Y. Yu, Y. Li, P. Huang, X. Zhou, S. Peng and Z. Sun, *Particle and fibre toxicology*, 2014, **11**, 50.
- 490. H. Li and J. Chang, *Acta biomaterialia*, 2013, **9**, 6981-6991.
- 491. L.-C. Gerhardt, K. L. Widdows, M. M. Erol, C. W. Burch, J. A. Sanz-Herrera, I. Ochoa, R. Stämpfli, I. S. Roqan, S. Gabe and T. Ansari, *Biomaterials*, 2011, **32**, 4096-4108.
- 492. R. Detsch, P. Stoor, A. Grünewald, J. A. Roether, N. C. Lindfors and A. R. Boccaccini, *Journal of Biomedical Materials Research Part A*, 2014, **102**, 4055-4061.
- 493. C. Mao, X. Chen, G. Miao and C. Lin, *Biomedical materials*, 2015, 10, 025005.
- 494. R. M. Day, A. R. Boccaccini, S. Shurey, J. A. Roether, A. Forbes, L. L. Hench and S. M. Gabe, *Biomaterials*, 2004, **25**, 5857-5866.
- 495. A. A. Gorustovich, J. A. Roether and A. R. Boccaccini, *Tissue Engineering Part B: Reviews*, 2009, **16**, 199-207.
- 496. H. Keshaw, A. Forbes and R. M. Day, *Biomaterials*, 2005, 26, 4171-4179.
- 497. V. Alt, D. V. Kögelmaier, K. S. Lips, V. Witt, S. Pacholke, C. Heiss, M. Kampschulte, S. Heinemann, T. Hanke and R. Schnettler, *Acta biomaterialia*, 2011, **7**, 3773-3779.
- 498. C. Wu and J. Chang, *Journal of Controlled Release*, 2014, **193**, 282-295.
- 499. G. Franci, A. Falanga, S. Galdiero, L. Palomba, M. Rai, G. Morelli and M. Galdiero, *Molecules*, 2015, **20**, 8856-8874.
- 500. S. Gurunathan, K.-J. Lee, K. Kalishwaralal, S. Sheikpranbabu, R. Vaidyanathan and S. H. Eom, *Biomaterials*, 2009, **30**, 6341-6350.
- 501. K. Kalishwaralal, E. Banumathi, S. R. K. Pandian, V. Deepak, J. Muniyandi, S. H. Eom and S. Gurunathan, *Colloids and Surfaces B: Biointerfaces*, 2009, **73**, 51-57.

- 502. J. Baharara, F. Namvar, M. Mousavi, T. Ramezani and R. Mohamad, *Molecules*, 2014, **19**, 13498-13508.
- 503. S. Bhagavathy and S. Kancharla, Int J Herb Med, 2016, 4, 24-29.
- 504. T. Yang, Q. Yao, F. Cao, Q. Liu, B. Liu and X. H. Wang, *International Journal of Nanomedicine*, 2016, **11**, 6679-6692.
- 505. K. Kang, D.-H. Lim, I.-H. Choi, T. Kang, K. Lee, E.-Y. Moon, Y. Yang, M.-S. Lee and J.-S. Lim, *Toxicology letters*, 2011, **205**, 227-234.
- 506. J. A. Milner, in *Nutrition and Cancer Prevention*, Springer, 2001, pp. 69-81.
- 507. B. Jiang, G. Tang, K. Cao, L. Wu and R. Wang, *Antioxidants & redox signaling*, 2010, **12**, 1167-1178.
- 508. C. Szabó and A. Papapetropoulos, British journal of pharmacology, 2011, 164, 853-865.
- 509. A. Papapetropoulos, A. Pyriochou, Z. Altaany, G. Yang, A. Marazioti, Z. Zhou, M. G. Jeschke, L. K. Branski, D. N. Herndon and R. Wang, *Proceedings of the National Academy of Sciences*, 2009, **106**, 21972-21977.
- 510. W.-J. Cai, M.-J. Wang, P. K. Moore, H.-M. Jin, T. Yao and Y.-C. Zhu, *Cardiovascular research*, 2007, **76**, 29-40.
- 511. X. M. van Wijk and T. H. van Kuppevelt, Angiogenesis, 2014, 17, 443-462.
- 512. K. Raman, R. Karuturi, V. P. Swarup, U. R. Desai and B. Kuberan, *Bioorganic & medicinal chemistry letters*, 2012, **22**, 4467-4470.
- 513. S. K. Nethi, A. K. Barui, V. S. Bollu, B. R. Rao and C. R. Patra, ACS Biomaterials Science & Engineering, 2017, **3**, 3635-3645.
- 514. D. Giustarini, A. Milzani, R. Colombo, I. Dalle-Donne and R. Rossi, *Clinica chimica acta*, 2003, **330**, 85-98.
