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Development of a Prototype for the Analysis of Multiple Responses of the Autonomic Nervous System

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Abstract

The autonomic nervous system (ANS) drives different non-voluntary responses, which can be investigated by multiple sensors. We propose a modular hardware prototype for the noninvasive acquisition, processing and transmission of biological signals to analyze the ANS in synchrony with the video recording of the pupil. The implementation includes 1) two noninvasive sensors, a pulse oximeter and an electrodermal activity sensor, 2) a module able to collect the information and send it to the PC via USB and 3) a graphic user interface for visualization, synchronization and data saving. A series of experimental tests were performed to investigate the effect of different stimulations: light, dental occlusion, transcutaneous electrical nerve stimulation (TENS) and mental efforts. They indicate the reliability of the system and the importance of the joint detection of more signals for discriminating different states of the ANS. Specifically, heart rate, Galvanic response and pupil size were compared, showing some coherence in their oscillations and a different ability to discriminate between the stimulation conditions. Their joint detection is thus important for discriminating different states of the ANS.

Keywords: Pulse oximetry, Electrodermal Activity (EDA), Galvanic Skin Response (GSR), Pupillogram, Autonomous Nervous System (ANS), Transcutaneous Electrical Nerve Stimulation (TENS), Embedded systems.

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1. Introduction

The autonomic nervous system (ANS) is the portion of the central nervous system that controls unconscious activities, such as visceral functions and homeostasis [1]. It is divided into two main branches, the sympathetic and the parasympathetic, the first promoting the activation of a physiological response and the other inhibiting it. The ANS is profoundly affected by emotions and somatosensory inputs and plays an important role in pain and stress modulation and perception.

Autonomic testing finds application in the clinical assessment of neurological disorders, particularly those affecting predominantly small nerve fibres [2]. Many studies have been devoted to the quantitative assessment of the ANS response, dating back to more than 3 decades [3][4][5]. However, most of the literature takes into account just one of the numerous physiological systems that are affected by the ANS in turn.

Very frequently the works that have dealt with ANS in different disorders related to its dysfunction have focused on cardio-circulatory parameters [6]. However, there are many other potential peripheral effects of ANS which could be deepened. For example, a physiological system related to the ANS can be investigated measuring skin conductance, reflecting the sweating of the sweat glands [7]. Moreover, pupil is strongly affected by the ANS [8]. The study of mydriasis and myosis (i.e., the dilation and contraction of pupil) is usually done in a different context (vestibular system). However, recent studies have indicated the possibility of characterizing the condition of the ANS in healthy or pathological conditions, by analyzing the nonlinear pupil oscillations [9][10][11][12][13][14]. Moreover, the study of pupillary dynamics was considered useful for evaluating the arousal state during mental effort due to cognitive tasks [15][16], in relation to the involvement of reward systems [17] and in the development of Human-Machine Interfaces [18]. Furthermore, it has been suggested that the use of different ANS parameters, including pupil size, may be useful for better characterizing and quantifying the emotional component linked to the autonomous response [19]. Moreover, further evidence was provided that pupil can be used to evaluate the state of emotional arousal as well as the generic activation of the ANS [20].

One of the limitations of the study of various ANS responses is the use of different instruments for the analysis of different signals. However, it was argued that pupillogram could provide useful information for the study of the arousal state and that it would receive a valid contribution from the

38 association with signals already used for this purpose, such as skin conduc-
39 tance and electrocortical activity [21]. These observations would involve an
40 important expansion of the combined and synchronous study of various pa-
41 rameters associated with pupillography to aspects not only of pathology, but
42 also related to the emotional / affective state. Thus, important outcomes
43 could be expected both for clinical patients and for any study involving the
44 psychic assessment of the arousal state.

45 Only recently, more reactions of the ANS have been investigated simul-
46 taneously [22][23]. The signals that have been often analyzed are the cardiac
47 pulse and the variations of skin conductance. In this study, we are inter-
48 ested in the investigation of those signals jointly with the pupil response.
49 A modular hardware prototype is developed for the noninvasive acquisition,
50 processing and transmission of biological signals and is synchronized with
51 a commercial system for pupil investigation. Our present implementation
52 includes:

- 53 • two noninvasive sensors, a pulse oximeter and an electrodermal ac-
54 tivity sensor (both often used in the study of the responses of ANS
55 [24][25][26]);
- 56 • a module able to collect the information and send it to the PC via USB;
- 57 • a graphic user interface (GUI) for visualization and data saving.

58 Pulse oximetry and skin conductivity are acquired in synchrony with the
59 video recording of the patient's dilation and constriction of the pupil (ac-
60 quired by a commercial system [27]). The system was developed to keep low
61 the production cost and energy consumption. It was tested in experiments
62 from healthy subjects under different stimulations: light, dental occlusion,
63 transcutaneous electrical nerve stimulation (TENS) and computational task.

64 **2. Design and Implementation**

65 The recording system is shown in Figure 1. Two sensors, described below,
66 are developed and are used together with a commercial system for pupil
67 investigation [27].

