POLITECNICO DI TORINO Repository ISTITUZIONALE

Stem cell-based therapy in periodontal regeneration: a systematic review and meta-analysis of randomized clinical studies

Original

Stem cell-based therapy in periodontal regeneration: a systematic

review and meta-analysis of randomized clinical studies / Campagna, Alessandro; Baima, Giacomo; Romano, Federica; Amoroso, Federico; Mussano, Federico; Oteri, Giacomo; Aimetti, Mario; Peditto, Matteo. - In: DENTISTRY JOURNAL. - ISSN 2304-6767. - (2024). [10.3390/dj12050145]

Availability: This version is available at: 11583/2988112 since: 2024-05-16T12:49:06Z

Publisher: Mdpi

Published DOI:10.3390/dj12050145

Terms of use:

This article is made available under terms and conditions as specified in the corresponding bibliographic description in the repository

Publisher copyright Wiley preprint/submitted version

This is the pre-peer reviewed version of the [above quoted article], which has been published in final form at http://dx.doi.org/10.3390/dj12050145.This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Use of Self-Archived Versions.

(Article begins on next page)





1

2

3

4

5

6

7 8

9

10

11

12

Systematic Review

Orally-derived stem cell-based therapy in periodontal regeneration: a systematic review and meta-analysis of randomized clinical studies

Alessandro Campagna^{1,2}, Giacomo Baima², Federica Romano², Federico Amoroso^{2,3}, Federico Mussano², Giacomo Oteri¹, Mario Aimetti²⁺ and Matteo Peditto¹⁺

- ¹ Department of Biomedical and Dental Sciences and Morphofunctional Imaging; University of Messina, 98122, Messina, Italy.
 - Department of Surgical Sciences; C.I.R. Dental School, University of Turin, 10126, Torino, Italy.
- ³ Politecnico di Torino; Turin; Italy.
- Correspondence: federico.mussano@unito.it
- + These authors contributed equally to this work.

Abstract: The present systematic review was performed study the application in the periodontal 13 regenerative therapy of the orally-derived stem cells, because of this, the following PICO question 14was proposed: "In patients with periodontitis, can the adjunctive use of orally-derived stem cells 15 provide additional clinical and radiographic benefits for periodontal regeneration?" Randomized 16 clinical studies were electronically and manually searched up to December 2023. Quantitative anal-17 yses were performed with the aim to evaluate mean differences (MD) between the treatment and 18 control groups in terms of clinical attachment level (CAL) gain, probing pocket depth (PPD) reduc-19 tion, gingival recession (GR), and radiographic bone gain (RBG) using random effect models. A total 20 of 7 studies were selected for the systematic review. Meta-analyses excluding studies with high risk 21 of bias highlighted a non-statistically significant result for the use of stem cells compared to the 22 control groups in terms of CAL gain [MD = 1.05; 95% CI (-0.88, 2.97) p = 0.29] and PPD reduction 23 [MD = 1.32; 95% CI (-0.25, 2.88) p = 0.10]. The same also applied to GR [MD = -0.08; 95% CI (-0.79, 24 0.63) p = 0.83] and RBG [MD = 0.50; 95% CI (-0.88, 1.88) p = 0.48]. Based on high heterogeneity, there 25 is not enough evidence to consider the adjunctive application of orally-derived mesenchymal stem 26 cells as a preferential approach for periodontal regenerative treatment compared to standard pro-27 cedures. 28

Keywords: periodontitis; periodontal regeneration; stem cells; biomaterials.

30

31

29

1. Introduction

Periodontitis is a biofilm-mediated disease with an important inflammatory compo-32 nent which causes the progressive breakdown of the supporting periodontal tissues [1– 33 3]. The objectives of the first steps of periodontal therapy are the control of the microbial 34 infection and the resolution of the inflammation, which clinically refers to the absence of 35 bleeding on probing (BoP) and presence of shallow probing pocket depths (PPD \leq 4 mm) 36 [4-6]. However, residual pockets often persist after non-surgical treatment, especially in 37 sites with important furcation involvement (FI) and/or deep intrabony defects [7,8]. After 38 the XVI European Workshop in Periodontology, there is a strong recommendation to treat 39 dental elements with deep residual PPD associated with intrabony defects of \geq 3 mm with 40 periodontal regenerative surgery [9–11]. Similarly, the recommendation was expressed 41 for the treatment of class II maxillary and mandibular molars [9]. As reported in the liter-42 ature, the various surgical techniques and biomaterials developed in the last 30-40 years 43 with the aim of predictable periodontal regeneration have achieved variable success [12– 44

14]. The benefits reported are often limited to deep intrabony defects and class II mandib-45 ular FI, while supracrestal defects, non-containing intrabony defects and maxillary class 46 II or III FI still have a less predictable outcome [15–18]. For this reason, new tissue engi-47 neering strategies are being sought, and the implementation of innovative techniques us-48 ing orally-derived stem cells is growing in terms of scientific research in periodontology 49 [19,20]. 50

