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*Original*

A Cellular Potts Model of single cell migration in presence of durotaxis / Rachele, Allena; Scianna, Marco; Preziosi, Luigi.  
- In: MATHEMATICAL BIOSCIENCES. - ISSN 0025-5564. - 275:(2016), pp. 57-70. [10.1016/j.mbs.2016.02.011]

*Availability:*

This version is available at: 11583/2649594 since: 2020-02-18T22:19:47Z

*Publisher:*

Elsevier Inc.

*Published*

DOI:10.1016/j.mbs.2016.02.011

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# A Cellular Potts Model of single cell migration in presence of durotaxis

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**Abstract**\_ Cell migration is a fundamental biological phenomenon during which cells sense their surroundings and respond to different types of signals. In presence of durotaxis, cells preferentially crawl from soft to stiff substrates by reorganizing their cytoskeleton from an isotropic to an anisotropic distribution of actin filaments. In the present paper, we propose a Cellular Potts Model to simulate single cell migration over flat substrates with variable stiffness. We have tested five configurations: i) a substrate including a soft and a stiff region, ii) a soft substrate including two parallel stiff stripes, iii) a substrate made of successive stripes with increasing stiffness to create a gradient and iv) a stiff substrate with four embedded soft squares. For each simulation, we have evaluated the morphology of the cell, the distance covered, the spreading area and the migration speed. We have then compared the numerical results to specific experimental observations showing a consistent agreement.

**Keywords**\_ Cell migration, Durotaxis, Cell polarity, Anisotropy, CPM

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## 1. Introduction

Cell migration is a critical phenomenon occurring in several biological processes, such as morphogenesis [1], wound healing [2] and tumorigenesis [3]. It takes place in successive and cyclic steps [4] and it is triggered by specific interactions with the ExtraCellular Matrix (ECM). Actually, cell migration may occur in the absence of external signals thereby typically resulting in a random

37 walk. However, in most situations, cells are able to sense their surrounding  
38 environment and to respond for instance to chemical (i.e., chemotaxis) [5],  
39 electrical (i.e., electrotaxis) [6] or mechanical (i.e., mechanotaxis) [7] fields or yet  
40 to stiffness gradients (i.e., durotaxis) [8,9]. The latter mechanism consists of the  
41 cell preferential crawling from soft matrix substrates to stiffer ones, even in the  
42 absence of any additional directional cues [10,11]. By forming local protrusions  
43 (i.e., pseudopodia), the cells are in fact able to probe the mechanical properties of  
44 the surrounding environment and to more strongly adhere over stiff regions.  
45 Additionally, such behaviour results in a substantial reorganisation of the  
46 intracellular cytoskeleton. In fact, over soft substrates cells typically show an  
47 unstable and isotropic distribution of actin filaments, which are poorly extended  
48 and radially oriented, whereas over stiff substrates cell morphology is more stable  
49 and exhibits significant spreading and often anisotropic arrangements of actin  
50 filaments in the direction of migration (i.e., polarization) [12–16].

51 Although several computational models have been proposed in literature to  
52 investigate single cell migration, only few of them deal with durotaxis. Among  
53 others, it is worth to cite the work by Moreo et al. [17] who proposed a continuum  
54 approach based on an extension of the Hill's model for skeletal muscle behaviour  
55 to investigate cell response on two-dimensional (2D) substrates. They showed, in  
56 agreement with experimental observations, that cells seem to have the same  
57 behaviour when crawling on stiffer substrate and on pre-strained substrates.  
58 Harland et al. [18] instead represented a cell as a collection of stress fibres  
59 undergoing contraction and birth/death processes and showed that on stiff  
60 substrates cells exhibit durotaxis and stress fibres significantly elongate.  
61 Dokukina and Gracheva [19] developed a 2D discrete model of a viscoelastic

62 fibroblast cell using a Delaunay triangulation. At each node the balance of the  
63 forces was determined by the contributions i) of the frictions between the cell and  
64 the substrate, ii) of a passive viscoelastic force and iii) of an intrinsic active force.  
65 The authors then evaluated cell behaviour over a substrate with a rigidity step in  
66 good agreement with specific experimental observations. In fact, they found that  
67 the cell preferentially moves on the stiffer substrate and turns away from the soft  
68 substrate as reported by [8]. Stefanoni et al. [20] proposed a finite element  
69 approach able to account for the local mechanical properties of the underneath  
70 substrate and to analyze selected cell migratory determinants on two distinct  
71 configurations: an isotropic substrate and a biphasic substrate (which consists of  
72 two adjacent isotropic regions with different mechanical properties). Trichet et al.  
73 [14] employed instead the active gel theory to demonstrate that cells preferentially  
74 migrate over stiff substrates founding an optimal range of rigidity for a maximal  
75 efficiency of cell migration. Further, in [21] a vertex-based approach (i.e., the so-  
76 called Subcellular Element Model, SCE) was set to represent intracellular  
77 cytoskeletal elements as well as their mechanical properties. In particular, the  
78 dynamics of such subcellular domains were described by Langevin equations,  
79 which account for a weak stochastic component (i.e., that mimic cytoplasmic  
80 fluctuations) and elastic responses (i.e., modelled by generalized Morse  
81 potentials) to both intracellular and intercellular biomechanical forces. The same  
82 method was successfully applied in [22] for modelling substrate-driven bacteria  
83 locomotion. Finally, in Allena and Aubry [23] a 2D mechanical model was  
84 proposed to simulate cell migration over an heterogeneous substrate including  
85 slipping regions and to show that over softer regions the cell slows down and is  
86 less efficient.

87           In the present work, we describe a Cellular Potts Model (CPM, developed in  
88 [24,25] and reviewed in [25–29]), which is a lattice-based stochastic approach  
89 employing an energy minimization philosophy, to reproduce single cell migration  
90 over flat substrates with different rigidity. In particular, we test four  
91 configurations: i) a substrate including a soft and a stiff region, ii) a soft substrate  
92 including two parallel stiff stripes, iii) a substrate made of successive stripes with  
93 increasing stiffness to create a gradient and iv) a stiff substrate with four  
94 embedded soft squares. For each scenario, we analyse cell behaviour in terms of  
95 morphology, distance covered, spreading/adhesive area and migration speed in  
96 order to capture the essential mechanisms of durotaxis. The computational  
97 outcomes are then compared with specific experimental observations taken from  
98 the existing literature.

99           The rest of this paper is organized as follows. In Section 2, we clarify the  
100 assumptions on which our approach is based and present the model components.  
101 The simulation results are then shown in Section 3. Finally, a justification of our  
102 model choices as well as a discussion on possible improvements is proposed in  
103 Section 4. Additionally, the article is equipped with an Appendix that deals with  
104 statistics and parameter estimates.

105

## 106           **2. Mathematical model**

107           The cell-substrate system is represented using a CPM environment [24,25].  
108 The simulation domain is a three-dimensional (3D) regular lattice  $W \in \mathbb{R}^3$   
109 constituted by identical closed grid sites, which are identified by their centre  
110  $\mathbf{x} \in \mathbb{R}^3$  and labelled by an integer number  $S(\mathbf{x}) \in N$  (which can be interpreted as  
111 a degenerate spin) [30,31]. The boundary of a generic site  $\mathbf{x}$ , one of its neighbours

112 and its overall neighbourhood are defined as  $\mathfrak{N}(\mathbf{x}, \mathbf{x}')$  and  $W'_{\mathbf{x}}$ , respectively.  
 113 Subdomains with identical label  $S$  form discrete objects  $S_s$  (with border  $\partial\Sigma_\sigma$ ),  
 114 which have an associated type  $t(S_s)$ . In the case of our interest,  $t = M$  stands  
 115 for the medium,  $t = C$  for the cells and  $t = S_i$  for the  $i$ -th type of substrate. In this  
 116 respect, we anticipate that each type of matrix region will differ for stiffness and  
 117 therefore for adhesive affinity with moving individuals.

118 Cell dynamics result from an iterative and stochastic reduction of the energy of  
 119 the overall system, given by a Hamiltonian  $H$  (units:  $\text{Kg}\cdot\text{m}^2/\text{s}^2$ ), whose expression  
 120 will be clarified below. The employed algorithm is a modification of the  
 121 Metropolis method for Monte Carlo-Boltzmann dynamics [24,32], which is  
 122 particularly suitable to simulate the exploratory behaviour of biological  
 123 individuals as cells. Procedurally, at each time step  $t$  of the algorithm, called  
 124 Monte Carlo Step (MCS), a randomly chosen lattice site  $\mathbf{x}_{source}$  belonging to a cell  
 125 tries to allocate its spin  $\sigma(\mathbf{x}_{source})$  to one of its unlike neighbours  $\mathbf{x}_{target} \in W'_{\mathbf{x}}$ ,  
 126 which is also randomly selected. Then, the net energy difference  $DH$  due to the  
 127 proposed change of system configuration is calculated as

$$128 \quad DH \Big|_{S(\mathbf{x}_{source}) \rightarrow S(\mathbf{x}_{target})} = H_{(after\ spin\ copy)} - H_{(before\ spin\ copy)} \quad (1)$$

129 The trial spin update is finally validated by a Boltzmann-like probability function  
 130 defined as

$$131 \quad P[S(\mathbf{x}_{source}) \rightarrow S(\mathbf{x}_{target})](t) = \min \left\{ 1, e^{-\frac{DH}{T_c}} \right\} \quad (2)$$

132 where  $t$  is the actual MCS and  $T_c \hat{=} R_+$  is a Boltzmann temperature, that has been  
 133 interpreted in several ways by CPM authors (see [33] for a comment on this

134 aspect). However, we here opt to give  $T_C$  the sense of a cell intrinsic motility (i.e.,  
 135 agitation rate), following the approach in [25]. Finally, it is useful to underline  
 136 that the matrix substrates are considered fixed and immutable.

137 As seen, the simulated system evolves to iteratively and stochastically reduce  
 138 its free energy, which is defined by a Hamiltonian function  $H$  which, for any  
 139 given time step  $t$ , reads

$$140 \quad H(t) = H_{adhesion}(t) + H_{shape}(t) \quad (3)$$

141  $H_{adhesion}(t)$  is deduced from the Steinberg's Differential Adhesion Hypothesis  
 142 (DAH) [24,34] and is due to the adhesion between cells and extracellular  
 143 components (i.e., the medium or a given type of substrate). In particular, it reads

$$144 \quad H_{adhesion}(t) = H_{adhesion}(t) = \frac{\dot{a}}{(\|\mathbf{x}_i\| \|\mathbf{S}_s\|) \zeta (\|\mathbf{x}_i\| \|\mathbf{S}_{st}\|)} \mathbf{J}_{t(S_s(\mathbf{x})), t(S_{st}(\mathbf{x}))} \quad (4)$$

145 with  $\mathbf{x}$  and  $\mathbf{x}^c$  two neighbouring sites and  $S_s$  and  $S_{st}$  two neighbouring objects  
 146 (with borders  $\|\mathbf{S}_s$  and  $\|\mathbf{S}_{st}$ , respectively).  $\mathbf{J}_{t(S_s(\mathbf{x})), t(S_{st}(\mathbf{x}))}$  and  $\hat{\Gamma} R_{\mp}$  are constant and  
 147 homogeneous binding forces per unit area. They are symmetric with respect to  
 148 their indices and can be specified as follows:

- 149 -  $J_{C,M}$  is the adhesive strength between the cells and the collagenous medium  
 150 which is constituted by a mixture of soluble adhesive ligands (i.e., carbohydrate  
 151 polymers and non-proteoglycan polysaccharides) and water solvent;
- 152 -  $J_{C,S_i}$  gives the adhesive strength between the cells and  $i$ -th type of substrate.

153 Recalling the minimization theory of the CPM, we assume that the stiffer the  
 154 substrate  $i$ , the lower the corresponding value  $J_{C,S_i}$  (i.e., the higher the adhesion  
 155 between the cells and the  $i$ -th type of substrate). This is a pivotal hypothesis of our  
 156 approach: it is consistent since it has been widely demonstrated in the

157 experimental literature that cells generate higher traction forces and generate more  
 158 stable focal adhesion points when migrating over stiffer substrates [16,35–38].