- 515. R. Osman and M. Swain, *Materials*, 2015, **8**, 932-958.
- 516. Y. Sun, Y. Li, B. Wu, J. Wang, X. Lu, S. Qu, J. Weng and B. Feng, *Acta biomaterialia*, 2017, **58**, 527-538.
- 517. M.-A. Saghiri, A. Asatourian, F. Garcia-Godoy and N. Sheibani, *Medicina oral*, *patologia oral y cirugia bucal*, 2016, **21**, e514.
- 518. S. Spriano, S. Yamaguchi, F. Baino and S. Ferraris, Acta Biomaterialia, 2018, 79, 1-22.
- 519. A. L. Raines, R. Olivares-Navarrete, M. Wieland, D. L. Cochran, Z. Schwartz and B. D. Boyan, *Biomaterials*, 2010, **31**, 4909-4917.

- 520. F. Rupp, L. Scheideler, D. Rehbein, D. Axmann and J. Geis-Gerstorfer, *Biomaterials*, 2004, **25**, 1429-1438.
- 521. G. Zhao, Z. Schwartz, M. Wieland, F. Rupp, J. Geis-Gerstorfer, D. L. Cochran and B. Boyan, *Journal of Biomedical Materials Research Part A: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials, 2005*, **74**, 49-58.
- 522. Z. Schwartz, P. Raz, G. Zhao, Y. Barak, M. Tauber, H. Yao and B. D. Boyan, *The Journal of Bone and Joint Surgery. American volume.*, 2008, **90**, 2485.
- 523. B. Shi, O. Andrukhov, S. Berner, A. Schedle and X. Rausch-Fan, *Dental Materials*, 2014, **30**, 839-847.
- 524. N. An, A. Schedle, M. Wieland, O. Andrukhov, M. Matejka and X. Rausch-Fan, Journal of Biomedical Materials Research Part A: An Official Journal of The Society for Biomaterials, The Japanese Society for Biomaterials, and The Australian Society for Biomaterials and the Korean Society for Biomaterials, 2010, **93**, 364-372.
- 525. G. Krenning, M. J. van Luyn and M. C. Harmsen, *Trends in molecular medicine*, 2009, **15**, 180-189.
- 526. T. Ziebart, C.-H. Yoon, T. Trepels, A. Wietelmann, T. Braun, F. Kiessling, S. Stein, M. Grez, C. Ihling and M. Muhly-Reinholz, *Circulation research*, 2008, **103**, 1327-1334.
- 527. T. Ziebart, A. Schnell, C. Walter, P. W. Kämmerer, A. Pabst, K. M. Lehmann, J. Ziebart, M. O. Klein and B. Al-Nawas, *Clinical oral investigations*, 2013, **17**, 301-309.
- 528. W. Chen, K. Xu, B. Tao, L. Dai, Y. Yu, C. Mu, X. Shen, Y. Hu, Y. He and K. Cai, *Acta biomaterialia*, 2018, **74**, 489-504.
- 529. M. Zong, L. Bai, Y. Liu, X. Wang, X. Zhang, X. Huang, R. Hang and B. Tang, *Materials Science and Engineering: C*, 2017, **71**, 93-99.
- 530. D. H. Jo, J. H. Kim, Y. S. Yu, T. G. Lee and J. H. Kim, *Nanomedicine: Nanotechnology*, *Biology and Medicine*, 2012, **8**, 784-791.
- 531. R. Augustine, A. Hasan, N. K. Patan, A. Augustine, Y. B. Dalvi, R. Varghese, R. N. Unni, N. Kalarikkal, A. E. Al Moustafa and S. Thomas, *Macromolecular bioscience*, 2019, 1900058.
- 532. M. N. Rahaman, A. Yao, B. S. Bal, J. P. Garino and M. D. Ries, *Journal of the American Ceramic Society*, 2007, **90**, 1965-1988.
- 533. D. Schubert, R. Dargusch, J. Raitano and S.-W. Chan, *Biochemical and biophysical research communications*, 2006, **342**, 86-91.

- 534. R. Augustine, Y. B. Dalvi, V. Y. Nath, R. Varghese, V. Raghuveeran, A. Hasan, S. Thomas and N. Sandhyarani, *Materials Science and Engineering: C*, 2019, **103**, 109801.
- 535. J. M. Berg and Y. Shi, *Science*, 1996, **271**, 1081-1085.
- 536. T. Boehm, M. S. O'Reilly, K. Keough, J. Shiloach, R. Shapiro and J. Folkman, *Biochemical and biophysical research communications*, 1998, **252**, 190-194.
- 537. T. Kaji, Y. Fujiwara, C. Yamamoto, M. Sakamoto and H. Kozuka, *Life sciences*, 1994, **55**, 1781-1787.