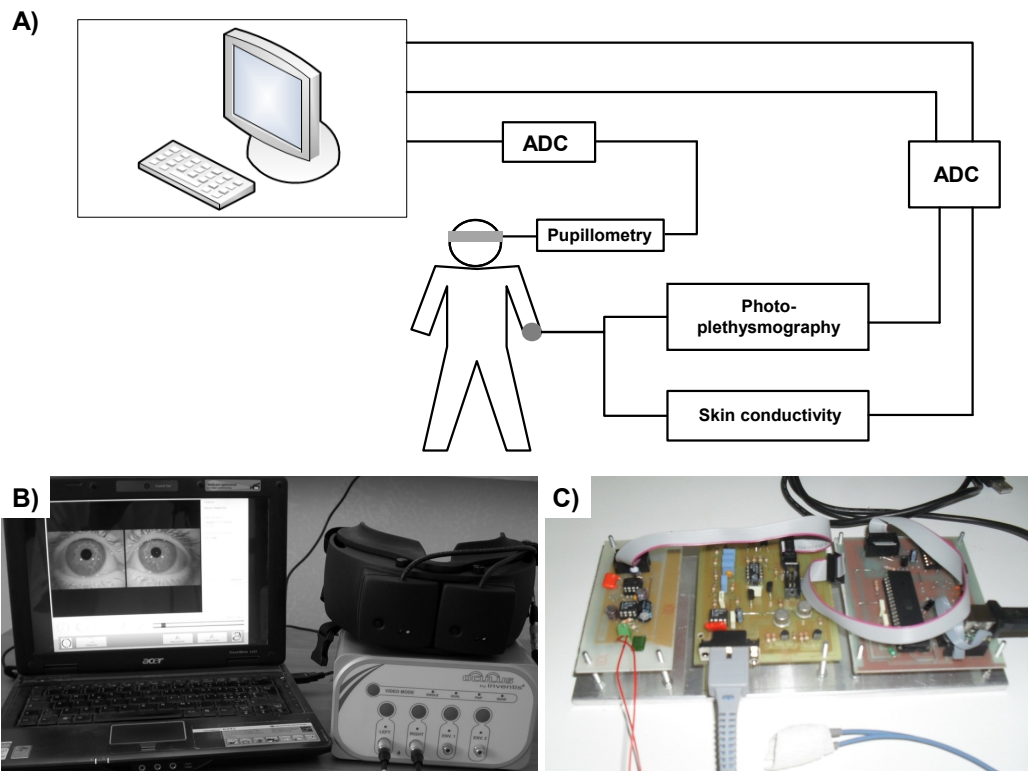


Figure 1: A) General scheme of the instrumentation used. B) Commercial system for pupil investigation [27]. C) Acquisition system recording photoplethysmogram and skin conductivity level.

68 *2.1. Design of the sensors*

69 *2.1.1. Pulse oximeter.*

70 The first sensor measures the absorption of red and infrared lights that
71 pass through a patient's finger by light sensors. The light is generated with
72 2 LEDs that are controlled alternately. A photodiode receives both ambient
73 and modulated light from the LED and generates a current that is related
74 to the oxygen saturation and the cardiac frequency [28][29][30]. The pulse
75 oximetry signal (photoplethysmogram - PPG) has an amplitude of approx-
76 imately 300 nA and a frequency range related to the heart rate (which is
77 about 1 Hz), with maximum frequency contributions of a few Hz [31]. The
78 signal is affected by the line interference (50 Hz), the neon lamps that are
79 usually used in offices and laboratories (giving an interference at 100 Hz)
80 and motion artifacts (notice that high performance professional oximeters
81 are stable to ambient light and motion artifacts [32], but our low cost system
82 had to rely on simple solutions). The circuit design is shown in Figure 2A
83 and B. The current signal is transformed into a voltage signal in the 0.1 V-
84 0.3 V range using a transimpedance amplifier with a gain of 820 kV/A. An
85 initial bandpass filter with cut-off frequencies of 0.8 and 15 Hz was imple-
86 mented; however powerline and neon lamps interferences were still visible in
87 the resulting signal, therefore two notch filters were added to remove their
88 contributions. A final active low-pass filter with a gain $G = 30$ V/V was
89 added to make up for losses in amplitude through the filtering stages, result-
90 ing in a total gain of 5 MV/A. For the actual implementation, the sampling
91 frequency of the PPG is 220 Hz. Notice that this sample rate is very high
92 (e.g., 100 Hz is usually used in clinical settings, but 25 Hz was found to be
93 sufficient to get a reliable estimation of the pulse rate variability [33]): thus,
94 it could be decreased for saving energy or memory.

95 *2.1.2. Skin conductivity level sensor.*

96 There are two skin resistance responses due to the electrodermal activity
97 (EDA): the tonic and the phasic levels. The tonic level is the absolute level of
98 resistance at a given moment in the absence of a measurable phasic response
99 and is referred to as Skin Conductance Level (SCL). The phasic level or
100 Skin Conductance Response (SCR) is superimposed on the tonic level and
101 corresponds to the response to stimuli. The sum of the tonic and phasic
102 response is the Galvanic Skin Response (GSR).