Compared to biomaterials, which are scaffolds characterized by unique chemical, 51 mechanical and biological properties, mainly osteoinductivity and osteoconductivity [21], 52 cell therapy relies on replenishing and/or empowering the inner healing body potential 53 [22,23]. In recent years, cell regeneration therapy has been introduced in many areas of 54 medicine, such as cardiology, neurology or traumatology [24,25], as well as for the treat-55 ment of orofacial dystrophies, diabetic problems and autoimmune diseases [26,27]. Re-56 generative medicine commonly employs stem cells, particularly mesenchymal stem cells 57 (MSCs), which possess unique faculties like self-renewal, clonality, and potency. These 58 adult stem cells exhibit anti-inflammatory properties and contribute to tissue repair pro-59 cesses, secreting mediators with various beneficial effects [28,29]. The expression of spe-60 cific surface antigens, including CD44, CD73, CD29, CD90, and CD105, helps characterize 61 MSCs, while lacking certain hematopoietic and endothelial markers [30]. Indeed, MSCs 62 are defined by their plastic adherence, capacity of self-renovation, and the potential for 63 differentiation in vitro into different types of cells, like osteoblasts, adipocytes, and chon-64 droblasts under specific stimuli [31,32]. Many intraoral and dental sources of MSCs are 65 available, for example dental pulp, periodontal ligament, bone marrow from alveolar 66 bone, dental follicle, gingival connective tissue or apical papilla [33–36]. In virtue of their 67 self-renewal, multipotentiality, immunomodulation, and tissue regeneration capacities, 68 MSCs can promote the growth of various periodontal tissues, like alveolar bone, root ce-69 mentum and periodontal ligament, even in situation with low intrinsic potential [23,37– 70 41]. A recent study assessed periodontal regenerative approaches in animal models, ob-71 serving that mesenchymal stem cells used alone or mixed with other biomaterials, such 72 as bovine bone, beta-tricalcium phosphate (β -TCP), or platelet-rich plasma (PRP), offered 73 better regenerative outcomes than those of the group with biomaterials alone [42]. Most 74 preclinical studies have indeed supported the biological rationale of employing MSCs to 75 promote osteoinduction and tenogenesis, while decreasing inflammation [26,43-45]. 76

In humans, recent systematic reviews evaluated the clinical results of periodontal re-77 generation by MSCs derived from different sources [46,47], reporting a significant ad-78 vantage of using cell therapy in terms of final outcomes. However, due to the presence of 79 highly heterogeneous results and the detection of methodological inconsistencies in data 80 handling, the purpose of the present systematic review was to elucidate through a meta-81 analytic approach the adjunctive clinical and radiographic effect of using orally-derived 82 stem cells for periodontal regeneration. 83

2. Materials and Methods

A systematic review protocol was written in the planning stages and registered on 85 the International prospective register of systematic reviews (PROSPERO; 86 CRD42024525702). The PRISMA statement was followed in both the planning and report-87 ing of the review [48]. 88

2.1. Focused question

This systematic review aimed to answer the following PICO question: "In patients 90 with periodontitis, can the adjunctive use of orally-derived stem cells provide additional 91 clinical benefits measured as clinical attachment level (CAL) gain, probing pocket depth 92 (PPD) reduction, recession (GR) and radiographic bone gain (RBG) for periodontal regen-93 eration procedures?" 94

2.2. Eligibility criteria

89

84

In the present systematic review, the criteria used to select the clinical studies were	96
based on the PICOS method and were the following:	97
• (P) Population: Adult patients with stage III-IV periodontitis presenting with	98
residual pockets and intra-bony defects with at least 3 mm of intra-bony	99
component after the completion of steps I-II of periodontal therapy (causal-	100
related therapy; supra- and sub-gingival instrumentation) [9];	101
• (I) Intervention: Periodontal regeneration with the use of orally-derived stem	102
cells;	103
• (C) Comparison: All other strategies for periodontal regeneration;	104
• (O) Outcome measures:	105
Primary outcomes: CAL gain, and PPD reduction.	106
Secondary outcomes: GR and RBG.	107
• (S) Types of studies: Only randomized controlled clinical trials (RCTs) were	108
considered.	109
The following additional inclusion criteria were applied:	110
English language;	111
• At least 6 months of follow-up;	112
These exclusion criteria were also applied to the selection process:	113
 Lack of pretreatment and post-treatment outcome measures 	114
• Case reports, case series, retrospective studies, animal studies, in vitro stud-	115
ies.	116
2.3. Search strategy	117
The search was conducted through various sources, both electronically and manu-	118

ally. The electronic research included Medline (PubMed), Scopus, and CENTRAL 119 (Cochrane Central Register of Controlled Trials) databases. All articles published until 120 December 2023 were searched adopting the strategy reported in Table 1. A screening of 121 the reference lists of the included studies and related reviews was also carried out to iden-122 tify any additional article of relevance. Hand search was also implemented of the follow-123 ing journals by the authors: Journal of Clinical Periodontology, Journal of Dental Research, 124 Journal of Periodontology, and Journal of Periodontal Research. 125

Table 1. – Search strategy.