159  $H_{shape}(t)$  defines the geometrical attributes of each cell  $S_s$ , which are written  
 160 as elastic potentials as it follows:

$$\begin{aligned}
 H_{shape}(t) &= H_{volume}(t) + H_{surface}(t) \\
 &= \frac{1}{2} k_{S_s} (v_{S_s}(t) - V_C)^2 + n_{S_s}(t) (s_{S_s}(t) - S_C)^2 \quad (5)
 \end{aligned}$$

162 where  $v_{S_s}(t)$  and  $s_{S_s}(t)$  are the actual volume and surface of the cell  $S_s$ ,

163 whereas  $V_C$  and  $S_C$  the corresponding cell characteristic measures in the initial

164 resting condition.  $k_{S_s}$  and  $n_{S_s}(t)$  are instead two mechanical moduli in units of

165 energy. The former is linked to volume changes and, assuming that cells do not

166 significantly grow during migration, is considered constant with a high value (i.e.,

167  $k_{S_s} = k_C \gg 1$ ) for any individual  $S_s$ . The latter refers to the rigidity of a cell. As

168 we will explain in details later on, for each cell  $S_s$ ,  $n_{S_s}$  is assumed to depend on

169 the underneath type of substrate. In particular, each cell decreases its initially high

170 (i.e.,  $\gg 1$ ) rigidity, thereby being more able to deform, if it comes in contact with

171 a stiff substrate. This assumption is consistent with experimental observations on

172 the fact that cell contact with stiff matrix regions activate downstream

173 intracellular pathways resulting in acto-myosin dynamics and therefore in

174 cytoskeletal remodelling [8,39]. More specifically, it seems that certain cells have

175 a binary sensor at their membrane junction sites that allows them to switch from a

176 relaxed and rounded morphology, when the substrate is softer than the cell's

177 elastic modulus [39–43], to a fan-shaped morphology with abundant stress fibres,

178 when the substrate is stiffer or as stiff as the cell itself [39]. Further, it has been



179 shown that cells tend to isotropically and poorly spread on soft substrates,  
180 whereas they form pseudopodia randomly distributed along the membrane on stiff  
181 substrates, resulting in a significant anisotropic spreading [16]. In this respect,  
182 according to several experimental observations [16,35–38,44], there exists a linear  
183 relationship between the adhesion forces exerted by the cell on the substrate and  
184 the spreading area of the cell. More specifically, the larger the contact area  
185 between the cell and the substrate, the higher the number of focal adhesion points  
186 that can be established. Nonetheless, the sequence of events is still unclear and  
187 two main processes may occur when a cell is seeded on a stiff substrate [45]:

- 188 i) the cell adheres because of the stiffness of the substrate, then it significantly  
189 spreads;
- 190 ii) the cell spreads because of the stiffness of the substrate, then it more strongly  
191 adheres.

192 Such uncertainty is the reason why in the present model both the adhesive  
193 parameters and the cell rigidity directly depend on the substrate stiffness, but are  
194 independent from each other.

195 The main components and the scales involved in the proposed model are  
196 summarized in the diagram in Fig. 1. Finally, all the parameters of the simulations  
197 are reported in Table 1, while the Appendix provides a careful explanation of how  
198 they have been estimated.

199

### 200 **3. Numerical simulations**

201 The characteristic size of each lattice site is  $4\ \mu\text{m}$  and the geometrical domain  
202  $W$  is a  $70 \times 70 \times 30$  regular grid ( $280\ \mu\text{m} \times 280\ \mu\text{m} \times 120\ \mu\text{m}$ ) with no-flux  
203 boundary conditions in all directions. This choice mimics the situation of a

204 delimited experimental device, where cells are not able to overcome the physical  
 205 barriers. All our CPM cells are initially a hemisphere of a radius of 20  $\mu\text{m}$ , whose  
 206 initial position will be specified for each simulation setting. A MCS is set to  
 207 correspond to 2 s of actual unit of time (see the Appendix for a comment on this  
 208 aspect), which results in simulations covering time intervals between 16 min to  
 209 5.5 h. This choice enables cells to migrate over sufficiently long paths in order to  
 210 compare numerical results and proper experimental observations. We have tested  
 211 several cell-matrix settings, which are presented in the followings. The resulting  
 212 simulations were performed on a modified version of the open source package  
 213 CompuCell3D (downloadable at [www.compuCell3d.org](http://www.compuCell3d.org)). In particular, a Python  
 214 script was specifically developed to account for substrate-dependent cell rigidity.

215

### 216 *3.1 Cells preferentially crawl over stiff substrates*

217 We first consider a substrate split into a soft and a stiff region, i.e.,  $t = \mathcal{S}_1$  and  
 218  $t = \mathcal{S}_2$  (see Fig. 2a). A cell  $S_1$  is then seeded at the centre of the substrate and it  
 219 is allowed to move for 500 MCS (approximately 16 minutes). The rigidity of  $S_1$ ,  
 220  $n_{S_1}$ , has initially a high value  $v_{\Sigma_1} = v_C = 25 \cdot 10^{-3} \text{ Kg/s}^2\text{m}^2$ . However, it is allowed  
 221 to decrease, of  $10^{-3} \text{ Kg/s}^2\text{m}^2$  for MCS until a threshold value  $n_t$  equal to  $10^{-2}$   
 222  $\text{Kg/s}^2\text{m}^2$ , while the cell is in contact with the stiff region  $S_2$ , thereby leading to a  
 223 flattening of the initially rigid cellular hemisphere. In mathematical terms, we  
 224 indeed have that

$$\begin{aligned}
 & v_{\Sigma_1}(t) \\
 &= \begin{cases} \max(v_{\Sigma_1}(t-1) - 10^{-3}; v_t) & \text{if } \exists(\mathbf{x}, \mathbf{x}' \in \Omega'_x): \mathbf{x} \in \Sigma_1 \text{ and } \mathbf{x}' \in S_2; \\ v_{\Sigma_1}(t-1) & \text{else,} \end{cases} \quad (6)
 \end{aligned}$$

225 for each MCS.

226 We then study how cell behaviour is affected by variations in the ratio between  
227 the adhesive affinity of the cell with either the soft or the stiff substrate region. In  
228 particular, we keep fixed  $J_{C,S_1} = J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$  while decreasing the  
229 value of  $J_{C,S_2}$  from  $25 \cdot 10^{-15} \text{ Kg/s}^2$  to  $1 \cdot 10^{-15} \text{ Kg/s}^2$  (which is equal to  $J_{stiff}$ ,  
230 the lowest value consistent with the case of our interest, see the Appendix). As  
231 summarized in Fig. 3c, when  $J_{C,S_2}$  decreases, the cell is biased to crawl towards  
232 the stiff domain, as it is confirmed by the plot of the trajectories of its center of  
233 mass deriving from independent simulations. In fact, over a period of 500 MCS ( $\approx$   
234 16 minutes), the cell randomly moves around the substrate centre when  $\frac{J_{C,S_1}}{J_{C,S_2}} = 1$   
235 (Fig. 3a) while, when  $\frac{J_{C,S_1}}{J_{C,S_2}} = 25$ , the cell trajectories dramatically shift over the  
236 stiff part of the substrate (Fig. 3b). Our numerical results are sustained and  
237 consistent with the experimental observations according to which cells (i.e.,  
238 fibroblasts, smooth muscle cells, Mesenchymal Stem Cells (MSCs)) crawl from  
239 soft (1 to 5 kPa) to stiff (34 to 80 kPa) substrates (i.e., gels or polyacrylamide  
240 sheets) [9–11,46]. Notably during motion towards the stiff substrate, our CPM  
241 cell is also allowed to increase its remodeling ability, as its rigidity  
242  $v_{\Sigma_1}$  progressively decreases upon contact with substrate  $S_2$ , according to Eq. (6).  
243 In this respect, a further set of simulations evaluates cell morphological  
244 differences due to the underneath type of substrate. Keeping the same domain as  
245 in Fig. 2a, two cells, i.e.,  $S_1$  and  $S_2$ , are initially seeded in the middle of the soft  
246 and the stiff regions, respectively. The rigidity of the two cells is then regulated  
247 by Eq. (6). As reproduced in Fig. 4 (in particular, panel (a) represents the final cell  
248 morphologies as resulted from a single representative simulation, whereas panel

249 (b) gives the mean final cell morphologies, as the plain ellipsoids derive from an  
 250 interpolation procedure of the cell adhesive areas coming from independent  
 251 simulations, see the Appendix for further details), both individuals do not  
 252 significantly move across the domain during a time lapse of 500 MCS  
 253 (approximately 16 min). However, the adhesive area of the cell located over the  
 254 soft region is almost 30% lower than the adhesive area of the cell that crawls over  
 255 the stiff substrate (Fig. 4c). Such a cell behaviour is consistent with the  
 256 experimental data by Lo and co-workers on 3T3 fibroblasts cultured on flexible  
 257 polyacrylamide sheets coated with type I collagen, where a transition in rigidity  
 258 was introduced by a discontinuity of the bis-acrylamide cross-linker, that resulted  
 259 in two substrate regions with Young's modulus equal to either 14 kPa and 30 kPa  
 260 [46]. In particular, on one hand, the value of the adhesive area of our CPM cell  
 261 seeded on the soft substrate is not surprisingly similar to the corresponding data  
 262 by Lo and co-workers [46], since we used such an experimental quantification for  
 263 our parameter estimate (see the Appendix). On the other hand, the adhesive area  
 264 of the CPM cell seeded on the stiff region is instead a completely independent and  
 265 self-emerging model outcome: therefore its consistency with the measurements by  
 266 Lo and colleagues [46] is relevant point of our work.

267

### 268 3.2 *Stiff vs soft substrate in the presence of an external cue*

269 For the second series of simulations we consider again a domain split into a  
 270 soft ( $t = \mathfrak{S}_1$  such as  $J_{C,S_1} = J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$ ) and a stiff ( $t = \mathfrak{S}_2$  such as  
 271  $J_{C,S_2} = J_{stiff} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ) region, but an additional external potential is  
 272 introduced. This results in an imposed artificial bias in the spin flip rate that is  
 273 able to affect the direction of cell migration. Entering more in details, the

274 expression of the Hamiltonian function presented in Eq. (3) is modified as it  
275 follows

$$276 \quad H(\mathbf{t}) = H_{adhesion}(\mathbf{t}) + H_{shape}(\mathbf{t}) + H_{potential} \quad (7)$$

277 where  $H_{potential} = -v_{ext}(\mathbf{x}_{target} - \mathbf{x}_{source})$  and  $v_{ext}$  is a vector whose components  
278 determine the direction of the potential and whose modulus gives the relative  
279 importance in the overall system energy. For the sake of simplicity, we assume  
280 that the potential is constant in time and homogeneous throughout the entire  
281 domain. As we will see later, it is in fact an artificial term that simply helps cells  
282 to maintain a sustained directional movement. In this respect, what is relevant is  
283 only its modulus, i.e.,  $|v_{ext}|$ . We then test two configurations:

- 284 a) a cell  $S_1$  placed at the south-east corner and the external potential directed  
285 towards the north-west corner ;
- 286 b) the same cell  $S_1$  placed at the south-west corner of the substrate and the  
287 external potential directed towards the north-east corner.

288 In both cases, we set  $|v_{ext}| = 7 \cdot 10^{-21} \text{Kg}\cdot\text{m}/\text{s}^2$ , which results in plausible cell  
289 velocities (see later) and the simulations last 10000 MCS (approximately 5.5 h).  
290 Further, cell rigidity is again regulated by Eq. (6).

291 In system configuration a) (see Movie 1 and Fig. 5a), the external cue guides the  
292 cell towards the north-west corner of the domain. In particular, when a part of the  
293 cell comes into contact with the stiffer substrate, it becomes the leading edge.  
294 Further, the moving individual clearly accelerates as soon as it crosses the  
295 boundary between the two matrix regions (3.6  $\mu\text{m}/\text{s}$  versus 4.5  $\mu\text{m}/\text{s}$ , Fig. 5c), as  
296 experimentally observed in [8] for fibroblasts crawling over polyacrylamide

297 sheets. An increment of the adhesive area is observed as well when the cell shifts  
298 over the stiff region.