103 The skin conductance is obtained by measuring the current flow through
104 the skin in response to a constant applied voltage. The circuit design is

105 shown in Figure 2C. For bipolar recordings, 0.5 V is recommended [34]. For
 106 a resolution of $0.01 \mu\text{S}$ (the minimum variation of the SCR [35][36]), the
 107 minimum current to be amplified is of 5 nA. The maximum input current of
 108 the amplifier is $15 \mu\text{A}$ (related to the maximum SCR and SCL), therefore
 109 the transimpedance of the circuit must be of 333 k Ω . This circuit has two
 110 independent outputs that allow the analysis of each component separately
 111 (however, their combination, i.e., the GSR, was considered in our tests shown
 112 in the following). The sampling frequency is 170 Hz. Notice that this sample
 113 rate is far beyond the Nyquist limit (even if some literature has suggested
 114 even to further increase it to allow to apply more easily complex analysis or
 115 advanced smoothing procedures [37]), so that it could be decreased if needed
 116 (e.g., to save power consumption or memory storage).

117 Table 1 summarizes the data acquired, the filters used and their frequency
 118 ranges, and the adopted sampling frequencies.

Signal	Filtered Frequency Range	Sampling Frequency
Pulse oximeter	0.8 Hz - 15 Hz Using a band-pass filter and additional 50 Hz and 100 Hz notch filters	220 Hz
Skin Conductivity Level (SCL)	0 - 15 mHz Using a low-pass passive filter	170 Hz
Skin Conductivity Response (SCR)	10 mHz - 1 Hz Using a decoupling capacitor and an active low-pass filter	170 Hz

Table 1: Signals acquired, frequency range of the signals and sampling frequency.

119 2.2. Digital signal processing and transmission

120 The information coming from the analogue sensors was converted into
 121 digital form, acquired, pre-processed and transmitted via USB to a PC. The
 122 microcontroller Atmel ATmega 16 accomplished these tasks [38][39]. Such
 123 a microcontroller was chosen as it is cheap and it has a high speed, a small
 124 code size and an analog multiplier (which can be used to implement digital
 125 filters). The connection of the microcontroller to the system is shown in Fig-
 126 ure 3. Notice that the internal 10-bit ADC was used for sampling the skin
 127 conductance, whereas an external 16 bits analog-to-digital converter (ADC)
 128 was included to sample the PPG (the considered external ADC is AD7715,

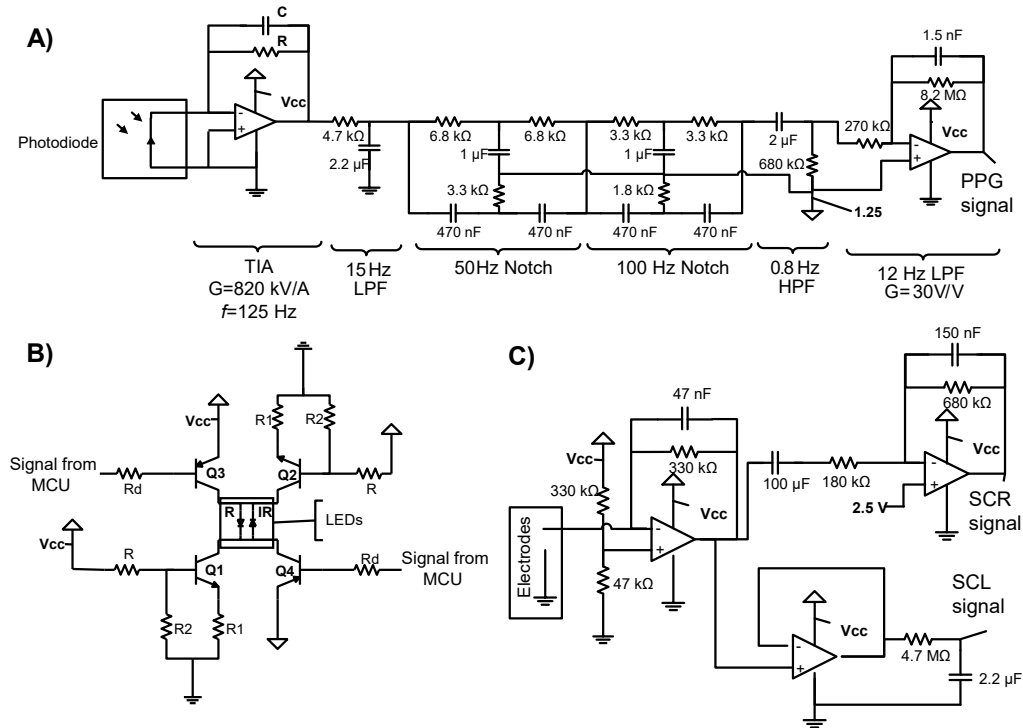


Figure 2: A) Schematic of the photoplethysmograph (abbreviations: PPG - photoplethysmogram; TIA - Transimpedance amplifier; LPF - low-pass filter; HPF - high-pass filter). B) Schematic of the LEDs driver. The signal from the microcontroller unit (MCU) controls which of the 2 LEDs is ON. C) Schematic of the skin conductance sensor. The 2 outputs are for the Skin Conductance Response (SCR) and the Skin Conductance Level (SCL).