(Periodontal defect OR periodontal lesion OR periodontal osseous defect OR intraosseous defect OR intra-osseous defect OR intrabony defect OR infra-bony defect OR angular defect OR bony defect OR osseous defect OR crater) AND (stem cells OR stem OR stem cell therapy OR cell therapy OR MSC OR mesenchymal stem cells OR human cord stem cells OR BMMSC OR bone marrow mesenchymal stem cell OR pluripotent stem cells OR embryonic stem cells OR ESC OR cell technology OR oral stem cells OR stem cell-delivery therapeutics OR induced pluripotent stem cells OR iPSC OR adiposederived stem cells OR dental stem cells OR pulp stem cells OR periodontal ligament stem cells OR PDLSC OR progenitor cells OR apical papilla stem cells OR dental follicle stem cells OR human exfoliated deciduous tooth cells) AND (clinical trial OR case series OR prospective study OR longitudinal study OR cohort study OR RCT OR randomized clinical trial) AND (GTR OR guided tissue regeneration OR periodontal regeneration)

2.4. Study selection

The results obtained from the manual search and from the various electronic data-129 base were downloaded and imported jointly into a reference management software, and duplicates or non-English language articles were automatically removed. The identified 131

126

127 128

articles were checked based on the pre-defined eligibility criteria. During the initial phase 132 the screening of potentially suitable titles and abstracts was performed: abstracts that at 133 this time met the inclusion criteria or did not provide sufficient information were admit-134 ted for the subsequent review phase. Once the eligible articles were defined, they were re-135 evaluated after reading the full-text by applying the selection criteria again. The studies 136 that satisfied all the inclusion criteria were included in the systematic review. Two review-137 ers (A.C. and M.P.) evaluated the abstracts, titles and full text for selection, and when 138 differences occurred, they were solved by discussion with a third party (G.O.). 139

2.5.	Data	extraction	for	analysis

The relevant data identified in the included studies were reported in a standardized 141 extraction form, including the following: 142

, 0 0	
 Author(s) and year of publication; 	143
 Number of patients included in the study; 	144
 Number of defects treated in both the test and control groups; 	145
 Type of stem cells used in the test group; 	146
 Type of bone defect treated; 	147
 Type of treatment of the test group; 	148
 Type of treatment of the control group; 	149
CAL gain;	150
PPD reduction;	151
• GR;	152
• RBG;	153
Study duration.	154
2.6. Risk of bias of individual studies	155
The quality <mark>evaluation</mark> of the <mark>selected</mark> studies was <mark>independently</mark> performed by two	156
review authors (A.C. and M.P.) through risk of bias analysis as it could impact on the	157
overall results and conclusions. The Cochrane Collaboration's tool was used for assessing	158
risk of bias [49,50]. We considered seven domains (sequence generation, allocation con-	159
cealment, blinding of the outcome assessor, blinding of participants and personnel, in-	160

complete outcome data, selective outcome reporting and other bias) and included in a specific table the results of the assessment. Then, the complessive risk of bias in the included studies was categorized as below:

A: Low risk of bias: little chance that bias would significantly affect the outcomes if all criteria were fulfilled; _______ 165

B: Unclear risk of bias: possibility of bias that casts some doubt on the outcomes if one or more criteria were only partially met; ________167

C: High risk of bias: likelihood of bias that substantially undermines confidence in the outcomes if one or more criteria were not met. 169

2.7. Statistical analysis

Studies were firstly summarized in a narrative form by key characteristics and ac-171 cording to type of regenerative surgery. A meta-analysis was carried out in the presence 172 of at least two studies of similar design. The variables were registered at patient level. In 173 each patient, only one tooth per technique was assessed. Weighted mean differences (MD) 174and 95% confidence intervals (95% CI) were calculated for CAL gain, PPD reduction, GR 175 reduction [51], and RBG using the generic inverse variance method. Forest plots were 176 graphically depicted to summarize the difference in outcomes between the groups using 177 the patient as the analysis unit. 178

We used the χ^2 test to assess the statistical heterogeneity among the different studies179and the percentage of variation in the global estimate due to heterogeneity was calculated180using I² index (25%: low; 50%: moderate; 75% high). [49] In case of values higher than 50%,181the random effect method was applied. Results were considered statistically significant182

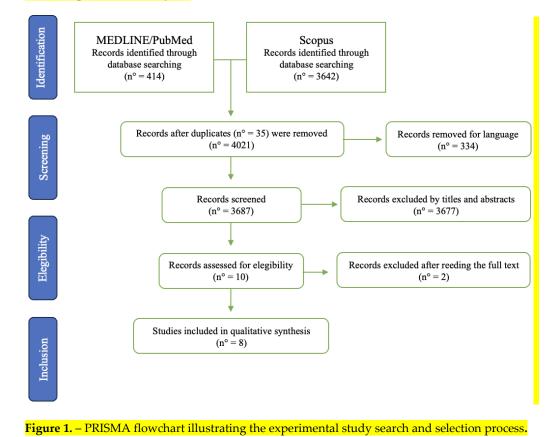
140

for p values < 0.05. Statistical analyses were carried out using the RevMan software ver-183 sion 5.4 (Copenhagen: The Nordic Cochrane Centre, The Cochrane Collaboration, 2014). 184