299 In the case b), the external potential forces the cell to move towards the north-  
300 east corner of the domain (see Movie 2 and Fig. 5b). However, as soon as the  
301 individual approaches the soft region, it changes orientation, and starts moving  
302 and elongating parallel to the boundary between the two substrate regions. These  
303 outcomes may be compared to the experimental observations obtained by Lo et al.  
304 in [46], who cultured fibroblasts on the already described substrate system, i.e.,  
305 characterized by two areas with different Young's modulus. In particular, Lo and  
306 colleagues seeded cells at low density to minimize the effects of intercellular  
307 interactions and to avoid that pulling or pushing forces from neighbours  
308 individuals may alter cell substrate probing processes (thereby impeding cells to  
309 freely move across the soft and the stiff regions). Then, cell migration was  
310 recorded over a time span of 10 h. Similarly to our numerical outcomes, the  
311 authors found that as cells move towards a stiffer substrate, new lamellipodia are  
312 formed in the direction of migration, thereby resulting in the dominant front end  
313 of the individuals. On the opposite, local retractions occur when cells approach a  
314 soft region, inducing therefore a change of direction. In a second series of  
315 experiments, Lo and co-workers showed that mechanical inputs triggered by  
316 substrate deformations might also control formation and retraction of  
317 lamellipodia. In particular, they externally pulled or pushed the substrate away or  
318 towards the cells centre to find that, due to the centripetal forces exerted by the  
319 3T3 fibroblasts on the substrate [46], in the first case less motion is produced,  
320 since cells experience a softening of the substrate, whereas in the second case the  
321 overall motion is increased, since cells perceive the substrate as stiffer. In the

322 CPM model proposed here, the matrix substrates are not deformable, therefore the  
323 numerical simulations are unable to capture the experimental observations coming  
324 from this second set of assays.

325

### 326 3.3 *Two stiff stripes embedded in a soft substrate*

327 The third configuration that has been tested includes a soft substrate (again  
328  $t = \mathfrak{S}_1$  with  $J_{C,S_1} = J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$ ) with two embedded stiff stripes (again  
329  $t = \mathfrak{S}_2$  with  $J_{C,S_2} = J_{stiff} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ), which are both 28  $\mu\text{m}$ -wide (Fig. 2b).  
330 A cell  $S_1$  is initially seeded at the south-west corner, whose rigidity is allowed to  
331 decrease following the constitutive law (Eq. (6)). An external potential is then  
332 introduced towards the north-east corner of the domain: its intensity  $|\mathbf{v}_{ext}|$  is  
333 allowed to vary from a minimal value of  $7 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  to a maximal value of  
334  $28 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ . All simulations last 10000 MCS, which correspond to nearly 5.5  
335 h. In the case of a low  $|\mathbf{v}_{ext}| = 7 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  (see Fig. 6a and Movie 3), the cell  
336 typically migrates towards the first stiff substrate stripe: then it remains stuck over  
337 it and goes on migrating along such a matrix region. Furthermore, its morphology,  
338 due to the dependency of its elasticity on the underneath type of substrate,  
339 changes as the crawling individual acquires an elongated shape. Such a behaviour  
340 is due to the fact that the external potential is too low to overcome the adhesive  
341 interactions between the cell and the stiffest substrate: in particular, the individual  
342 has not energetic benefits (deriving from the external bias) to move further in the  
343 domain, i.e., to pass the first stiff stipe. The outcomes of our CPM are consistent  
344 to that observed for cells (i.e., endothelial cells or fibrosarcoma cells) seeded on

345 2D substrates (i.e., maleic acid copolymer surfaces) structured with fibronectin  
346 stripes which orient their actin fibres along the stripe direction [47–49].

347 On the other hand, if the modulus of the external potential increases, we have a  
348 higher percentage of cells that are able to cross the entire domain (Fig. 6b). In  
349 particular, when  $|\boldsymbol{v}_{ext}|$  is maximal (i.e.,  $28 \cdot 10^{-21}$  Kg·m/s<sup>2</sup>, Fig. 6c and Movie 4),  
350 the cells constantly migrate at the north-east corner of the domain passing also the  
351 second stiff stripe. In this case, the cell average velocity increases over the stiff  
352 stripes (about 4.4  $\mu\text{m/s}$ ) whereas it varies between 3.6  $\mu\text{m/s}$  and 3.9  $\mu\text{m/s}$  over the  
353 soft regions (Fig. 6d). With the maximal external potential, cell morphology does  
354 not significantly vary, as the moving individuals typically maintain an almost  
355 hemispheric shape, without substantial elongation or increments in the adhesive  
356 area during the entire motion. They in fact behave as translating rigid bodies,  
357 subjected to an external high force. This interesting behaviour is the consequence  
358 of the fact that the cells do not need to reorganize (nor have enough time to do it)  
359 to be able to crawl, as their motion is mainly due to the external bias: the specific  
360 substrate regions are only able to further accelerate (or partially slow down) cell  
361 movement, as previously commented. The numerical outcomes in the case of low  
362 or intermediate values of  $|\boldsymbol{v}_{ext}|$  can be compared to those experimentally tested by  
363 Choi et al. [50] and Vincent et al. [51], where different cell phenotypes were  
364 seeded on micropatterned hydrogels with stiffness gradient. Although no external  
365 bias was introduced in such experimental configurations, a similar behaviour may  
366 be observed. In the former work [50], the authors proposed two mechanically-  
367 patterned hydrogels: one constituted by 100  $\mu\text{m}$  stiff (10 kPa) and 500  $\mu\text{m}$  soft (1  
368 kPa) stripes and one containing 500  $\mu\text{m}$  stiff (10 kPa) and 100  $\mu\text{m}$  soft (1 kPa).  
369 First, Adipose-derived Stem Cells (ASCs) and C2C12 myoblasts were allowed to



370 adhere and both were able to sense the stiffness gradient and to migrate towards  
371 the stiffer stripes (i.e., durotaxis) [46]. Such behaviour was also observed when  
372 cells were far away from the stripe interface (around 250  $\mu\text{m}$ ). Nevertheless, since  
373 cells only detect stiffness differences over short distances (around some microns)  
374 [52], in this case the authors implied that the phenomenon was mostly due to  
375 random walk towards the interface rather than to durotaxis itself. Regarding the  
376 morphology of the cells, both ASCS and C2C12 myoblasts aligned in the  
377 direction of the long axis of the stripe as we observe in our numerical simulations  
378 (Fig. 5a and Movie 3) in the case of low intensity of the external potential.  
379 Second, less contractile cells such as neurons were seeded on the hydrogels,  
380 which did not show any preferential adhesion confirming previous experimental  
381 observations according to which they prefer a softer niche [53].

382 In the latter work [51], the authors developed three types of polyacrylamide  
383 (PA) hydrogel systems of stiffness gradients: physiological (1 Pa/ $\mu\text{m}$ ),  
384 pathological (10 Pa/ $\mu\text{m}$ ) and step (100 Pa/ $\mu\text{m}$ ). The step stiffness gradient, which  
385 is the configuration of interest for the simulations presented above in this section,  
386 was constituted by 500  $\mu\text{m}$  wide regions of soft PA alternated with  $\sim 100$   $\mu\text{m}$  wide  
387 stripes of stiff hydrogel producing a stripped stiffness profile. MSCs were plated  
388 and they spread and attached independently of the gradient strength or the  
389 stiffness within hours after the seeding, whereas after 3 days they started to  
390 migrate towards stiffer regions. Additionally, cells crawled at  $18 \pm 0.7$   $\mu\text{m/hr}$ ,  
391 which is approximately 6-fold faster than on the other gradient configurations  
392 discussed in the same paper (i.e. physiological and pathological) and confirms that  
393 durotaxis velocity is influenced by gradient strength [11].

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### 3.4 Stiffness gradient

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In this section, we present the results for a simulation involving a substrate made of six successive stripes (i.e.,  $t = S_i$  where  $i = 1, \dots, 6$ , each 46  $\mu\text{m}$ -wide) which are organized to obtain a soft-to-stiff gradient from the left to the right side of the domain (from the red to the yellow subdomains). Such substrate regions are characterized by different cell adhesive affinity, i.e.,  $J_{C,S_i}$ , which vary from  $J_{C,S_1} = J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$  to  $J_{C,S_6} = J_{stiff} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ , respectively (see Fig. 2c and the corresponding caption for the specific details). A cell  $S_1$  is initially seeded at the south-west corner and an external potential is introduced towards the north-east corner of the domain, whose magnitude  $|\mathbf{v}_{ext}|$  is varied again from a minimal value of  $7 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  (Movie 5) to a maximal value of  $28 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  (Movie 6). The rigidity  $\eta_{S_1}$  of  $S_1$  is allowed to decrease (from the usual initial high value of  $\nu_C = 25 \cdot 10^{-3} \text{ Kg/s}^2\text{m}^2$ ) with a law analogous with Eq. (6), but which takes into account of the presence of different types of substrates, i.e.,

$$\nu_{\Sigma_1}(t) = \begin{cases} \max(\nu_{\Sigma_1}(t-1) - \nu_i; \nu_t) & \text{if } \exists (\mathbf{x}, \mathbf{x}' \in \Omega'_x): \mathbf{x} \in \Sigma_1 \text{ and } \mathbf{x}' \in S_i; \\ \nu_{\Sigma_1}(t-1) & \text{else,} \end{cases} \quad (8)$$

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where  $t$  is the actual MCS,  $\eta_t$  is the usual threshold value (equal to  $10^{-2} \text{ Kg/s}^2\text{m}^2$ ) and  $i = 2, \dots, 6$ . In this respect,  $\nu_i = 0.05 \cdot 10^{-3}, 0.06 \cdot 10^{-3}, 0.1 \cdot 10^{-3}, 0.2 \cdot 10^{-3}, 1 \cdot 10^{-3} \text{ Kg/s}^2\text{m}^2$  while the cell is in contact with substrate  $S_2, S_3, S_4, S_5, S_6$ , respectively.  $\eta_{S_1}$  remains indeed constant and equal to  $\nu_C$  if the cell is located over the softest substrate  $S_1$ . All the resulting simulations last 10000 MCS (5.5 h). As reproduced in Fig. 7a, the percentage of cells able to reach the north-east corner increases

417 concomitantly with increments of  $|\mathbf{v}_{ext}|$ . Moreover, by fixing  $|\mathbf{v}_{ext}| = 28 \cdot 10^{-21}$   
418  $\text{Kg}\cdot\text{m}/\text{s}^2$ , it is possible to observe that the cell average velocity increases from 3.6  
419  $\mu\text{m}/\text{s}$  to 4.4  $\mu\text{m}/\text{s}$  as they move from softer to stiffer substrates (Fig. 7b). This  
420 result is coherent with the model outcomes of the previous set of simulations (i.e.,  
421 see Fig. 6c and d), where we have observed that in the case of very high external  
422 potential cells accelerate while crossing on stiffer matrix regions, even if they do  
423 not significantly undergo morphological transitions.

424 A similar configuration was experimentally proposed by Cheung et al. [10]  
425 who, using a microfluidics-based lithography technique, fabricated a  
426 micropatterned cell-adhesive substrate made of a series of PEG-fibrinogen  
427 hydrogels with uniform stiffness ranging from 0.7 to 50 kPa. Human Foreskin  
428 Fibroblasts (HFFs) were then plated and their migratory trajectories were analysed  
429 over 22 h. The authors found that the cells that were initially seeded on a stiffness  
430 frontier tended to migrate towards the stiffer region, while cells plated on uniform  
431 stiffness spread in both directions.