129 which is a 16-bit Sigma-Delta ADC that can be interfaced with microcon-
130 trollers using the Serial Peripheral Interface, SPI). The ADC requires up to
131 $195 \mu\text{s}$ for a single conversion, thus setting the sampling frequency of the
132 signals once they have been converted to approximately 5 kHz. To remove
133 noise added by the ADC, an average filter (of 5 samples for pulse oximetry
134 and 20 samples for the Galvanic skin response) was implemented in the mi-
135 crocontroller. The filtered samples were then transmitted to the PC with
136 the USART (Universal Synchronous/Asynchronous Receiver/Transmitter),
137 which was interfaced with a USB transceiver.

138 The red and infrared lights for the pulse oximetry sensor must be ON in
139 different cycles: the signal period was 1 ms, with a duty cycle of 0.25 and
140 the phase between the two lights was 0.5 ms. The state of the LEDs was
141 controlled using timer interrupt.

142 To manage the data, a software interface was developed in Visual Basic,
143 with the following main functions.

- 144 • Registration of patient's information. The user should also be able to
145 import the information from the database of pupillograms.
- 146 • Visualization of the signals. The charts update constantly to show the
147 signals while they are being acquired.
- 148 • Storage of the signals. The information was stored in a file. This file
149 contains the patient's information, the starting time of the acquisition
150 (with a resolution of 10 ms), the time of each sample (recorded as lag
151 from the starting time of acquisition) and the samples of the signals.

152 *2.3. Pupillometry.*

153 Images of the pupils were acquired by the Oculus system (Inventis srl,
154 Padova, Italy), using two infrared CCD cameras (resolution 720x576 pixels,
155 256 grey levels) mounted on a light helmet (1.5 kg), with sampling frequency
156 of 25 frame/s. The eyes were illuminated with an infrared diode with 880
157 nm of wave-length; moreover, during experiments on pupil dynamics under
158 constant light conditions, illumination was provided by a yellow-green LED
159 with 740 nm of wave-length.

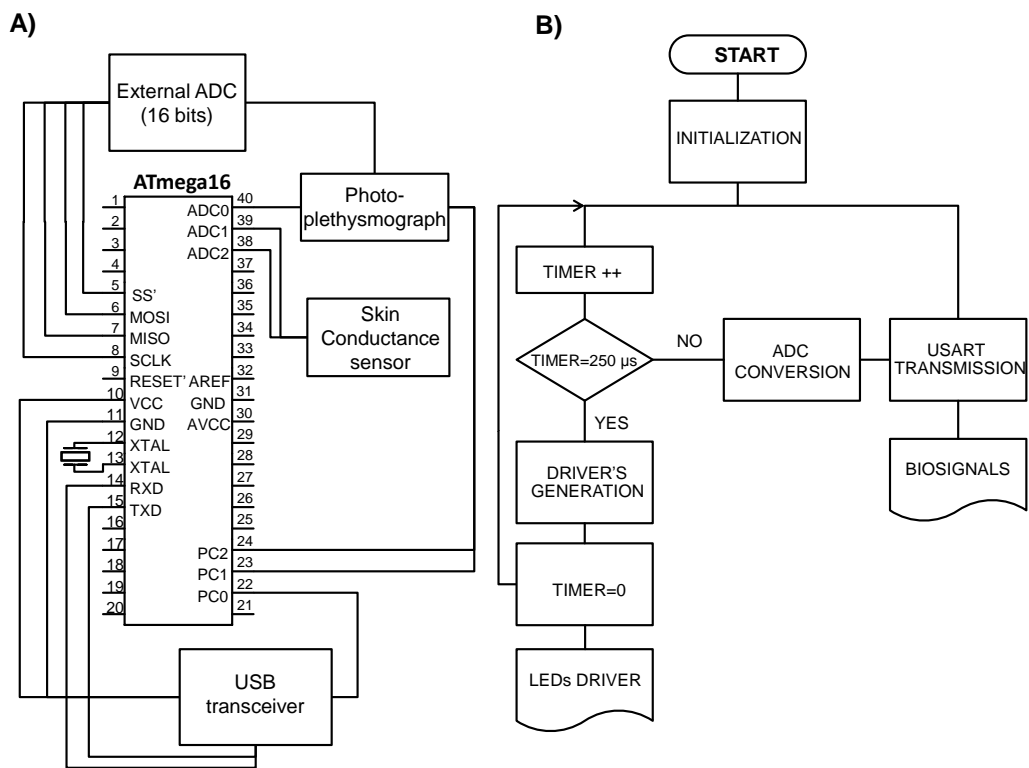


Figure 3: A) General schematic of the digital processing unit. B) Microcontroller general flow diagram.

160 3. Experimental tests

161 In order to assess the reliability of our acquisition system, a pilot study
162 on a few healthy subjects was performed. Specifically, signals were acquired,
163 synchronized and then processed, extracting some descriptors to characterize
164 different experimental conditions.