3. Results

3.1. Study selection

The selection process was conducted according to the PRISMA guideline (Figure 1). 187 The search on the MEDLINE/PubMed, Scopus and Cochrane databases provided a total 188 of 4086 studies; there were 65 duplicates, while articles discarded for non-English lan-189 guage were 334. A number of 3687 studies were screened and, of these, 3678 were ex-190 cluded after first-stage reading of titles and abstracts due to the type of publication (chap-191 ter of book or thesis), objective and/or design of the study. Two articles were removed 192 after full-text reading. Finally, 7 articles met all the inclusion criteria and were included 193 into the qualitative analysis. 194



3.2. Risk of bias

Out of the 7 included RCTs, 2 were not included in the meta-analysis because were 198 rated at high risk of bias [52,53]. Of the remaining 5 studies, 3 were considered as unclear 199 risk of bias and only 2 as low risk of bias (Figure 2) [19,54–57]. The lack of blinding of the 200 outcome assessor, among the seven domains, was the most frequent source of bias.

201

195

196

197

202

185

Complessive evaluation	Unclear Included	Unclear Included	Low Included	Low Included	High Excluded	Unclear Included	High Excluded
Incomplete outcome data	¢	+	+	+	X	+	X
Blinding of outcome assessment	×	-	+	t	2	×	X
Blinding of participants and personnel	~	×	+	t	2	*	X
Other bias	Ŧ	t	t	t	+	+	X
Selective reporting	+	+	+	+	+	+	×
Allocation concealment	t		t	-	Z	+	2
Random sequence generation	t	Ŧ	t	Ŧ	+	t	+
Study	Abdal-Wahab et al. (2020)	Apatzidou et al. (2021)	Chen et al. (2016)	Ferrarotti et al. (2018)	Hernández- Monjaraz et al. (2020)	Sánchez et al. (2020)	Shalini & Vandana (2018)

Figure 2. – Assessment of the risk of bias in the included studies.

3.3. Study characteristics

Data extracted from the RCTs included in the review are presented in Table 2. There was a certain heterogeneity about the specific type of stem cells used in the control groups between the 5 studies included in the meta-analysis. Indeed, 2 studies [19,57] used periodontal ligament stem cells (PDLSCs), 1 study [56] applied dental pulp stem cells (DPSCs), 209 1 study [55] used bone marrow mesenchymal stem cells extracted from alveolar bone (AB-MMSCs) and 1 study [54] gingival mesenchymal stem cells (GMSCs). The follow-up lasted 6 months in 1 study [54] and 12 months in 4 studies [19,55–57]. 212

- 203
- 204
- 205

Abdal-Wahab and colleagues [54] included a total of 20 patients, excluding current 213 smokers. A full-mouth non-surgical periodontal therapy was performed in all selected 214 patients and, then, they were randomly assigned in the following test or control groups: 215

- Test: 10 intrabony periodontal defects were treated with GMSCs associated 216 with a β-TCP scaffold and a collagen membrane. 217
- Control: 10 intrabony periodontal defects treated with β-TCP and collagen 218 membrane alone.
 219

Apatzidou and colleagues [55] included 27 patients which were allocated in the following 3 groups: 221

- Test: 9 intrabony defects were treated with ABMMSCs embedded on a collagen scaffold enriched with a fibrin lysate and autologous platelets, using the minimally invasive surgical technique (MIST) [9].
- Control B: 10 intrabony defects were treated using MIST with only the collagen scaffold enriched with fibrin lysate and autologous platelets.

• Control C: 8 intrabony defects were treated with the MIST technique alone.

Chen and colleagues [19] selected 30 patients, randomly assigned to one of the two groups: 228

- Test: 20 intrabony defects treated with heterologous bone graft and the adjunctive use of PDLSCs 231
- Control: 21 intrabony defects treated with heterologous bone graft only.

Ferrarotti and colleagues [56] enrolled 29 patients with severe periodontitis, randomly assigning them to one of two groups: 234

- Test: 15 intrabony defects accessed with the MIST technique and treated with 235 DPSCs soaked on a collagen sponge. 236
- Control: 14 intrabony defects treated with only insertion of collagen sponge 237 using MIST technique. 238

Sánchez and colleagues [57] included a total of 20 patients. After initial periodontal239therapy, the subjects were placed in one of two groups with a quasi-randomized approach, i.e. the patients assigned to the treatment group have previously obtained successful in vitro stem cell expansion process:240241242

- Test: 10 intrabony defects treated with PDLSCs together with a heterologous 243 bone substitute. 244
- Control: 10 intrabony defects treated with heterologous bone substitute 245 alone. 246

Table 2. Summary of studies included in the systematic review.