432

### 433 *3.5 Role of the characteristic dimension of the gradient stiffness*

434 The external potential introduced in most of the previous sets of simulations, is  
435 an artificial term that is included in the Hamiltonian to bias and sustain cell  
436 movement across the entire matrix substrate. In experimental assays, the  
437 directional component in cell motion is typically established by geometrical cues,  
438 such as microtracks and microchannels [54,55], or gradients of soluble or  
439 insoluble chemical substances (chemotaxis and haptotaxis, respectively) or, in the  
440 case of our interest, gradients of substrate stiffness [10,51]. However, we have  
441 observed from our simulations that the sequence of different types of substrate

442 stripes employed in the previous section does not suffice to determine a persistent  
443 cell movement across the entire matrix, since a high enough external potential has  
444 to be included to allow cells reach the north-east corner of the domain (see the  
445 plot in Fig. 7a). The reason of this discrepancy between computational and  
446 experimental outcomes relies in the fact that “real cells”, once established a  
447 direction of movement, are able to dramatically orient their cytoskeleton (via the  
448 polarization of actin filaments) and, eventually, start a **persistent** shape-dependent  
449 locomotion. This way, real individuals are able to cross also large portions of  
450 substrates without slowing down or changing direction. Such a cell behaviour can  
451 not be captured in our approach since we do not include a proper model  
452 component reproducing selected intracellular cytoskeletal dynamics (in this  
453 respect, the interested reader may refer to [56,57], where polarization processes  
454 and the subsequent cell persistent movements are simulated in CPMs either by  
455 introducing an asymmetric correction to the Boltzman probability law or by  
456 adding a further inertial term in the Hamiltonian). The CPM cells of our model are  
457 only able to isotropically spread (due to decrements in their rigidity upon contact  
458 with stiff substrates) or elongate following the geometry of the underlying matrix  
459 region in order to maximize their adhesive interactions with the stiffer areas of the  
460 domain (but only when the external potential is substantially low, see Fig. 6a and  
461 c). However, the model presented in this paper can be used to predict if a  
462 sustained cell motion can be achieved by only varying the geometrical  
463 characteristics of the matrix substrate. With this purpose in mind, we employ the  
464 same type of domain as in Section 3.4, but we progressively decrease the width of  
465 the substrate stripes. We then evaluate the minimal magnitude of the external  
466 potential needed by cells to reach the border of the domain opposite to their initial

467 position (again the south-west corner). Cell rigidity follows the law in Eq. [8] and  
468 the simulations last 10000 MCS (5.5 h). As summarized in panel (a) of Fig. 8, we  
469 can observe a tri-phasic behaviour. For sufficiently wide stripes (i.e.,  $> 45 \mu\text{m}$ ), a  
470 cell sustained movement results only with very high external potentials (i.e.,  
471  $> 25 \cdot 10^{-21} \text{ Kg}\cdot\text{m}/\text{s}^2$ ). Then, for lower stripe widths (i.e., in the range of 35-45  $\mu\text{m}$ )  
472 the critical value of the external potential modulus decreases almost linearly.  
473 Finally, for low enough stripe widths (i.e.,  $< 35 \mu\text{m}$ ), the potential necessary to  
474 have a sustained cell movement significantly drops, until becoming negligible for  
475 stripe widths lower than 35  $\mu\text{m}$  (Movie 7). Summing up, we can state that the  
476 characteristic dimension of the stiffness gradient (here determined by the width of  
477 the matrix stripes), which allows a persistent cell movement without the artificial  
478 help of an external bias, is lower than the mean cell diameter (i.e., that in our  
479 simulations is around 40-45  $\mu\text{m}$ ). From a computational viewpoint, the rationale  
480 of this behaviour is that when a CPM cell is located on a given substrate stripe it  
481 is however able to wandering its close proximity (due to the stochastic Metropolis  
482 algorithm) which, if the stripe width is low enough, includes the neighbouring  
483 matrix region. In this respect, the CPM cell simultaneously experiences the  
484 adhesive affinity with a couple of neighbouring substrate stripes and then it moves  
485 towards the stiffer one, thereby advancing across the domain. Such a process is  
486 reiterated for all pairs of substrate stripes, thereby resulting in a sustained  
487 directional movement. These results can be interpreted from an experimental  
488 viewpoint as a prediction on the fact that cells may exhibit a persistent motion  
489 also without an intracellular polarization, i.e., by only maintain an amoeboid  
490 movement, if the substrate stiffness gradient is sufficiently fine-grained.

491 We finally conclude this section by analysing how cell velocity is affected by the  
492 wide of the substrate stripes, in the range of values sufficiently low to have a  
493 sustained cell crawling in the absence of an external potential (i.e.,  $< 35 \mu\text{m}$ ). As it  
494 is possible to see in panel (b) of Fig. 8, lower widths of the substrate regions  
495 (which means, as previously seen, more fine-grained stiffness gradients) results in  
496 increments in cell average velocity. From the computational viewpoint, this is due  
497 to the fact that the more the different stripes of the matrix are small, the more the  
498 previously described cell probing mechanism is facilitated and accelerated,  
499 thereby resulting in higher cell average velocities.

500

### 501 3.6 *Soft squares embedded in a stiff substrate*

502 As a final simulation, we test the substrate configuration in Fig. 2d, where four  
503 soft squares ( $t = \mathfrak{S}_1$  with  $J_{C,S_1} = J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$ ) are embedded in a stiff  
504 substrate ( $t = \mathfrak{S}_2$  with  $J_{C,S_2} = J_{stiff} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ) at its three corners (north-  
505 west, north-east and south-east) and at the centre. A cell  $S_1$  is initially seeded at  
506 the south-west corner and an external potential is introduced towards the north-  
507 east corner of the domain, whose magnitude  $|\mathbf{v}_{ext}|$  has been set equal to an  
508 intermediate  $14 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ . As usual, cell rigidity is allowed to decrease  
509 according to Eq. (6) and the observation time is 10000 MCS (i.e., nearly 5.5. h).  
510 The cell starts moving in the direction determined by the potential with a  
511 trajectory of approximately  $45^\circ$  but, as soon as it encounters the central soft  
512 square, the cell avoids and circumvents it. As the original path is recovered, the  
513 cell needs to squeeze between the north-east square and the substrate frontier in  
514 order to achieve the target corner of the domain (Movie 8 and Fig. 9). The choice

515 of the migration track may depend on the initial position of the cell. In the present  
516 simulation, the cell is seeded along the substrate diagonal, thus the probability of  
517 circumventing the central soft square counter clockwise (as it happens here) or  
518 clockwise are the same. However, if the cell is seeded slightly downward and/or  
519 right, it will most certainly employ a counter clockwise trajectory, whereas if it is  
520 plated upward, it will probably follow a clockwise path. It is useful to notice that  
521 with a significantly higher modulus of the external potential the cell would have  
522 been able to pass across the soft regions, without deforming to avoid them,  
523 coherently with the simulations proposed in Fig. 6c.

524 This configuration is similar to that proposed in [23] where the cell must  
525 avoid two slipping regions in order to reach the external cue placed at  $45^\circ$ .  
526 Although the employed numerical approaches are substantially different, taken  
527 together the outcomes confirm the tendency of the cell to migrate over stiffer  
528 substrates where the higher adhesion forces may be developed.

529

#### 530 **4. Conclusions**

531 In this paper we have proposed a three-dimensional CPM approach to simulate  
532 single cell migration over matrix domains in which soft and stiff regions are  
533 combined.

534 The CPM method is becoming an increasingly common technique for the  
535 mathematical modelling of a wide range of biological phenomena, including  
536 avascular and vascular tumor growth [58–61], gastrulation [62], skin pigmentation  
537 [63], yeast colony growth [64], stem cell differentiation [65], fruiting body  
538 formation of *Dictyostelium discoideum* [66], epidermal formation [67], hydra  
539 regeneration [66], retinal patterning [68], wound healing [69,70], biofilms [71],

540 chick limb-bud growth [72–74], cellular differentiation and growth of tissues,  
541 blood flow and thrombus development [75–77], angiogenesis [70,78–81],  
542 dynamics of vascular cells [82–85], cell scattering [86], cell migration on and  
543 within matrix environments [56,57,87]. Notably, in [88] the authors introduced a  
544 compartmentalized approach to subdivide a *Myxococcus Xanthus* into strings of  
545 subcellular domains with different rigidity, this in order to give the bacterium a  
546 particular geometry and to control its overall length. Further, in [89] a keratocyte  
547 has been represented with a set of undifferentiated hexagonal subunits, which has  
548 allowed to reproduce its polarization during motion. In this respect, it is useful to  
549 underline that, as commented in [25], although these approaches are correct, the  
550 fact that the proposed subcellular compartments do not have an immediate or  
551 direct correspondence with real subcellular elements, has limited the practicality  
552 and the usefulness of the relative models. The most accurate way of realistically  
553 reproducing different and extremely complex cell morphologies is to  
554 compartmentalize them according to the compartmentalization “suggested in  
555 nature”, and thus to explicitly represent for instance the plasmamembrane (PM),  
556 the cytosolic region, the nucleus, and other intracellular organelles (e.g.,  
557 mitochondria, ribosomes, Golgi apparatus, and secretory granules). This way is in  
558 fact possible, for example, to localize within the proper cell compartment selected  
559 biochemical pathways and/or to study the role play by the nucleus in cell  
560 movement, given its higher rigidity with respect to the surrounding cytoplasm  
561 [56,57,87].

562 Key benefits of the CPM energetic formalism are its simplicity and  
563 extensibility: almost any biological mechanism can in fact be included in the  
564 model, simply by adding an appropriate generalized potential term in the



565 Hamiltonian functional. For instance, it is possible to easily comprehend the  
566 importance of each mechanism involved in the simulated phenomenon by only  
567 altering the relative Potts parameter, so that the other terms in the Hamiltonian  
568 scale accordingly. In particular, by equating all the other terms to zero, it is  
569 possible to understand whether a mechanism is individually capable of producing  
570 the phenomenon of interest or whether it requires cooperative processes. Further  
571 critical features of the CPM (compared to alternative cell-based modelling  
572 approaches that represent biological individuals as point particles, such as  
573 Interacting Particle Systems or purely discrete models, or fixed-sized spheres or  
574 ellipsoids, such as Cellular Automata) is that i) it differentiates between bound  
575 and unbound regions of cell membranes and ii) morphological changes can be  
576 easily and realistically reproduced. These characteristics have been fundamental  
577 in our choice of using a CPM to describe the phenomenon of our interest since  
578 they are particularly suitable to implement our two main model assumptions,  
579 drawn according to the experimental observations reported in the literature: i) the  
580 adhesiveness of cells changes according to the substrate stiffness, that models the  
581 fact that higher traction forces and more stable focal points are generated over a  
582 stiffer substrate [16,35–38] and ii) each cell adapts its morphology as a function  
583 of the substrate stiffness so that over a soft region it maintains a rounded shape,  
584 whereas over a stiffer domain a significant spreading occurs [39–43]. The  
585 considerations above are in remarkable agreement with the scholarly dissertation  
586 proposed by Voss-Böhme in the conclusive section of her article [33]. She in fact  
587 argued that the application of CPMs is reasonable when the biological problem of  
588 interest involves “considerable variability in cell sizes and shapes”, which is the  
589 case of the cell morphological transitions due to contact with soft/stiff substrates.

590 On the opposite, when “essentially isotropic, non-polarized cells of uniform size  
591 are considered”, it would be preferable the use of more coarse-grained modelling  
592 approaches, like the already cited Cellular Automata or Interacting Particle  
593 Systems, which are better analysed both mechanistically and analytically.

594 Further, we have opted for a 3D setting since the adhesive interactions between  
595 cells and matrix substrates occur *under* the cell body (i.e., they are localized over  
596 the contact area between the cells themselves and the underneath substrate). In bi-  
597 dimensional CPMs cell-matrix interactions instead occur only “laterally”, as the  
598 cells do not move *on* substrates but *within* the same plane as the matrix. Indeed, a  
599 three-dimensional domain is more appropriate to reproduce an adhesive-driven  
600 cell migration.