165 3.1. Signal acquisition

166 Three signals were acquired and then synchronized for subsequent pro-
167 cessing. Specifically, videos of the pupils were acquired by a commercial
168 system (Section 2.3), whereas pulse oximetry and variations of sweat were
169 monitored with the system described in the previous sections, considering
170 the PPG and the GSR time series.

171 3.1.1. Synchronization of the signals.

172 The software of the considered system for pupil investigation [27] does not
173 allow any kind of modifications or access. However, it allows to record the
174 machine time in which each frame is acquired. The data from our prototype
175 were recorded on the same PC. The starting time of the acquisition was
176 saved (as mentioned above), with a resolution of 10 ms. Given that the
177 sampling period of the pupillogram was 40 ms, this resolution was enough to
178 synchronize the two systems.

179 3.1.2. Experimental set up.

180 The subjects were sitting in a high-back chair. The environment was kept
181 at a constant temperature of 21°C. Visual predominance was determined.
182 The acquisition system was then connected to the patient's non dominant
183 hand. The helmet was applied and was maintained until the end of the
184 recording. This phase took about 4 minutes.

185 The correct procedure and execution of tests was first explained to the
186 subjects. Then, they were asked for brief tests to make sure that the instruc-
187 tions were well understood. This phase took about 2 minutes.

188 Two operators worked within the experimental set. The first took care
189 of the subject (pretest and test instructions, helmet handling, check of the
190 correctness of execution), the second controlled hardware and software.

191 *3.1.3. Experiments.*

192 A few experimental tests were performed on 8 young and healthy subjects
193 (age 25.1 ± 1.1 years; 6 females, 2 males). Small stimulations were admin-
194 istered to the participants, to test the ability of our prototype to register
195 reliable data characterizing the response of the ANS. Specifically, stationary
196 conditions of one minute duration which require a different involvement of
197 the sympathetic and parasympathetic controls were considered, separated by
198 5 minutes rest. The following conditions were investigated: neutral position
199 of the jaw (rest position: RP) and habitual dental occlusion (HDO)¹, in light
200 or darkness condition; moreover, a test was performed during a computa-
201 tional task, which was assumed to induce a detectable stress of the subject.
202 These 5 conditions (tested in the following order: RP in light and dark-
203 ness, HDO in light and darkness and computational task in darkness) were
204 performed before, during and after the application of low-frequency Transcu-
205 taneous Electrical Nerve Stimulation (TENS), which was expected to induce
206 relaxation. TENS was applied as in previous studies [40], using a J5 My-
207 omonitor TENS Unit device (Myotronics-Noromed, Inc., Tukwila, WA, USA)
208 with disposable electrodes (Myotrode SG Electrodes, Myotronics-Noromed,
209 Inc., Tukwila, WA, USA). Synchronous and bilateral stimuli, with amplitude
210 adjusted in the range 0–24 mA and duration of 500 μ s, have been delivered
211 over the cutaneous projection of the notch of the fifth pair of cranial nerves
212 (grounding electrode in the center of the back of the neck) at 0.66 Hz.

213 *3.2. Signal processing and results*

214 *3.2.1. Pre-processing of recorded data.*

215 Pupillometric recordings were processed through the algorithm of strongly
216 connected components [41] to measure frame by frame the area of the pupil,
217 expressed as the number of pixels covering it. The area of the pupil was then
218 low-pass filtered under 2 Hz (non-causal, zero-phase, Butterworth filter of
219 order 2). This frequency range is enough to characterize the response of the
220 ANS of our interest, which mainly includes the following frequency ranges:
221 low-frequency (LF) 0.04–0.15 Hz and high-frequency (HF) 0.15–0.5 Hz [9].
222 The local maxima in the PPG were used to identify the heartbeats, from

¹During dental occlusion, the effect of muscle fatigue and the massive involvement of the autonomic system were excluded by avoiding prolonged teeth clenching. Subjects were asked to swallow and then to contact the teeth lightly without clenching. Attention was paid to check the activity of mimic muscles.

223 which the heart rate (HR) was estimated as the reciprocal of the inter-pulse
224 interval.

225 The GSR was low-pass filtered under 2 Hz.

226 3.2.2. Estimation of indexes.

227 The following simple descriptors were estimated from the pre-processed
228 data (pupillogram, i.e., pupil size over time), HR and GSR.

- 229 • The mean and standard deviation (STD) were computed to explore
230 average values and variability, respectively.
- 231 • The linear trend was estimated as a basic indicator of the evolution in
232 time of the signals (the trend was defined as the slope of the interpo-
233 lation line of the data after scaling the time to range between 0 and 1
234 and normalizing the time series to have zero mean and unit STD).

235 3.2.3. Statistical analysis.

236 The indexes mentioned above were used as descriptors of the signals in
237 the different experimental conditions. The two-sided Wilcoxon signed rank
238 test (considering paired data) was then applied to investigate differences of
239 each of the indexes in specific pairs of conditions of interest, after pooling
240 data: RP in darkness compared to the computational task, RP compared to
241 HDO (in darkness), pre-TENS compared to TENS or post-TENS conditions.
242 The significance level was set to $p < 0.05$.