Study	MSC type	Defect inclusion criteria	Group characteristics		oup characteristics Number of patients		Number of defects		Primary outcomes
			Test	Control	Test	Control	Test	Control	
Apatzidou et al. 2021	mesenchy mal stem cells	wall defects	ABMMSCs + autologous fibrin/platel et lysate	Group B: autologous fibrin/platele	9	10 + 8	9	Group B: 10 Group C: 8	CAL; PPD; GR; BDD; BC-BD
	,	in test group		101101					

247

225

226

227

		and 10 in							
		control groups)							
Sanchez et al. 2020	us periodont al ligament- derived mesenchy mal stem cells	Infrabony defect (3 1- wall defects in test group; 7 2- wall defects in test group and 10 in control	PDLSCs + bone xenograft	Bone xenograft	10	10	10	10	CAL; PPD; GR
Abdal-Wahab et al. 2020	us gingival associated mesenchy mal stem cells (GMSCs)	Infrabony defect (7 2- wall defects in test group and 6 in control group; 3 3- wall defects in test group and 4 in control group)	GMSC + (beta- tricalcium phosphate (β-TCP) + collagen membrane	Beta- tricalcium phosphate (β-TCP) + collagen membrane	10	10	10	10	CAL; PPD
Hernández- Monjaraz et al. 2020		Infrahony	DPSCs + collagen scaffold	Collagen scaffold	11	10	11	10	PPD
Ferrarotti et al. 2018	Autologo us dental pulp stem cells (DPSCs)	Infrabony defect (7 1- wall defects in test group and 5 in control group; 4 2- wall defects in test group and 5 in control group; 4 3- wall defects in test group and 4 in control group)	DPSCs + MIST + collagen sponge	MIST + collagen sponge	15	14	15	14	CAL; PPD; GR; BC-BD
Shalini & Vandana 2018	Autologo us periodont al	Infrabony	OFD + PDLSCs	OFD	14	14	14	14	CAL, PPD

ligament- derived mesenchy mal stem cells (PDLSCs)			
Autologo us periodont Infrabony al defect Chen et al. ligament- (Defects 2016 derived characteristi mesenchy c not mal stem mentioned) cells (PDLSCs)	OSS	20	CAL; PPD; 21 GR; BDD

Legend: CAL: clinical attachment levels; PPD: probing pocket depth; GR: gingival recession; BDD:248linear distance from cementoenamel junction to bottom of defect; BC-BD: linear distance from bone249crest to bottom of defect; MIST: minimally invasive surgical technique; GTR: guided tissue regener-250ation; OFD: open flap debridement.251

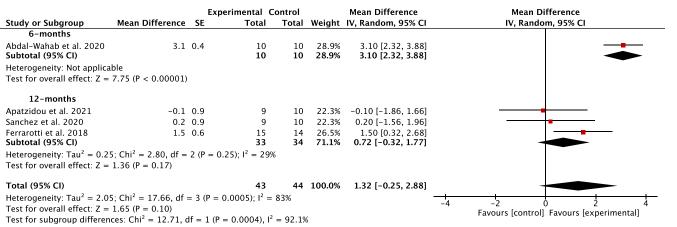
3.4. Results of the analyses

The results of individual studies as they relate with the main outcomes are reported 253 in Table S1. The mean improvements reported in the meta-analyses for the different study outcomes can be summarized as follows: 255

- CAL gain: A total of 4 studies [54–57] compared post-operative CAL gain256with a minimum of 6-month follow up between the test and control groups.257Very high heterogeneity was encountered between the groups (P < 0.001; I^2 258= 90%). The meta-analysis conducted using a random-effect model revealed259a non-statistically significant improvement in the test group [MD = 1.05; 95%260CI (-0.88, 2.97) p = 0.29] (Figure 3).261
- PPD reduction: A total of 4 studies [54–57] compared post-operative PPD reduction with a minimum of 6-month follow up between the experimental 263 group and the control group. There was high heterogeneity between the 264 groups (P < 0.001; I² = 83%). A non-statistically significant adjunctive improvement in PPD reduction in the experimental group [MD = 1.32; 95% CI 266 (-0.25, 2.88) p = 0.10] was shown by the meta-analysis (Figure 4).
- GR: 3 studies [55–57] compared GR between the test and the control arm with 268 a 12-month follow-up. The meta-analysis results displayed low heterogeneity between the groups (P = 0.37; $I^2 = 0\%$), so using a random effect model 270 they revealed a non-statistically significant difference between the test and 271 control groups [MD = -0.08; 95% CI (-0.79, 0.63) p = 0.83] (Figure 5). 272
- RBG: A total of 3 studies [19,55,56] compared RGB between the test group 273 and the control group. The heterogeneity was high between the groups (p < 274 0.01; I² = 84%). There was no statistically significant difference in RBG between test and control groups [MD = 0.50; 95% CI (-0.88, 1.88) p = 0.48] (Figure 6).