601 We have then used our CPM-based approach to test cell behaviour in different  
602 domain configurations, where soft and stiff substrates coexisted. In particular, the  
603 numerical outcomes have been consistently compared to specific experimental  
604 data, in terms of cell morphology, distance covered, spreading/adhesive area and  
605 migration speed. In this respect, following the dichotomy proposed in the already  
606 cited work by Voss-Böhme [33], we have interpreted our CPM as a  
607 *phenomenological* method. In particular, the resulting remarkable agreement (not  
608 only qualitative but also quantitative) between *in vitro* and *in silico* data has  
609 allowed us to conclude that our approach, although strongly simplified, was able  
610 to capture the main mechanisms underlying cell migration in presence of  
611 durotaxis. We have finally turned to use our model in a *predictive* manner, with  
612 the aim to analyse how the external potential and the critical dimensions of a  
613 substrate stiffness gradient (here represented by the width of the different types of  
614 matrix stripes) affect cell movement. In this respect, we have found that cells are

615 able to achieve a sustained cell migration in the absence of an external bias (and  
616 in the absence of intracellular polarization mechanisms) where the underneath  
617 matrix is characterized by a sufficient fine-grained gradient of rigidity.

618       However, our approach is not free of some serious shortcomings. First, it does  
619 not reproduce the active and continuous reorganization of the cytoskeleton, which  
620 provides the support for cells and mediates their coordinated and directed  
621 movements, mainly in response to mechanical tensions and stresses exchanged  
622 with the underneath substrate. In this respect, selected geometrical and mechanical  
623 properties of the cells, such their elongation and elasticity, should evolve  
624 according to a model of actin filament dynamics, which are powered, for example,  
625 by ATP (adenosine triphosphate) hydrolysis and controlled by inside-out  
626 signalling mechanisms transmitted from and by the extracellular matrix via focal  
627 adhesion points. Further, in our model, the substrates are not deformable and  
628 therefore it has not been possible to account how the matrix reacts to the probing  
629 processes exerted by crawling cells. Finally, it is useful to underline that our  
630 specific CPM application does not suffer of the limitation that Voss-Böhme  
631 proved to characterize most CPMs (see again [33]), i.e., cells die out in the long-  
632 run due to modifications in the original Metropolis algorithm. We have in fact  
633 focused on relatively short observation times: our model has indeed worked in a  
634 well-behaved parameter regime where the temporal evolution of the simulated  
635 system has been still directed towards the minimization of the Hamiltonian  
636 functional and the non-controlled, voter-like part of the lattice updates has been  
637 negligible.

638 **Appendix**

639 *A.1 Morphological and migratory determinants*

640 The *position* of a cell was established by the coordinate of its center of mass  
641 (CM). In particular, a cell was assumed to be located on a given type of substrate  
642 if its center of mass was located on that matrix region. In this respect, the  
643 migratory *trajectory* of a cell was generated by tracking the position of its center  
644 of mass at each time step (i.e., at each MCS).

645 The *adhesive area* of a cell was defined as the extension of its surface in  
646 contact with the substrate of interest at the final observation time.

647 The *average velocity* of an individual on a given type of substrate was  
648 measured as the ratio between the width of the substrate region itself (which is  
649 clarified for each simulation setting) and the time needed by the cell to cross it. In  
650 this respect, to obtain the amount of time spent by a cell to pass a given matrix  
651 region it is sufficient to multiply the corresponding average velocity for the width  
652 of the substrate of interest.

653

654 *A.2 Statistics*

655 In the plots, we represented cell trajectories coming from 10 independent and  
656 randomly chosen simulations. A number of 10 was chosen since we observed that  
657 it was sufficient to have a correct interpretation of the simulation outcomes but it  
658 was also low enough to have an acceptable graphical quality, as too many cell  
659 paths overlapped one to each other, thereby resulting undistinguishable.

660 Cell average velocity and adhesive area were instead given in the  
661 corresponding graphs as mean  $\pm$  sd over 100 independent simulations.

662 In the plots representing the cell final distribution on the different types of  
663 substrate, the relative frequency was given by the number of individuals that, over  
664 100 independent simulations, were located over each matrix region at the end of  
665 the observation time. Indeed, the sum of the relative frequencies is, in all cases,  
666 equal to 100.

667 Finally, dashed and plain ellipsoids representing, respectively, initial and final  
668 cell morphologies in a given simulation setting were established by interpolating  
669 the cell adhesive areas coming from 10 independent simulations (typically the  
670 ones used to track the cell trajectories for the same simulation setting). Obviously,  
671 the initial cell position was the constant for each simulation setting, whereas the  
672 initial cell shape was the same for all cases (i.e., an hemisphere of 20  $\mu\text{m}$  of  
673 radius).

674

### 675 *A.3 Parameter estimates*

676 Given the energetic nature of the CPM, a direct one-to-one correspondence  
677 between model parameters and experimental quantities is not straightforward (as  
678 commented also in [27] and in [90]). In particular, as explained in details in [33],  
679 the CPM parameters can be subdivided in i) directly interpretable and measurable  
680 quantities, such as cell geometrical dimensions ii) effective parameters that  
681 subsume various cellular and subcellular properties, such as the adhesive strength  
682 and the elastic moduli determining cell geometrical attributes iii) “merely  
683 technical coefficients”, such as the Boltzmann temperature, that has been  
684 interpreted in different ways by CPM authors (in this work, we opted to link the  
685 value of  $T_C$  to an intrinsic cell motility, i.e., the amplitude of cell boundary  
686 fluctuations: consistently, we therefore added the subscript “C” to the coefficient

687  $T_C$ ). However, a plausible parameter setting was inferred by a proper comparison  
688 with experimental findings, taking also advantage of selected sensitivity analysis  
689 performed in other CPM-based works. First, the initial/target dimensions of our  
690 virtual cells were consistent with the measures of NIH 3T3 mouse fibroblasts used  
691 by Lo and colleagues [8] for their assays on durotaxis. Since, as previously seen,  
692 we did not include in our model any nutrients and therefore cells were not allowed  
693 to grow during migration, we set a high  $\kappa_C = 25 \cdot 10^{-9} \text{ Kg}/(\text{s}^2\text{m}^4)$  to keep cell  
694 volume fluctuations within a few per cent. Such a specific value was taken from  
695 other CPMs dealing both with single cell dynamics [82] and with multicellular  
696 phenomena [91], where it was estimated after some trials. Further, observing from  
697 the data by Lo et al. [46] that 3T3 cells seeded on soft enough substrates did not  
698 significantly spread or undergo morphological transitions, we set a high value  
699  $\nu_C = 25 \cdot 10^{-3} \text{ Kg}/(\text{s}^2\text{m}^2)$  also for the intrinsic cell rigidity which, as previously  
700 seen, can decrease (in our work) only upon cell contact with stiffer substrates.  
701 This choice is consistent with other CPM-based approaches [57,70,82,91] that  
702 employed similar values (i.e.,  $\geq 15$ ) to model an initially low cell deformability.

703 We then turned to estimate both the Boltzmann temperature  $T_C$  and the cell-  
704 substrate adhesiveness, denoted as  $J_{soft}$  throughout the paper, in the case of the  
705 softest matrix regions considered in this paper (i.e., the ones pseudo-coloured in  
706 red in the simulations, typically identified by  $\tau = S_1$ ). In particular, we looked for  
707 the couple of coefficients  $(T_C, J_{soft})$  that simultaneously best fitted the *in vitro*  
708 results by Lo and co-workers in the corresponding experimental setting (i.e.,  
709 collagen-coated polyacrylamide substrate properly manipulated to obtain a low  
710 Young's modulus of 14 kPa) in terms of cell adhesive area (which was called by  
711 Lo and colleagues "projected area" [46]). As it is possible to observe in Fig. 10,

712 there is a quite large range of values that matched experimental and computational  
713 data (i.e., the yellow area of the graph): however, we opted for the intermediate  
714 couple of coefficients  $T_C = 50 \cdot 10^{-27} \text{ Kg m}^2/\text{s}^2$  and  $J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$ .  
715 Decrements in cell-substrate adhesive strength and in cell rigidity, allowed in our  
716 model as a consequence of cell contact with stiffer matrix regions, were then  
717 performed until selected threshold values, i.e.,  $\nu_t = 10^{-2} \text{ Kg/s}^2 \text{ m}^2$  and  $J_{stiff} = 1 \cdot$   
718  $10^{-15} \text{ Kg/s}^2$ , respectively. In particular,  $\nu_t$  was the lowest value of cell rigidity  
719 that permitted to avoid unrealistic (often disconnected) cell shapes.  $J_{stiff}$ , i. e., the  
720 adhesive force between cells and the stiffest substrates (the ones pseudo-coloured  
721 in yellow in the simulation snapshots, typically labelled by  $\tau = S_2$ , except from  
722 the case of the simulations dealing with the stiffness gradient) was instead the  
723 lowest value for which cells did not start to slow down during migration. In fact,  
724 as studied in details in [87], a too high cell-substrate adhesiveness partially  
725 inhibits cell movement, as CPM cells are not able to detach from the matrix  
726 component if the corresponding J-parameter is too low. Finally, the cell-medium  
727 contact strength was set equal to  $J_{soft}$  (i. e.,  $= 25 \cdot 10^{-15} \text{ Kg/s}^2$ ). The rationale of  
728 this choice relied in the fact that for lower values of  $J_{C,M}$  cells detached from the  
729 substrate and fluctuate in the middle of the medium, which was obviously an  
730 unrealistic situation. On the opposite, too high values of  $J_{C,M}$  (i. e.,  $> 30 \cdot 10^{-15}$   
731  $\text{Kg/s}^2$ ) forced cells to completely lay down on the matrix, in order to minimize  
732 their contact surface with the medium, but also this situation was not plausible.  
733 Finally, the correspondence between 1 MCS and 2 seconds of actual time was  
734 taken from another CPM reproducing three-dimensional cell migration in matrix  
735 environments [87]. Further, we observed that this setting resulted in a remarkable  
736 accordance, in terms of cell velocity, between computational and experimental

737 results: our CPM cells in fact move at speeds in a range of (3, 5)  $\mu\text{m/s}$ , which is  
738 consistent with the values measured by Vincent and colleagues [51] in the case of  
739 MSCs plated on polyacrylamide (PA) hydrogels with selected stiffness.

## 740 **Acknowledgements**

741 This work was initiated and partially completed while Rachele Allena was a  
742 visitor to the Dipartimento di Scienze Matematiche of the Politecnico di Torino  
743 funded by the Gruppo Nazionale per la Fisica Matematica (GNFM).

## 744 **Bibliography**

745 [1] R.L. Juliano, S. Haskill, Signal transduction from the extracellular matrix, *J. Cell Biol.*  
746 120 (1993) 577–585.

747

748 [2] P. Martin, Wound healing--aiming for perfect skin regeneration, *Science.* 276 (1997)  
749 75–81.

750

751 [3] L.R. Bernstein, L.A. Liotta, Molecular mediators of interactions with extracellular  
752 matrix components in metastasis and angiogenesis, *Curr Opin Oncol.* 6 (1994) 106–113.

753

754 [4] D.A. Lauffenburger, A.F. Horwitz, Cell migration: a physically integrated molecular  
755 process, *Cell.* 84 (1996) 359–369.

756

757 [5] H. Harris, Role of chemotaxis in inflammation, *Physiol. Rev.* 34 (1954) 529–562.

758

759 [6] K.R. Robinson, The responses of cells to electrical fields: a review, *J. Cell Biol.* 101  
760 (1985) 2023–2027.

761

762 [7] S. Li, P. Butler, Y. Wang, Y. Hu, D.C. Han, S. Usami, et al., The role of the dynamics  
763 of focal adhesion kinase in the mechanotaxis of endothelial cells, *PNAS.* 99 (2002) 3546–  
764 3551. doi:10.1073/pnas.052018099.