243 3.2.4. Results.

244 Examples of recorded signals are shown in Figure 4. The pupil shows an
245 irregular oscillatory behavior, as also the HR. There is a decreasing trend
246 in both HR and GSR. The figure shows also the spectral coherence of the
247 pupillogram and the HR at frequencies lower than 1 Hz. The two signals were
248 found to be coherent in subjects under controlled breathing conditions, where
249 a respiratory component was visible in both the pupillogram and the HR [8].
250 Here, the considered normal breathing and the short acquisitions resulted in
251 significant coherence (over 0.5) only in a few subjects and conditions.

252 The significance of the differences of indexes extracted from the signals
253 recorded in different conditions is shown in Table 2. Different indexes have
254 a greater discriminatory value comparing different conditions:

- 255 • an index from the HR showed the maximal significance (i.e., minimum p
256 value) in discriminating computational task and RP in darkness (which
257 can be considered as a rest state);

- 258 • indexes estimated from the pupillogram had the maximal significance
259 in distinguishing light and darkness, RP and HDO, or pre-TENS and
260 TENS conditions;
- 261 • indexes extracted from GSR were the most statistically different in the
262 conditions pre-TENS versus post-TENS and TENS versus post-TENS.

263 Specifically, HR and mean pupil size increased when comparing computa-
264 tional task with RP in darkness (due to the mental stress induced by the
265 task). The subject started sweating during the computational task, as indi-
266 cated by the high positive trend of GSR. Pupil was the only system showing
267 significant differences between HDO and RP: pupil size increased as a result
268 of HDO stimulation [9][10][11]. Moreover, it was the only system showing
269 significant differences comparing light and darkness conditions (obviously,
270 increasing the diameter in darkness).

271 Pupil also indicated the relaxation induced by TENS (there is a significant
272 reduction of pupil size during and after the application of TENS). On the
273 other hand, GSR increased during and after TENS, with respect to the pre-
274 TENS condition.

275 4. Discussion

276 The ANS controls many different visceral functions, including heart rate,
277 perspiration, digestion, salivation, respiratory rate, pupillary dynamics, mic-
278 turation (urination), sexual arousal, breathing and swallowing. The joint
279 acquisition of different autonomic responses could be useful for a deeper in-
280 sight into ANS physiology and pathology. For example, the study of the
281 autonomic response is important in the following situations [42]:

- 282 • sympathetic and parasympathetic lesions after surgical procedures;
- 283 • drug's collateral effects;
- 284 • diagnosis and follow-up of ANS diseases;
- 285 • poisoning;
- 286 • involuntary reactions of the patient.