			Experimental Co	ontrol		Mean Difference	Mean Difference
Study or Subgroup	Mean Difference	SE	Total	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
6-months							
Abdal-Wahab et al. 2020	2.9	0.5	10	10	26.3%	2.90 [1.92, 3.88]	_
Subtotal (95% CI)			10	10	26.3%	2.90 [1.92, 3.88]	
Heterogeneity: Not applica	able						
Test for overall effect: Z =	5.80 (P < 0.00001)						
12-months							
Apatzidou et al. 2021	-0.9	0.5	9	10	26.3%	-0.90 [-1.88, 0.08]	
Ferrarotti et al. 2018	1.6	0.8	15	14	23.7%	1.60 [0.03, 3.17]	_
Sanchez et al. 2020	0.6	0.8	9	10	23.7%	0.60 [-0.97, 2.17]	
Subtotal (95% CI)			33	34	73.7%	0.33 [-1.21, 1.88]	
Heterogeneity: $Tau^2 = 1.3$	8; Chi ² = 7.80, df =	2 (P	$= 0.02$; $I^2 = 74\%$				
Test for overall effect: $Z =$	0.42 (P = 0.67)						
Total (95% CI)			43	44	100.0%	1.05 [-0.88, 2.97]	
Heterogeneity: $Tau^2 = 3.43$	2; $Chi^2 = 29.68$, df	= 3 (F	$P < 0.00001$); $I^2 =$	= 90%			- <u>t</u> ttt
Test for overall effect: Z =							-4 -2 0 2 4
Test for subgroup differen		= 1	$(P = 0.006), I^2 = 3$	86.7%			Favours [control] Favours [experimental]

Figure 3. - Comparison between the results of studies comparing periodontal regeneration with or 279 without the adjunctive use of orally-derived stem cells in terms of clinical attachment level (CAL) 280 gain. 281



282

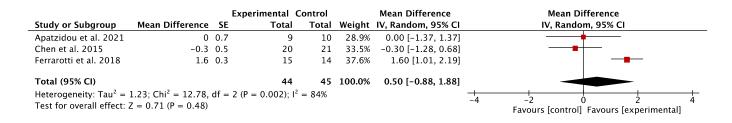
278

Figure 4. - Comparison between the results of studies comparing periodontal regeneration with or 283 without the adjunctive use of orally-derived stem cells in terms of probing pocket depth (PPD) re-284 duction. 285

Study or Subgroup	Mean Difference		Experimental C Total		Weight	Mean Difference IV, Random, 95% CI	Mean Difference IV, Random, 95% Cl
Apatzidou et al. 2021	0.6	0.7	9	10	26.8%	0.60 [-0.77, 1.97]	
Ferrarotti et al. 2018	-0.1	0.5	15	14	52.6%	-0.10 [-1.08, 0.88]	_
Sanchez et al. 2020	-0.9	0.8	0	0	20.6%	-0.90 [-2.47, 0.67]	
Total (95% CI)			24	24	100.0%	-0.08 [-0.79, 0.63]	
Heterogeneity: Tau ² = 0 Test for overall effect: 2	f = 2	$P = 0.37$; $I^2 =$	0%			-2 -1 0 1 2 Favours [control] Favours [experimental]	

²⁸⁶

Figure 5. - Comparison between the results of studies comparing periodontal regeneration with or 287 without the adjunctive use of orally-derived stem cells in terms of gingival recession (GR). 288



289

292

Figure 6. – Comparison between the results of studies comparing periodontal regeneration with or 290 without the adjunctive use of orally-derived stem cells in terms of radiographical bone gain (RBG). 291

4. Discussion

293 294 295

The objective of this systematic literature review was to assess the clinical and therapeutic effectiveness of regenerative periodontal treatment when orally-derived stem cells are used as adjunctive therapy by selecting RCTs. To date, MSCs can be isolated from diverse sources in the oral cavity, with dento-periodontal derived stem cells seeming the 296 best candidates for periodontal tissue regeneration [37,39,58]. Therefore, the focus ques-297 tion of this systematic review was: "In patients with periodontitis, can the adjunctive use of 298 orally-derived MSCs provide additional clinical benefits measured as CAL gain, PPD reduction, 299 GR, and RBG for periodontal regeneration?". A total of 7 RCTs including a total of 186 pa-300 tients were included after screening. Risk of bias test led to the exclusion of 2 RTCs for the 301 meta-analyses. The overall findings showed a lack of significant benefits in the adjunctive 302 use of orally-derived MSCs at 12 months during periodontal regeneration procedures. 303 Heterogeneity in methodology, study design, and outcomes was high. 304