765  
766 [8] C.M. Lo, H.B. Wang, M. Dembo, Y.L. Wang, Cell movement is guided by the rigidity  
767 of the substrate, *Biophys. J.* 79 (2000) 144–152. doi:10.1016/S0006-3495(00)76279-5.  
768  
769 [9] M. Raab, J. Swift, P.C.D.P. Dingal, P. Shah, J.-W. Shin, D.E. Discher, Crawling from  
770 soft to stiff matrix polarizes the cytoskeleton and phosphoregulates myosin-II heavy chain, *J*  
771 *Cell Biol.* 199 (2012) 669–683. doi:10.1083/jcb.201205056.  
772  
773 [10] Y.K. Cheung, E.U. Azeloglu, D.A. Shiovitz, K.D. Costa, D. Seliktar, S.K. Sia,  
774 *Microscale Control of Stiffness in a Cell-Adhesive Substrate Using Microfluidics-Based*  
775 *Lithography*, *Angewandte Chemie International Edition.* 48 (2009) 7188–7192.  
776 doi:10.1002/anie.200900807.  
777  
778 [11] B.C. Isenberg, P.A. DiMilla, M. Walker, S. Kim, J.Y. Wong, *Vascular Smooth Muscle*  
779 *Cell Durotaxis Depends on Substrate Stiffness Gradient Strength*, *Biophysical Journal.* 97  
780 (2009) 1313–1322. doi:10.1016/j.bpj.2009.06.021.  
781  
782 [12] I.B. Bischofs, U.S. Schwarz, *Cell organization in soft media due to active*  
783 *mechanosensing*, *PNAS.* 100 (2003) 9274–9279. doi:10.1073/pnas.1233544100.  
784  
785 [13] A. Saez, M. Ghibaudo, A. Buguin, P. Silberzan, B. Ladoux, *Rigidity-driven growth*  
786 *and migration of epithelial cells on microstructured anisotropic substrates*, *PNAS.* 104 (2007)  
787 8281–8286. doi:10.1073/pnas.0702259104.  
788  
789 [14] L. Trichet, J.L. Digabel, R.J. Hawkins, S.R.K. Vedula, M. Gupta, C. Ribault, et al.,  
790 *Evidence of a large-scale mechanosensing mechanism for cellular adaptation to substrate*  
791 *stiffness*, *PNAS.* 109 (2012) 6933–6938. doi:10.1073/pnas.1117810109.  
792  
793 [15] M. Raab, J. Swift, P.C.D.P. Dingal, P. Shah, J.-W. Shin, D.E. Discher, Crawling from  
794 soft to stiff matrix polarizes the cytoskeleton and phosphoregulates myosin-II heavy chain, *J*  
795 *Cell Biol.* 199 (2012) 669–683. doi:10.1083/jcb.201205056.  
796  
797 [16] C.A. Reinhart-King, M. Dembo, D.A. Hammer, *The dynamics and mechanics of*  
798 *endothelial cell spreading*, *Biophys. J.* 89 (2005) 676–689. doi:10.1529/biophysj.104.054320.

799

800 [17] P. Moreo, J.M. García-Aznar, M. Doblaré, Modeling mechanosensing and its effect on  
801 the migration and proliferation of adherent cells, *Acta Biomater.* 4 (2008) 613–621.  
802 doi:10.1016/j.actbio.2007.10.014.

803

804 [18] B. Harland, S. Walcott, S.X. Sun, Adhesion dynamics and durotaxis in migrating cells,  
805 *Phys Biol.* 8 (2011) 015011. doi:10.1088/1478-3975/8/1/015011.

806

807 [19] I.V. Dokukina, M.E. Gracheva, A Model of Fibroblast Motility on Substrates with  
808 Different Rigidities, *Biophys J.* 98 (2010) 2794–2803. doi:10.1016/j.bpj.2010.03.026.

809

810 [20] F. Stefanoni, M. Ventre, F. Mollica, P.A. Netti, A numerical model for durotaxis, *J.*  
811 *Theor. Biol.* 280 (2011) 150–158. doi:10.1016/j.jtbi.2011.04.001.

812

813 [21] S.A. Sandersius, T.J. Newman, Modeling cell rheology with the Subcellular Element  
814 Model, *Phys Biol.* 5 (2008) 015002. doi:10.1088/1478-3975/5/1/015002.

815

816 [22] C.W. Harvey, F. Morcos, C.R. Sweet, D. Kaiser, S. Chatterjee, X. Liu, et al., Study of  
817 elastic collisions of *Myxococcus xanthus* in swarms, *Phys Biol.* 8 (2011) 026016.  
818 doi:10.1088/1478-3975/8/2/026016.

819

820 [23] R. Allena, D. Aubry, “Run-and-tumble” or “look-and-run”? A mechanical model to  
821 explore the behavior of a migrating amoeboid cell, *J. Theor. Biol.* 306 (2012) 15–31.  
822 doi:10.1016/j.jtbi.2012.03.041.

823

824 [24] Graner, Glazier, Simulation of biological cell sorting using a two-dimensional  
825 extended Potts model, *Phys. Rev. Lett.* 69 (1992) 2013–2016.

826

827 [25] M. Scianna, L. Preziosi, Multiscale Developments of the Cellular Potts Model,  
828 *Multiscale Model. Simul.* 10 (2012) 342–382. doi:10.1137/100812951.

829

830 [26] A. Balter, R.M.H. Merks, N.J. Popławski, M. Swat, J.A. Glazier, The Glazier-Graner-  
831 Hogeweg Model: Extensions, Future Directions, and Opportunities for Further Study, in:  
832 D.A.R.A. Anderson, P.M.A.J. Chaplain, D.K.A. Rejniak (Eds.), *Single-Cell-Based Models in*

833 Biology and Medicine, Birkhäuser Basel, 2007: pp. 151–167.  
834 [http://link.springer.com/chapter/10.1007/978-3-7643-8123-3\\_7](http://link.springer.com/chapter/10.1007/978-3-7643-8123-3_7) (accessed July 29, 2015).  
835

836 [27] J.A. Glazier, A. Balter, N.J. Popławski, Magnetization to Morphogenesis: A Brief  
837 History of the Glazier-Graner-Hogeweg Model, in: D.A.R.A. Anderson, P.M.A.J. Chaplain,  
838 D.K.A. Rejniak (Eds.), *Single-Cell-Based Models in Biology and Medicine*, Birkhäuser  
839 Basel, 2007: pp. 79–106. [http://link.springer.com/chapter/10.1007/978-3-7643-8123-3\\_4](http://link.springer.com/chapter/10.1007/978-3-7643-8123-3_4)  
840 (accessed July 29, 2015).  
841

842 [28] N.J. Savill, R.M.H. Merks, The Cellular Potts Model in Biomedicine, in: D.A.R.A.  
843 Anderson, P.M.A.J. Chaplain, D.K.A. Rejniak (Eds.), *Single-Cell-Based Models in Biology  
844 and Medicine*, Birkhäuser Basel, 2007: pp. 137–150.  
845 [http://link.springer.com/chapter/10.1007/978-3-7643-8123-3\\_6](http://link.springer.com/chapter/10.1007/978-3-7643-8123-3_6) (accessed July 29, 2015).  
846

847 [29] A. Szabó, R.M.H. Merks, Cellular potts modeling of tumor growth, tumor invasion,  
848 and tumor evolution, *Front Oncol.* 3 (2013) 87. doi:10.3389/fonc.2013.00087.  
849

850 [30] E. Ising, Beitrag zur Theorie des Ferromagnetismus, *Z. Physik.* 31 (1925) 253–258.  
851 doi:10.1007/BF02980577.  
852

853 [31] R.B. Potts, Some generalized order-disorder transformations, *Mathematical  
854 Proceedings of the Cambridge Philosophical Society.* 48 (1952) 106–109.  
855 doi:10.1017/S0305004100027419.  
856

857 [32] N. Metropolis, A.W. Rosenbluth, M.N. Rosenbluth, A.H. Teller, E. Teller, Equation of  
858 State Calculations by Fast Computing Machines, *The Journal of Chemical Physics.* 21 (1953)  
859 1087–1092. doi:10.1063/1.1699114.  
860

861 [33] A. Voss-Böhme, Multi-Scale Modeling in Morphogenesis: A Critical Analysis of the  
862 Cellular Potts Model, *PLoS ONE.* 7 (2012) e42852. doi:10.1371/journal.pone.0042852.  
863

864 [34] M.S. Steinberg, Reconstruction of tissues by dissociated cells. Some morphogenetic  
865 tissue movements and the sorting out of embryonic cells may have a common explanation,  
866 *Science.* 141 (1963) 401–408.

867  
868 [35] J.P. Califano, C.A. Reinhart-King, Substrate Stiffness and Cell Area Predict Cellular  
869 Traction Stresses in Single Cells and Cells in Contact, *Cell Mol Bioeng.* 3 (2010) 68–75.  
870 doi:10.1007/s12195-010-0102-6.  
871  
872 [36] J.L. Tan, J. Tien, D.M. Pirone, D.S. Gray, K. Bhadriraju, C.S. Chen, Cells lying on a  
873 bed of microneedles: an approach to isolate mechanical force, *Proc. Natl. Acad. Sci. U.S.A.*  
874 100 (2003) 1484–1489. doi:10.1073/pnas.0235407100.  
875  
876 [37] C.A. Reinhart-King, M. Dembo, D.A. Hammer, Endothelial Cell Traction Forces on  
877 RGD-Derivatized Polyacrylamide Substrata†, *Langmuir.* 19 (2003) 1573–1579.  
878 doi:10.1021/la026142j.  
879  
880 [38] A.D. Rape, W.-H. Guo, Y.-L. Wang, The regulation of traction force in relation to cell  
881 shape and focal adhesions, *Biomaterials.* 32 (2011) 2043–2051.  
882 doi:10.1016/j.biomaterials.2010.11.044.  
883  
884 [39] T. Yeung, P.C. Georges, L.A. Flanagan, B. Marg, M. Ortiz, M. Funaki, et al., Effects  
885 of substrate stiffness on cell morphology, cytoskeletal structure, and adhesion, *Cell Motil.*  
886 *Cytoskeleton.* 60 (2005) 24–34. doi:10.1002/cm.20041.  
887  
888 [40] M. Ghibaudo, A. Saez, L. Trichet, A. Xayaphoummine, J. Browaeys, P. Silberzan, et  
889 al., Traction forces and rigidity sensing regulate cell functions, *Soft Matter.* 4 (2008) 1836.  
890 doi:10.1039/b804103b.  
891  
892 [41] A. Engler, L. Bacakova, C. Newman, A. Hategan, M. Griffin, D. Discher, Substrate  
893 compliance versus ligand density in cell on gel responses, *Biophys. J.* 86 (2004) 617–628.  
894 doi:10.1016/S0006-3495(04)74140-5.  
895  
896 [42] A.J. Engler, S. Sen, H.L. Sweeney, D.E. Discher, Matrix elasticity directs stem cell  
897 lineage specification, *Cell.* 126 (2006) 677–689. doi:10.1016/j.cell.2006.06.044.  
898  
899 [43] N. Zaari, P. Rajagopalan, S.K. Kim, A.J. Engler, J.Y. Wong, Photopolymerization in  
900 Microfluidic Gradient Generators: Microscale Control of Substrate Compliance to Manipulate

901 Cell Response, *Adv. Mater.* 16 (2004) 2133–2137. doi:10.1002/adma.200400883.  
902

903 [44] C.S. Chen, J.L. Alonso, E. Ostuni, G.M. Whitesides, D.E. Ingber, Cell shape provides  
904 global control of focal adhesion assembly, *Biochem. Biophys. Res. Commun.* 307 (2003)  
905 355–361.  
906

907 [45] S.J. Han, K.S. Bielawski, L.H. Ting, M.L. Rodriguez, N.J. Sniadecki, Decoupling  
908 Substrate Stiffness, Spread Area, and Micropost Density: A Close Spatial Relationship  
909 between Traction Forces and Focal Adhesions, *Biophys J.* 103 (2012) 640–648.  
910 doi:10.1016/j.bpj.2012.07.023.  
911

912 [46] C.M. Lo, H.B. Wang, M. Dembo, Y.L. Wang, Cell movement is guided by the rigidity  
913 of the substrate, *Biophys. J.* 79 (2000) 144–152. doi:10.1016/S0006-3495(00)76279-5.  
914

915 [47] S.I. Fraley, Y. Feng, A. Giri, G.D. Longmore, D. Wirtz, Dimensional and temporal  
916 controls of three-dimensional cell migration by zyxin and binding partners, *Nat Commun.* 3  
917 (2012) 719. doi:10.1038/ncomms1711.  
918

919 [48] P.P. Girard, E.A. Cavalcanti-Adam, R. Kemkemer, J.P. Spatz, Cellular  
920 chemomechanics at interfaces: sensing, integration and response, *Soft Matter.* 3 (2007) 307–  
921 326. doi:10.1039/B614008D.  
922

923 [49] A. Müller, J. Meyer, T. Paumer, T. Pompe, Cytoskeletal transition in patterned cells  
924 correlates with interfacial energy model, *Soft Matter.* 10 (2014) 2444–2452.  
925 doi:10.1039/c3sm52424h.  
926

927 [50] Y.S. Choi, L.G. Vincent, A.R. Lee, K.C. Kretchmer, S. Chirasatitsin, M.K. Dobke, et  
928 al., The alignment and fusion assembly of adipose-derived stem cells on mechanically  
929 patterned matrices, *Biomaterials.* 33 (2012) 6943–6951.  
930 doi:10.1016/j.biomaterials.2012.06.057.  
931

932 [51] L.G. Vincent, Y.S. Choi, B. Alonso-Latorre, J.C. del Álamo, A.J. Engler,  
933 Mesenchymal Stem Cell Durotaxis Depends on Substrate Stiffness Gradient Strength,  
934 *Biotechnol J.* 8 (2013) 472–484. doi:10.1002/biot.201200205.