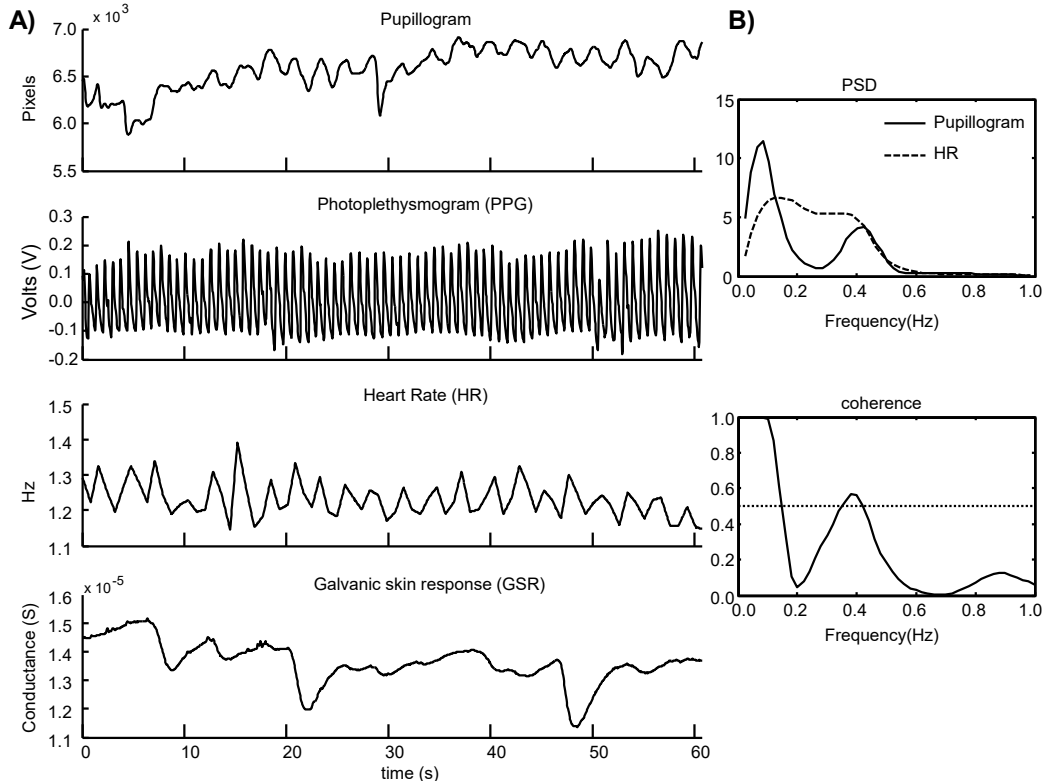


Figure 4: A) Example of data (pupillogram, PPG from which the HR is obtained, GSR), with the subject at rest position of the jaw in darkness. The HR was computed as the reciprocal of the inter-pulse interval. Each HR value was located in the average time between the two instants used to compute it, obtaining a time series (sampled not uniformly). Then, this time series, PPG and GRS were interpolated at 25 Hz (i.e., the sampling frequency of the pupillogram). B) Power spectrum densities (PSD) of pupillogram and HR (using Welch's overlapped segment averaging estimator, considering 8 segments with 50% overlap) and magnitude squared coherence.

287 The diagnosis in most of these situations is currently vague [43][44][45].
288 The joint investigation of different autonomic responses could help in clar-
289 ifying the complex dynamics of the ANS in such conditions. One of the
290 problems when designing an experimental setup is being constrained by the
291 functionalities of commercial systems, which are usually closed and allow only
292 specific protocols. Here, we were interested in investigating synchronously
293 the joint responses of the heart (i.e., the cardiac pulse), the skin (i.e., sweat
294 production) and pupil. Some devices have been developed for the joint ac-
295 quisition of different ANS responses: both research prototypes [46][47][48]
296 and commercial products (Q sensor, MOXO, FeelTM, UP4, VIVOGRAPH,
297 Microsoft Band, E4, Embrace) have been proposed. Despite the interest in
298 the study of pupillary dynamics, at present there are no tools that are able to
299 simultaneously obtain the acquisition and processing of pupillary dynamics
300 and those of other systems such as the cardiovascular (e.g., through pulse
301 oximetry) and sudorific system (through skin conductivity). Being able to
302 expand the analysis to several systems in a synchronous way would allow
303 a better knowledge of the coupling pathways between somatosensory and
304 affective / emotional needs and responses of the whole of the ANS in physio-
305 logical and/or pathological conditions, probably indicating the activation of
306 different autonomous circuits in different conditions.

307 In this work, we have designed an experimental setup able to record in
308 synchrony the video of pupil (done with a commercial system [27]) and dif-
309 ferent responses of the ANS. A modular sensor network was designed and
310 implemented to acquire, process and transmit via USB some biomedical sig-
311 nals reflecting the state of the ANS. Two sensors have been included in the
312 prototype, for pulse oximetry and skin conductivity. However, it is feasible
313 for being extended to include more body sensors, miniaturized and embedded
314 in a portable system. This could be important for future extension of the
315 work, as body sensor networks are finding many applications in the continu-
316 ous monitoring of sensitive people [49][50][51][52][53]. Indeed, many different
317 sensors are available to monitor physiological data and could be included:
318 accelerometer, blood glucose sensor, electrodes for bioelectric signals, blood
319 pressure sensor, gyroscope, carbon dioxide gas sensor, etc. [53][54].

320 The developed system, even if it is only a prototype, provided robust estima-
321 tions of the electrodermal activity and of the pulse oximetry, which was then
322 processed to investigate the heart inter-beat interval. Note that the PPG
323 is immune to electrical artifacts, which could be observed on the electrocar-
324 diogram during TENS application (however, it is less precise in detecting

325 the HR, as it also depends on the pulse wave velocity). Thus, our system is
326 adequate for the study of the ANS response to TENS reflected in the HR.

327 Preliminary experimental tests are shown. Some trends have been ob-
328 served. For example, both HR and GSR are decreasing during the experi-
329 ment shown in Figure 4. This could reflect that the subject was relaxing, as
330 the recordings were acquired in a rest condition (i.e., rest position of the jaw
331 in darkness). Moreover, GSR slowly increased during the experiments, with
332 the results that its mean value was larger during and after TENS, with re-
333 spect to the pre-TENS condition, even if TENS should relax the participants
334 (as indicated by variations in pupil size). Possibly, this trend of GSR was
335 only due to an accumulation of sweat during the experiment, as the sensors
336 were kept fixed for all its duration.

337 However, some consistent outcomes were also observed. Specifically, even
338 if we considered only short recordings, weak ANS stimulations (dental oc-
339 clusion, light, TENS and a computational task) in healthy subjects and we
340 extracted simple indexes (mean, STD and trend), we have shown statisti-
341 cally significant variations of at least an index in each pair of conditions.
342 This indicates that the overall information provided by the joint recordings,
343 not just that of each individual signal, should be used for the discrimination
344 of the ANS responses in the different considered conditions. The results are
345 in line with our expectations: HDO and computation elicit the sympathetic
346 response (which should result in HR and GSR increasing and pupil dilation),
347 TENS induces relaxation (determining a decrease of HR, GSR and pupil
348 size). The sensory amplitude TENS that we administered has been shown
349 to induce the response of different branches of the ANS, as the salivary [55],
350 the cardiovascular [56], the neuromuscular systems [57] and, in particular,
351 the one that governs the static and dynamic dimension of the pupil [40]. The
352 effects of the sensory trigeminal TENS stimulation are probably to be at-
353 tributed to a central system that modulates the mechanisms of arousal and,
354 therefore, the autonomous efferent response [58][59] (that was observed in
355 this study).