- 538. S.-G. Shian, Y.-R. Kao, F. Y.-H. Wu and C.-W. Wu, *Molecular pharmacology*, 2003, **64**, 1076-1084.
- 539. L. Nardinocchi, V. Pantisano, R. Puca, M. Porru, A. Aiello, A. Grasselli, C. Leonetti, M. Safran, G. Rechavi and D. Givol, *PloS one*, 2010, **5**, e15048.
- 540. M. Sheffer, A. J. Simon, J. Jacob-Hirsch, G. Rechavi, E. Domany, D. Givol and G. D'Orazi, *Oncotarget*, 2011, **2**, 1191.
- 541. R. Augustine, E. A. Dominic, I. Reju, B. Kaimal, N. Kalarikkal and S. Thomas, *RSC Advances*, 2014, **4**, 51528-51536.
- 542. A. K. Barui, S. K. Nethi and C. R. Patra, *Journal of Materials Chemistry B*, 2017, **5**, 3391-3403.
- 543. R. Augustine, P. Dan, A. Sosnik, N. Kalarikkal, N. Tran, B. Vincent, S. Thomas, P. Menu and D. Rouxel, *Nano Research*, 2017, **10**, 3358-3376.
- 544. S. Mohamed, N. N. Parayath, S. Taurin and K. Greish, *Therapeutic delivery*, 2014, **5**, 1101-1121.
- 545. S. Giri, A. Karakoti, R. P. Graham, J. L. Maguire, C. M. Reilly, S. Seal, R. Rattan and V. Shridhar, *PLoS ONE*, 2013, **8**.
- 546. D. H. Jo, J. H. Kim, J. G. Son, N. W. Song, Y.-I. Kim, Y. S. Yu, T. G. Lee and J. H. Kim, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2014, **10**, e1109-e1117.
- 547. A. P. Pathak, W. E. Hochfeld, S. L. Goodman and M. S. Pepper, *Angiogenesis*, 2008, **11**, 321.
- 548. L. Tang, X. Sun, N. Liu, Z. Zhou, F. Yu, X. Zhang, X. Sun and X. Chen, *ACS Applied Nano Materials*, 2018, **1**, 1741-1749.
- 549. T. Wu, X. Ding, B. Su, A. Soodeen-Lalloo, L. Zhang and J.-Y. Shi, *Clinical and Translational Oncology*, 2018, **20**, 599-606.
- 550. F. Yan, X. Xu, Y. Chen, Z. Deng, H. Liu, J. Xu, J. Zhou, G. Tan, J. Wu and H. Zheng, *Ultrasound in medicine & biology*, 2015, **41**, 2765-2773.

- 551. M. O'Donnell, *Physica C: Superconductivity and its Applications*, 2018, **548**, 103-106.
- 552. G. Loudos, G. C. Kagadis and D. Psimadas, *European journal of radiology*, 2011, **78**, 287-295.
- 553. G. J. Strijkers, E. Kluza, G. A. Van Tilborg, D. W. van der Schaft, A. W. Griffioen, W. J. Mulder and K. Nicolay, *Angiogenesis*, 2010, **13**, 161-173.
- 554. H.-x. Yuan, W.-p. Wang, J.-x. Wen, L.-w. Lin, A. A. Exner, P.-s. Guan and X.-j. Chen, *Ultrasound in medicine & biology*, 2018, **44**, 1460-1467.
- 555. F. Debordeaux, L. Chansel-Debordeaux, J.-B. Pinaquy, P. Fernandez and J. Schulz, *Nuclear medicine and biology*, 2018, **62**, 31-46.
- 556. Y.-l. Chen, F.-q. Liu, Y. Guo, J. Cheng, L. Yang, M. Lu, P. Li, J. Xu, T. Yu and Z.-g. Wang, *Biomaterials science*, 2018, **6**, 2130-2143.
- 557. C. Giraudeau, F. Geffroy, S. Mériaux, F. Boumezbeur, P. Robert, M. Port, C. Robic, D. Le Bihan, F. Lethimonnier and J. Valette, *Angiogenesis*, 2013, **16**, 171-179.
- 558. M. Wu, Y. Zhang, Y. Zhang, M. Wu, M. Wu, H. Wu, L. Cao, L. Li, X. Li and X. Zhang, *RSC advances*, 2018, **8**, 1706-1716.
- 559. J. H. Ryu, J.-Y. Shin, S. A. Kim, S.-W. Kang, H. Kim, S. Kang, K. Choi, I. C. Kwon, B.-S. Kim and K. Kim, *Biomaterials*, 2013, **34**, 6871-6881.
- 560. J. D. Lewis, G. Destito, A. Zijlstra, M. J. Gonzalez, J. P. Quigley, M. Manchester and H. Stuhlmann, *Nature medicine*, 2006, **12**, 354.
- 561. N. F. Steinmetz, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2010, **6**, 634-641.