935

936 [52] A.J. Engler, M.A. Griffin, S. Sen, C.G. Bönnemann, H.L. Sweeney, D.E. Discher,  
937 Myotubes differentiate optimally on substrates with tissue-like stiffness: pathological  
938 implications for soft or stiff microenvironments, *J. Cell Biol.* 166 (2004) 877–887.  
939 doi:10.1083/jcb.200405004.

940

941 [53] L.A. Flanagan, Y.-E. Ju, B. Marg, M. Osterfield, P.A. Janmey, Neurite branching on  
942 deformable substrates, *Neuroreport.* 13 (2002) 2411–2415.  
943 doi:10.1097/01.wnr.0000048003.96487.97.

944

945 [54] O. Ilina, G.-J. Bakker, A. Vasaturo, R.M. Hofmann, P. Friedl, Two-photon laser-  
946 generated microtracks in 3D collagen lattices: principles of MMP-dependent and -  
947 independent collective cancer cell invasion, *Phys Biol.* 8 (2011) 015010. doi:10.1088/1478-  
948 3975/8/1/015010.

949

950 [55] D. Irimia, G. Charras, N. Agrawal, T. Mitchison, M. Toner, Polar stimulation and  
951 constrained cell migration in microfluidic channels, *Lab Chip.* 7 (2007) 1783–1790.  
952 doi:10.1039/b710524j.

953

954 [56] M. Scianna, L. Preziosi, A cellular Potts model for the MMP-dependent and -  
955 independent cancer cell migration in matrix microtracks of different dimensions, *Comput*  
956 *Mech.* 53 (2014) 485–497. doi:10.1007/s00466-013-0944-6.

957

958 [57] M. Scianna, L. Preziosi, Modeling the influence of nucleus elasticity on cell invasion  
959 in fiber networks and microchannels, *J. Theor. Biol.* 317 (2013) 394–406.  
960 doi:10.1016/j.jtbi.2012.11.003.

961

962 [58] A.L. Bauer, T.L. Jackson, Y. Jiang, A cell-based model exhibiting branching and  
963 anastomosis during tumor-induced angiogenesis, *Biophys. J.* 92 (2007) 3105–3121.  
964 doi:10.1529/biophysj.106.101501.

965

966 [59] B.M. Rubenstein, L.J. Kaufman, The Role of Extracellular Matrix in Glioma Invasion:  
967 A Cellular Potts Model Approach, *Biophys J.* 95 (2008) 5661–5680.  
968 doi:10.1529/biophysj.108.140624.

969  
970 [60] A. Shirinifard, J.S. Gens, B.L. Zaitlen, N.J. Popławski, M. Swat, J.A. Glazier, 3D  
971 multi-cell simulation of tumor growth and angiogenesis, PLoS ONE. 4 (2009) e7190.  
972 doi:10.1371/journal.pone.0007190.  
973  
974 [61] S. Turner, J.A. Sherratt, Intercellular adhesion and cancer invasion: a discrete  
975 simulation using the extended Potts model, J. Theor. Biol. 216 (2002) 85–100.  
976 doi:10.1006/jtbi.2001.2522.  
977  
978 [62] D. Drasdo, G. Forgacs, Modeling the interplay of generic and genetic mechanisms in  
979 cleavage, blastulation, and gastrulation, Dev. Dyn. 219 (2000) 182–191. doi:10.1002/1097-  
980 0177(200010)219:2<182::AID-DVDY1040>3.0.CO;2-A.  
981  
982 [63] J. Moreira, A. Deutsch, Pigment pattern formation in zebrafish during late larval  
983 stages: a model based on local interactions, Dev. Dyn. 232 (2005) 33–42.  
984 doi:10.1002/dvdy.20199.  
985  
986 [64] T. Walther, H. Reinsch, A. Grosse, K. Ostermann, A. Deutsch, T. Bley, Mathematical  
987 modeling of regulatory mechanisms in yeast colony development, J. Theor. Biol. 229 (2004)  
988 327–338. doi:10.1016/j.jtbi.2004.04.004.  
989  
990 [65] V.P. Zhdanov, B. Kasemo, Simulation of the growth and differentiation of stem cells  
991 on a heterogeneous scaffold, Physical Chemistry Chemical Physics. 6 (2004) 4347–4350.  
992 doi:10.1039/b407371c.  
993  
994 [66] A.F.M. Marée, P. Hogeweg, How Amoeboids Self-Organize into a Fruiting Body:  
995 Multicellular Coordination in Dictyostelium discoideum, Proceedings of the National  
996 Academy of Sciences of the United States of America. 98 (2001) 3879–3883.  
997  
998 [67] N.J. Savill, J.A. Sherratt, Control of epidermal stem cell clusters by Notch-mediated  
999 lateral induction, Dev. Biol. 258 (2003) 141–153.  
1000  
1001 [68] A. Mochizuki, Pattern formation of the cone mosaic in the zebrafish retina: a cell  
1002 rearrangement model, J. Theor. Biol. 215 (2002) 345–361. doi:10.1006/jtbi.2001.2508.

1003  
1004 [69] P.K. Maini, L. Olsen, J.A. Sherratt, Mathematical models for cell-matrix interactions  
1005 during dermal wound healing, *Int. J. Bifurcation Chaos*. 12 (2002) 2021–2029.  
1006 doi:10.1142/S0218127402005674.  
1007  
1008 [70] M. Scianna, An extended Cellular Potts Model analyzing a wound healing assay,  
1009 *Comput. Biol. Med.* 62 (2015) 33–54. doi:10.1016/j.compbiomed.2015.04.009.  
1010  
1011 [71] N.J. Popławski, A. Shirinifard, M. Swat, J.A. Glazier, SIMULATION OF SINGLE-  
1012 SPECIES BACTERIAL-BIOFILM GROWTH USING THE GLAZIER-GRANER-  
1013 HOGEWEG MODEL AND THE COMPUCELL3D MODELING ENVIRONMENT, *Math*  
1014 *Biosci Eng.* 5 (2008) 355–388.  
1015  
1016 [72] R. Chaturvedi, C. Huang, J.A. Izaguirre, S.A. Newman, J.A. Glazier, M. Alber, A  
1017 Hybrid Discrete-Continuum Model for 3-D Skeletogenesis of the Vertebrate Limb, in: P.M.A.  
1018 Sloot, B. Chopard, A.G. Hoekstra (Eds.), *Cellular Automata*, Springer Berlin Heidelberg,  
1019 2004: pp. 543–552. [http://link.springer.com/chapter/10.1007/978-3-540-30479-1\\_56](http://link.springer.com/chapter/10.1007/978-3-540-30479-1_56)  
1020 (accessed July 29, 2015).  
1021  
1022 [73] R. Chaturvedi, C. Huang, B. Kazmierczak, T. Schneider, J.. Izaguirre, T. Glimm, et  
1023 al., On multiscale approaches to three-dimensional modelling of morphogenesis, *J R Soc*  
1024 *Interface*. 2 (2005) 237–253. doi:10.1098/rsif.2005.0033.  
1025  
1026 [74] N.J. Popławski, M. Swat, J.S. Gens, J.A. Glazier, Adhesion between cells, diffusion of  
1027 growth factors, and elasticity of the AER produce the paddle shape of the chick limb, *Physica*  
1028 *A*. 373 (2007) 521–532. doi:10.1016/j.physa.2006.05.028.  
1029  
1030 [75] C. Xue, A. Friedman, C.K. Sen, A mathematical model of ischemic cutaneous  
1031 wounds, *PNAS*. 106 (2009) 16782–16787. doi:10.1073/pnas.0909115106.  
1032  
1033 [76] Z. Xu, J. Lioi, J. Mu, M.M. Kamocka, X. Liu, D.Z. Chen, et al., A multiscale model of  
1034 venous thrombus formation with surface-mediated control of blood coagulation cascade,  
1035 *Biophys. J.* 98 (2010) 1723–1732. doi:10.1016/j.bpj.2009.12.4331.  
1036



- 1037 [77] Z. Xu, N. Chen, M.M. Kamocka, E.D. Rosen, M. Alber, A multiscale model of  
1038 thrombus development, *J R Soc Interface*. 5 (2008) 705–722. doi:10.1098/rsif.2007.1202.  
1039
- 1040 [78] R.M.H. Merks, J.A. Glazier, Dynamic mechanisms of blood vessel growth,  
1041 *Nonlinearity*. 19 (2006) C1–C10. doi:10.1088/0951-7715/19/1/000.  
1042
- 1043 [79] R.M.H. Merks, S.V. Brodsky, M.S. Goligorksy, S.A. Newman, J.A. Glazier, Cell  
1044 elongation is key to in silico replication of in vitro vasculogenesis and subsequent remodeling,  
1045 *Dev. Biol.* 289 (2006) 44–54. doi:10.1016/j.ydbio.2005.10.003.  
1046
- 1047 [80] R.M.H. Merks, E.D. Perryn, A. Shirinifard, J.A. Glazier, Contact-Inhibited  
1048 Chemotaxis in De Novo and Sprouting Blood-Vessel Growth, *PLoS Comput Biol.* 4 (2008).  
1049 doi:10.1371/journal.pcbi.1000163.  
1050
- 1051 [81] M. Scianna, L. Munaron, L. Preziosi, A multiscale hybrid approach for vasculogenesis  
1052 and related potential blocking therapies, *Prog. Biophys. Mol. Biol.* 106 (2011) 450–462.  
1053 doi:10.1016/j.pbiomolbio.2011.01.004.  
1054
- 1055 [82] M. Scianna, A multiscale hybrid model for pro-angiogenic calcium signals in a  
1056 vascular endothelial cell, *Bull. Math. Biol.* 74 (2012) 1253–1291. doi:10.1007/s11538-011-  
1057 9695-8.  
1058
- 1059 [83] A. Szabo, E.D. Perryn, A. Czirok, Network Formation of Tissue Cells via Preferential  
1060 Attraction to Elongated Structures, *Phys. Rev. Lett.* 98 (2007) 038102.  
1061 doi:10.1103/PhysRevLett.98.038102.  
1062
- 1063 [84] A. Szabó, R. Unnep, E. Méhes, W.O. Twal, W.S. Argraves, Y. Cao, et al., Collective  
1064 cell motion in endothelial monolayers, *Phys Biol.* 7 (2010) 046007. doi:10.1088/1478-  
1065 3975/7/4/046007.  
1066
- 1067 [85] A. Szabó, A. Czirók, The Role of Cell-Cell Adhesion in the Formation of  
1068 Multicellular Sprouts, *Math Model Nat Phenom.* 5 (2010) 106. doi:10.1051/mmnp/20105105.  
1069
- 1070 [86] M. Scianna, R.M.H. Merks, L. Preziosi, E. Medico, Individual cell-based models of

1071 cell scatter of ARO and MLP-29 cells in response to hepatocyte growth factor, *J. Theor. Biol.*  
1072 260 (2009) 151–160. doi:10.1016/j.jtbi.2009.05.017.