When considering the primary outcome CAL gain, 4 studies were selected. The con-305 fidence interval of the data relating to the study by Apatzidou et al. and the study by 306 Sánchez et al. exceeded the vertical line of the reference value, so there was no statistically 307 significant difference between the test and control groups. Conversely, Ferrarotti et al. and 308 Abdal-Wahab et al. provided a statistically significant advantage for the adjunctive appli-309 cation of MSCs both at 6- and 12-month follow-up. When the studies were combined, the 310 final meta-analysis revealed a non-statistically significant improvement in CAL gain for 311 the test group of 1.05 mm [95% CI (-0.88, 2.97) p = 0.29]. For PPD reduction, the same 4 312 studies were selected [54–57], with the overall result of the meta-analysis being not statis-313 tically significant at 12 months, but with a significant advantage in the study at 6 months 314 [54]. This finding may suggest a greater rapidity of the periodontal regeneration process 315 following the use of MSCs, although in the long term the results of regenerative surgical 316 treatment appear to be comparable to those of other regenerative methods. When delving 317 deeper into study characteristics, Ferrarotti et al. [56] showed the highest difference in 318 outcome measures between test and control group with respect to other included studies 319 with low risk of bias. This discrepancy can be both ascribed to (i) the use of DPSCs and 320 (ii) the nature of the regenerative procedure in the control group. Indeed, DPSCs hold 321 significant promise due to their accessibility, shared origin, and similar antigenic pattern 322 with PDLSCs, making them particularly attractive for therapeutic applications [33]. 323 DPSCs exhibit an extended lifespan, display compatibility with biomaterials, and can be 324 safely preserved through cryopreservation methods [59]. Building on this foundation, ex-325 perimental findings from studies conducted in vivo and in animal models suggest that 326 DPSCs have the capability to produce lamellar bone with proper vascularization. Moreo-327 ver, DPSCs demonstrate the potential for differentiation into various periodontal tissues, 328 emphasizing their versatility and potential therapeutic efficacy in the field of periodontal 329 regeneration [59,60]. Regarding PDLSCs, they can be found both on the root and alveolar 330 bone surfaces after tooth extraction, although those on the root demonstrate superior dif-331 ferentiation capabilities [61]. Recognized for their safety and efficacy, they became the pi-332 oneering treatment in periodontal regeneration therapy [35,41,62]. Indeed, PDLSCs 333 exhibited the ability to differentiate into mesenchymal cell lineages, generating cells capa-334 ble of forming collagen, adipocytes, cementum tissue, Sharpey's fibers, and osteoblast-like 335 cells in vivo [63]. However, translatability of PDLSCs to the clinics has been hindered by 336 several limitations, including the necessity for tooth extraction and the possibility that the 337 chronic exposure to a chronic inflammatory environment could lead to the depletion of 338 their potential through senescence [36]. Finally, bone marrow-derived mesenchymal stem 339 cells are a specific type of multipotent MSCs that can be obtained from the alveolar bone 340 during surgery, proving comparable biologic features to iliac BMMSCs [64]. They have 341 also shown the potential of inducing not only reconstruction of bone, but also periodontal 342 and dental tissue regeneration in preclinical models. Indeed, they have the ability to in-343 crease the expression of genes related to tooth development, and they can transform into 344 cells resembling ameloblasts and periodontal tissue cells [65]. Lastly, although presenting 345 a biological rationale to hypothesize their use [66], no study was found testing the appli-346 cation of adipose-derived stem cells in periodontal tissue regeneration. 347

Three recent systematic reviews are present in literature focusing on this topic 348 [46,47,67], with their results and conclusion disagreeing substantially from the present 349 study. Indeed, their meta-analyses revealed significant differences between the experi-350 mental and control groups in terms of PPD, CAL, radiographic intrabony defect depth, 351 and GR, emphasizing how the use of MSCs can be beneficial in periodontal regeneration. 352 In contrast to these optimistic trends, the present systematic review revealed an overall 353 lack of significant benefits at 12 months. Notably, the observed heterogeneity in method-354 ology, study design, and measured outcomes was consistently pronounced. Indeed, in a 355 plausible attempt to broaden the focus, previous systematic review combined studies us-356 ing MSCs derived from diverse body sources (such as umbilical stem cells) for different 357 oral surgical interventions (i.e., alveolar bone reconstruction) at different time-points (3, 358 6, and 12 months). Indeed, the inclusion of multiple follow-up groups from the same RCT 359 may lead to excessive weight in the meta-analyses. Overall conclusions cannot overlook 360 these important heterogeneities, in order to provide a clear snapshot of the state-of-the-361 art and guide future research endeavors. This raises important considerations about the 362 standardization of protocols and the need for more homogeneity in future research en-363 deavors to elucidate the specific conditions under which orally-derived MSCs may or may 364 not be effective in enhancing periodontal regeneration. 365

The attention towards the use of MSCs in periodontal therapy derives from the need 366 to implement treatment options for lesions resulting from periodontitis, due to its preva-367 lence globally [68]. In recent years, MSCs have achieved increasing success in the treat-368 ment of many pathologies studied by various branches of medicine, based on their regen-369 erative and immunoregulatory properties. Our knowledge is still limited, and this means 370 that the prospects regarding their clinical use are very broad. It should be considered that 371 there are critical steps to increase the frequency of stem cell-based therapeutic approaches. 372 In fact, it must be said that the safety of cell therapies, in general, has not yet been fully 373 evaluated. Notably, no RCT in the present review showed adverse events for the use of 374 dento-periodontal derived stem cells. Furthermore, questions such as cell delivery, immu-375 nogenicity, use of autologous or allogeneic cells, culture quality control, and cost-effec-376 tiveness are critical to address. The next phase of research should aim to identify the tis-377 sues that can optimally serve as the source of stem cells and, in this sense, future attention 378 should be even more marked on dental and periodontal tissues. It should be emphasized 379 that until recently, medical stem cell research has not prioritized periodontal tissues due 380 to the non-life-threatening nature of periodontitis. 381