- 562. P. H. Beatty and J. D. Lewis, Advanced Drug Delivery Reviews, 2019, 145, 130-144.
- 563. X. Li, M. Wu, J. Wang, Y. Dou, X. Gong, Y. Liu, Q. Guo, X. Zhang, J. Chang and Y. Niu, *Nanomedicine: Nanotechnology, Biology and Medicine*, 2019, **15**, 252-263.
- 564. B. P. Burke, C. Cawthorne and S. J. Archibald, *Philosophical Transactions of the Royal* Society A: Mathematical, *Physical and Engineering Sciences*, 2017, **375**, 20170261.
- 565. X. Q. Zhang, R. Lam, X. Xu, E. K. Chow, H. J. Kim and D. Ho, *Advanced materials*, 2011, **23**, 4770-4775.
- 566. Y. Sun, X. Zhu, J. Peng and F. Li, Acs Nano, 2013, 7, 11290-11300.
- 567. S. Shi, K. Yang, H. Hong, F. Chen, H. F. Valdovinos, S. Goel, T. E. Barnhart, Z. Liu and W. Cai, *Biomaterials*, 2015, **39**, 39-46.

- 568. Y. Yang, S. Chen, H. Li, Y. Yuan, Z. Zhang, J. Xie, D. W. Hwang, A. Zhang, M. Liu and X. Zhou, *Nano letters*, 2018, **19**, 441-448.
- 569. B. Dubertret, P. Skourides, D. J. Norris, V. Noireaux, A. H. Brivanlou and A. Libchaber, *Science*, 2002, **298**, 1759-1762.
- 570. J. Zhang, G. Hao, C. Yao, J. Yu, J. Wang, W. Yang, C. Hu and B. Zhang, ACS applied materials & interfaces, 2016, 8, 16612-16621.
- 571. X. Xing, B. Zhang, X. Wang, F. Liu, D. Shi and Y. Cheng, *Biomaterials*, 2015, **48**, 16-25.
- 572. Y. Ma, H. Shen, M. Zhang and Z. Zhang, in *Nanomaterials for Tumor Targeting Theranostics: A Proactive Clinical Perspective*, World Scientific, 2016, pp. 85-141.
- 573. M. Stroh, J. P. Zimmer, D. G. Duda, T. S. Levchenko, K. S. Cohen, E. B. Brown, D. T. Scadden, V. P. Torchilin, M. G. Bawendi and D. Fukumura, *Nature medicine*, 2005, 11, 678.
- 574. S. Chen and P. I. Imoukhuede, Analytical Chemistry, 2019, 91, 7603-7612.
- 575. M. W. Kieran, R. Kalluri and Y.-J. Cho, *Cold Spring Harbor perspectives in medicine*, 2012, **2**, a006593.
- 576. J. M. Ebos and R. S. Kerbel, *Nature reviews Clinical oncology*, 2011, 8, 210.
- 577. P. Auguste, S. Lemiere, F. Larrieu-Lahargue and A. Bikfalvi, *Critical reviews in oncology/hematology*, 2005, **54**, 53-61.
- 578. I. H. Park, J. H. Sohn, S. B. Kim, K. S. Lee, J. S. Chung, S. H. Lee, T. Y. Kim, K. H. Jung, E. K. Cho and Y. S. Kim, *Cancer research and treatment: official journal of Korean Cancer Association*, 2017, **49**, 569.
- 579. B. A. Saeed, V. Lim, N. A. Yusof, K. Z. Khor, H. S. Rahman and N. A. Samad, *International journal of nanomedicine*, 2019, **14**, 5135.
- 580. D. H. Jo, J. H. Kim, J. G. Son, Y. Piao, T. G. Lee and J. H. Kim, *Nano research*, 2014, **7**, 844-852.
- 581. D. Guarnieri, M. A. Malvindi, V. Belli, P. P. Pompa and P. Netti, *Journal of nanoparticle research*, 2014, **16**, 2229.
- 582. R. Augustine, P. Prasad and I. M. N. Khalaf, *Materials Science and Engineering: C*, 2019.
- 583. R. Augustine, A. P. Mathew and A. Sosnik, Applied Materials Today, 2017, 7, 91-103.
- 584. S. K. Nethi, A. K. Barui, S. Mukherjee and C. R. Patra, *Antioxidants & redox signaling*, 2019, **30**, 786-809.

- 585. A. K. Barui, S. K. Nethi, S. Haque, P. Basuthakur and C. R. Patra, ACS Applied Bio Materials, 2019, **2**, 5492-5511.
- 586. I. Bilecka and M. Niederberger, *Nanoscale*, 2010, **2**, 1358-1374.
- 587. P. Rajasekharreddy, C. Huang, S. Busi, J. Rajkumari, M.-H. Tai and G. Liu, *Current medicinal chemistry*, 2019, **26**, 1311-1327.