1073

1074 [87] M. Scianna, L. Preziosi, K. Wolf, A Cellular Potts Model simulating cell migration on  
1075 and in matrix environments, *Math Biosci Eng.* 10 (2013) 235–261.

1076

1077 [88] J. Starruß, T. Bley, L. Søgaaard-Andersen, A. Deutsch, A New Mechanism for  
1078 Collective Migration in *Myxococcus xanthus*, *J Stat Phys.* 128 (2007) 269–286.

1079 doi:10.1007/s10955-007-9298-9.

1080

1081 [89] A.F.M. Marée, A. Jilkine, A. Dawes, V.A. Grieneisen, L. Edelstein-Keshet,  
1082 Polarization and movement of keratocytes: a multiscale modelling approach, *Bull. Math. Biol.*

1083 68 (2006) 1169–1211. doi:10.1007/s11538-006-9131-7.

1084

1085 [90] R.M.H. Merks, P. Koolwijk, Modeling Morphogenesis *in silico* and *in vitro* :  
1086 Towards Quantitative, Predictive, Cell-based Modeling, *Mathematical Modelling of Natural*  
1087 *Phenomena.* 4 (2009) 149–171. doi:10.1051/mmnp/20094406.

1088

1089 [91] M. Scianna, E. Bassino, L. Munaron, A cellular Potts model analyzing differentiated  
1090 cell behavior during *in vivo* vascularization of a hypoxic tissue, *Comput. Biol. Med.* 63

1091 (2015) 143–156. doi:10.1016/j.combiomed.2015.05.020.

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## 1097 **Figures, Movies and Tables**

1098 Fig. 1 Cell behaviour is determined by a modified Metropolis algorithm, which is based on a  
1099 iterative and stochastic minimization of the cell-matrix system energy, defined by a Hamiltonian  
1100 functional  $H$ . In particular, it includes energetic contributions for cell geometrical attributes and  
1101 cell-substrate adhesive affinity. A Boltzmann-like law finally controls the likelihood of the

1102 acceptance of domain configuration updates, which is further biased by the intrinsic cell motility,  
 1103 established by parameter  $T_C$ .

1104 Fig. 2 Snapshots of the tested substrate configurations: (a) soft (red:  $t_{S_1}$  with  $J_{C,S_1} = 25 \cdot 10^{-15}$   
 1105  $\text{Kg/s}^2$ ) and stiff (yellow:  $t_{S_2}$  with  $J_{C,S_2} = 1 \cdot 10^{-15} \text{Kg/s}^2$ ) substrates, (b) soft (red:  $t_{S_1}$  with  
 1106  $J_{C,S_1} = 25 \cdot 10^{-15} \text{Kg/s}^2$ ) substrate with two stiff (yellow:  $t_{S_2}$  with  $J_{C,S_2} = 1 \cdot 10^{-15} \text{Kg/s}^2$ ) stripes,  
 1107 (c) sequence of stripes with different stiffness (red:  $t_{S_1}$  with  $\text{Kg/s} J_{C,S_1} = 25 \cdot 10^{-15}$ , dark orange:  
 1108  $t_{S_2}$  with  $J_{C,S_2} = 20 \cdot 10^{-15} \text{Kg/s}^2$ , orange:  $t_{S_3}$  with  $J_{C,S_3} = 15 \cdot 10^{-15} \text{Kg/s}^2$ , light orange:  $t_{S_4}$  with  
 1109  $J_{C,S_4} = 10 \cdot 10^{-15} \text{Kg/s}^2$ , dark yellow:  $t_{S_5}$  with  $J_{C,S_5} = 5 \cdot 10^{-15} \text{Kg/s}^2$ , yellow:  $t_{S_6}$  with  $J_{C,S_6} =$   
 1110  $1 \cdot 10^{-15} \text{Kg/s}^2$ ), (d) stiff (yellow:  $t_{S_2}$  with  $J_{C,S_2} = 1 \cdot 10^{-15} \text{Kg/s}^2$ ) substrate with embedded soft  
 1111 (red:  $t_{S_1}$  with  $J_{C,S_1} = 25 \cdot 10^{-15} \text{Kg/s}^2$ ) squares.

1112  
 1113 Fig. 3 Simulation for a substrate with soft (red) and stiff (yellow) regions. As the ratio  $\frac{J_{C,S_1}}{J_{C,S_2}}$ ,  
 1114 increases, the cells are typically biased to migrate towards the stiff region (c). This is also  
 1115 confirmed by the trajectories of the cell centre of mass, which are relatively close to the centre of  
 1116 the substrate when  $\frac{J_{C,S_1}}{J_{C,S_2}} = 1$  (a), whereas they are substantially shifted on the stiff region when

1117 
$$\frac{J_{C,S_1}}{J_{C,S_2}} = 25 \text{ (b)}.$$

1118  
 1119 Fig. 4 Two cells are initially seeded on a soft (red) and a stiff (yellow) substrate, respectively. (a)  
 1120 Simulation snapshot of the final positions (i.e., at MCS = 500 corresponding to nearly 16 minutes)  
 1121 of the two cells. (b) Initial (dashed) and final (plain) contour shapes give an idea of the position  
 1122 and the morphology of the two cells. (c) Cell adhesive area as a function of the type of substrate.  
 1123 The area is about 30% higher in the case of the cell seeded over the stiff substrate, due to the  
 1124 specific constitutive law given to cell rigidity (i.e., Eq. (6)).

1125  
 1126 Fig. 5 Simulation for a substrate with soft (red) and stiff (yellow) subdomains. The trajectories of  
 1127 the cell centre of mass as well as the initial (dashed) and the final (plain) cell contours are traced

1128 respectively for (a) a cell initially seeded at the south-east corner and an external potential  
1129 introduced towards the north-west corner and (b) a cell initially seeded at the south-west corner  
1130 and an external potential directed towards the north-east corner. (c) Cell average velocity over  
1131 either the stiff and soft substrate.

1132

1133 Fig. 6 Configuration with a soft (red) substrate with two embedded stiff stripes (yellow). (a) and  
1134 (c) Simulations with  $|\mathbf{v}_{ext}| = 7 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  and  $|\mathbf{v}_{ext}| = 28 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$  respectively.

1135 Representative cell trajectories are plotted together with the initial (dashed) and the final (plain)  
1136 cell contours. (b) Relative cell frequency as function of  $|\mathbf{v}_{ext}|$ . (d) Cell average velocity over the  
1137 different substrate regions in the case of  $|\mathbf{v}_{ext}| = 28 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ .

1138

1139 Fig. 7 Results for the simulation with a soft to stiff gradient (Sec. 3.4). (a) Relative cell frequency  
1140 as  $|\mathbf{v}_{ext}|$  increases. (b) Average cell velocity over the different substrate regions in case of  $|\mathbf{v}_{ext}| =$   
1141  $28 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ .

1142

1143 Fig. 8 Results for the simulations with a soft to stiff gradient (Sec. 3.5). (a)  $|\mathbf{v}_{ext}|$  necessary to  
1144 allow cells reach the opposite border of the domain vs width of substrate stripes. (b) Average cell  
1145 velocity over the different substrate regions for different widths of the matrix stripes in case of a  
1146 stiffness gradient sufficient fine-grained to have a cell persistent movement even in the absence of  
1147 an external potential.

1148

1149 Fig. 9 Snapshots from a representative simulation dealing with a domain with four soft squares (red) embedded  
1150 in a stiff substrate (yellow). The cell is initially seeded at the south-west corner and migrates in the direction of  
1151 an external potential ( $|\mathbf{v}_{ext}| = 14 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ ), i.e., towards the north-east corner. Snapshots are taken at 2  
1152 min (a), 30 min (b), 1.5 h (c), 2 h (d), 2.5 h (e), 3.5 h (f), 4.5 h (g) and 5.5 h (g).

1153

1154 Fig. 10 Cell adhesive area at 5.5 h, obtained from CPM simulations for different values both of the Boltzmann  
1155 temperature  $T_C$  and of the adhesiveness between the cell and the softest substrate, i.e.,  $J_{soft}$ . Values are given as  
1156 the mean over 100 simulations. The experimental value measured by Lo and co-workers in corresponding

1157 conditions is  $1.74 \cdot 10^3 \mu\text{m}^2$ . From this plot, it was indeed possible to observe that the parameter region pseudo-  
1158 colored in yellow gave the best fitting couples of coefficients. In particular, we opted for  $T_C = 50 \cdot$   
1159  $10^{-27} \text{ Kg m}^2/\text{s}^2$  and  $J_{\text{soft}} = 25 \cdot 10^{-15} \text{ Kg/s}^2$ .

1160

1161 Movie 1 Simulation of cell migration over a stiff-soft substrate (yellow = stiff region, red = soft  
1162 region) in presence of an external potential directed towards the north-west corner (Sec. 3.2). The  
1163 cell is initially seeded at the south-east corner.

1164

1165 Movie 2. Simulation of cell migration over a stiff-soft substrate (yellow = stiff region, red = soft  
1166 region) in presence of an external potential directed towards the north-east corner (Sec. 3.2). The  
1167 cell is initially seeded at the south-west corner.

1168

1169 Movie 3. Simulation of cell migration over a soft substrate (red) including two parallel stiff stripes  
1170 (yellow) in presence of an external potential ( $|\mathbf{v}_{ext}| = 7 \cdot 10^{-21} \text{ Kg} \cdot \text{m/s}^2$ ) directed towards the north-  
1171 east corner (Sec. 3.3). The cell is initially seeded at the south-west corner.

1172

1173 Movie 4. Simulation of cell migration over a soft substrate (red) including two parallel stiff stripes  
1174 (yellow) in presence of an external potential ( $|\mathbf{v}_{ext}| = 28 \cdot 10^{-21} \text{ Kg} \cdot \text{m/s}^2$ ) directed towards the  
1175 north-east corner (Sec. 3.3). The cell is initially seeded at the south-west corner.

1176

1177 Movie 5. Simulation of cell migration over a substrate with a soft-to-stiff gradient from the left to  
1178 the right part of the domain (the cell-substrate adhesive affinities vary from  $J_{\text{soft}} = 25 \cdot 10^{-15} \text{ Kg/s}^2$   
1179 to  $J_{\text{stiff}} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ) (Sec. 3.4). A low external potential ( $|\mathbf{v}_{ext}| = 7 \cdot 10^{-21} \text{ Kg} \cdot \text{m/s}^2$ ) is directed  
1180 towards the north-east corner of the substrate, whereas the cell is seeded at the south-west corner.

1181

1182 Movie 6. Simulation of cell migration over a substrate with a soft-to-stiff gradient from the left to  
1183 the right part of the domain (the cell-substrate adhesive affinities vary from  $J_{\text{soft}} = 25 \cdot 10^{-15} \text{ Kg/s}^2$   
1184 to  $J_{\text{stiff}} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ) (Sec. 3.4). A high external potential ( $|\mathbf{v}_{ext}| = 28 \cdot 10^{-21} \text{ Kg} \cdot \text{m/s}^2$ ) is

1185 directed towards the north-east corner of the substrate, whereas the cell is seeded at the south-west  
1186 corner.

1187

1188 Movie 7. Simulation of cell migration over a substrate with a soft-to-stiff gradient from the left to the right part  
1189 of the domain (the cell-substrate adhesive affinities vary from  $J_{soft} = 25 \cdot 10^{-15} \text{ Kg/s}^2$  to  $J_{stiff} = 1 \cdot 10^{-15} \text{ Kg/s}^2$ ).  
1190 Here, the width of the stripes has been highly reduced (i.e., = 28  $\mu\text{m}$ ) and no external potential is introduced  
1191 (Sec. 3.5).

1192

1193 Movie 8. Simulation of cell migration over a stiff substrate (yellow) with four embedded soft  
1194 squares (red) (Sec. 3.6). The cell is initially seeded at the south-west corner and an external  
1195 potential ( $|\mathbf{v}_{ext}| = 14 \cdot 10^{-21} \text{ Kg}\cdot\text{m/s}^2$ ) is directed towards the north-east corner of the domain.

1196

1197 Table 1 Main parameters of the model.