356 Pupil appeared to be the most sensible system, as it reflected even the
357 small stimuli given by HDO or TENS. HR showed significant variations only
358 in a few conditions (e.g., mean HR was significantly increased only by the
359 computation task). GSR showed an important trend during computation,
360 but it was prone to accumulation effects.

361 Our study has some limitations: a few experiments have been conducted
362 and all on healthy volunteers; only three signals have been included; our

363 system for the recording of PPG and GSR is a simple prototype, which is
364 surely more delicate and less stable than engineered commercial products.
365 However, it pointed out the importance of integrating information from dif-
366 ferent systems controlled by the ANS, in particular pupil (which was the most
367 sensitive to weak stimulations). Further studies are then suggested, both to
368 improve and extend our preliminary results. For example, other tests and
369 advanced processing techniques could provide specific indications in physi-
370 ology or pathology, in future joint acquisitions of different responses of the
371 ANS. Additional measures to guarantee reproducibility of the tests need to
372 be developed, such as careful preparation in order to stabilize hemodynamic
373 parameters. Moreover, additional sensors or stimulation signals could be
374 included in the proposed device, due to its modular architecture.

375 5. Conclusions

376 We discuss the implementation of a modular system for the acquisition
377 of different signals reflecting the responses of the ANS to stimulations. The
378 signals were synchronized to the pupillogram recorded by a commercial sys-
379 tem, showing the joint variations of pupil size, Galvanic response and pho-
380 toplethysmogram. Small stimulations were given to the participants and
381 only considering all different recorded signals the different responses of the
382 ANS could be discriminated. Thus, these preliminary results suggest the
383 importance of recording jointly multiple data in order to better characterize
384 autonomic responses.

385 References

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CONDITIONS	INDEXES with SIGNIFICANT DIFFERENCE
Computation versus RP (D)	<i>Mean HR</i> (p=0.0002) Comp.: 1.4(1.17, 2.04) – RP: 1.08(0.94, 1.57) <i>STD of HR</i> (p=0.001) Comp.: 0.28(0.17, 0.54) – RP: 0.11(0.08, 0.38) <i>STD of GSR</i> (p=0.0015) Comp.: 1.25(0.54, 2.77) – RP: 0.2(0.1, 1.0) <i>Trend of GSR</i> (p=0.0078) Comp.: 3.03(0.27, 3.28) – RP: –0.12(–1.88, 2.06) <i>Mean pupil area</i> (p=0.01) Comp.: 6952(6180, 9050) – RP: 6565(5839, 8072)
HDO (D) versus RP (D)	<i>Mean pupil area</i> (p=0.0029) HDO: 6817(6318, 8636) – RP: 6565(5839, 8072) <i>Trend of pupil area</i> (p=0.012) HDO: 1.05(–0.62, 1.86) – RP: 2.10(1.55, 2.63) <i>STD of pupil area</i> (p=0.024) HDO: 177(142, 279) – RP: 267(202, 308)
pre-TENS versus TENS	<i>Mean of pupil area</i> (p=0.009) pre: 6425(3806, 7391) – tens: 5818(3134, 7082) <i>STD of GSR</i> (p=0.031) pre: 0.11(0.05, 0.27) – tens: 0.21(0.72, 0.83) <i>STD of HR</i> (p=0.036) pre: 0.11(0.07, 0.24) – tens: 0.12(0.09, 0.57)
TENS versus post-TENS	<i>Mean of GSR</i> (p=0.0001) tens: 12.91(9.09, 14.83) – post: 13.65(12.86, 16.61)
pre-TENS versus post-TENS	<i>Mean of GSR</i> (p=0.006) pre: 12.88(10.88, 13.68) – post: 13.65(12.73, 16.61) <i>Mean area of pupil</i> (p=0.012) pre: 6425(3806, 7392) – post: 5823(3151, 6874) <i>STD of HR</i> (p=0.022) pre: 0.12(0.08, 0.25) – post: 0.16(0.08, 0.35)
Light versus Darkness	<i>Mean area of pupil</i> (p<<0.001) L: 2987(2545, 3981) – D: 7312(6613, 9059) <i>STD of pupil</i> (p<<0.001) L: 417(333, 539) – D: 220(161, 283) <i>Trend of pupil</i> (p=0.014) L: 0.23(–0.79, 1.29) – D: 1.70(0.21, 2.32)

Table 2: Statistical analysis of the data using Wilcoxon sign rank test. Mean, standard deviation and trend of data were considered (L: light; D: darkness). Median and quartiles of the indexes showing significant differences (p<0.05) are reported (in order of increasing p). Pupil size is indicated in pixels, HR in Hz, GSR in μ S; their trends are measured in arbitrary units (they were computed on normalized data and time).