Although efforts were made to enhance the quality of data regarding the topic, this study has certain limitations primarily stemming from the nature of the existing literature. Indeed, the RCTs provided periodontal regeneration with very diverse flap design, scaffolds, MSCs vectors and cell handling technologies. Despite the promising outcomes highlighted in the broader literature, our synthesis points to a nuanced perspective, suggesting that the use of orally-derived MSCs may not consistently confer additional clinical 382 benefits in the specified timeframe. While recognizing the potential of stem cell therapies, 388 including MSCs, our findings underscore the complexity of translating these approaches 389 into consistently successful clinical outcomes in the context of periodontitis. 390

5. Conclusions

In conclusion, it was not possible to demonstrate that the additional use of dento/per-392 iodontal stem cells in periodontal regenerative surgical procedures determines an im-393 provement in clinical and radiographic parameters compared to other biomaterials or 394 techniques more studied in the literature. The regenerative approach supported by tissue 395 engineering and cell therapy should be explored in depth with a significantly higher num-396 ber of randomized controlled clinical trials, with larger samples and at least 12 months 397 follow-up to allow the detection of even long-term outcomes. In consideration of the re-398 sults expressed, the low number of RCTs, the inherent costs of using MSCs and the possi-399 bility of adverse events still too little addressed in the literature, regenerative periodontal 400 surgery with the use of stem cells for bone defects could not be currently considered a preferential approach for clinical treatment compared to other periodontal regeneration 402 procedures. 403

6. Indications for future research

- RCTs evaluating the clinical efficacy, as well as patient related outcomes and 405 cost-benefit analyses of periodontal regeneration using dento-periodontal 406 stem cells. 407
- This RCTs should be designed with an increased number of patients enrolled 408 and long-term follow-up. 409
- Studies focused on clinical protocols to obtain an efficient number of MSCs 410 from the oral cavity. 411
- Studies focused on side effects in short-term and long-term with the use of 412 MSCs. 413

List of abbreviations: MD (mean difference); CAL (clinical attachment leve); PPD (probing pocket 414depth); GR (gingival recession); RGB (radiographic bone gain); CI (confidence interval); BoP (bleed-415 ing on probing); FI (furcation involvement); MSCs (mesenchymal stem cells); β -TCP (beta-tricalcium 416 phosphate); PRP (platelet-rich plasma); RCTs (randomized clinical trials); BMMSC (bone marrow 417 mesenchymal stem cells); ESC (embryonic stem cells); iPSC (induced pluripotent stem cells); 418 PDLSCs (periodontal ligament stem cells); GTR (guided tissue regeneration); DPSCs (dental pulp 419 stem cells); ABMMSCs (alveolar bone marrow mesenchymal stem cells); GMSCs (gingival mesen-420 chymal stem cells); MIST (minimally invasive surgical technique); BDD (linear distance from ce-421 mentoenamel junction to bottom of defect); BC-BD (linear distance from bone crest to bottom of 422 defect); OFD (open flap debridement). 423

Supplementary Materials: The following supporting information can be downloaded at: 424 www.mdpi.com/xxx/s1, Table S1: Included studies: summary of the results for the main outcomes 425 of interest. 426

Author Contributions: Conceptualization, A.C., M.P. and G.O.; X.X. and Y.Y.; methodology, A.C., 427 G.B., M.P., F.R. and G.O.; validation, A.C. and M.P.; formal analysis, A.C., G.B. and F.R.; investiga-428 tion, A.C., M.P. and F.A.; resources, X.X.; data curation, F.M.; writing-original draft preparation, 429 A.C., M.P., F.A.; writing-review and editing, G.B., F.R., F.M., M.A. and G.O.; supervision, M.A. 430 and G.O.; project administration, F.M. and G.O. All authors have read and agreed to the published 431 version of the manuscript. 432

Funding: This research received no external funding. 433 434

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All data used in this study are available in the manuscript or its sup-436 porting material. 437

391

401

404

Conflicts of Interest: The authors declare no conflicts of interest.

References

- Papapanou, P.N.; Sanz, M.; Buduneli, N.; Dietrich, T.; Feres, M.; Fine, D.H.; Flemmig, T.F.; Garcia, R.; Giannobile, 1. W.V.; Graziani, F.; et al. Periodontitis: Consensus Report of Workgroup 2 of the 2017 World Workshop on the 441 Classification of Periodontal and Peri-Implant Diseases and Conditions. Journal of Periodontology 2018, 89, S173-442 S182, doi:10.1002/JPER.17-0721. 443
- Pihlstrom, B.L.; Michalowicz, B.S.; Johnson, N.W. Periodontal Diseases. Lancet 2005, 366, 1809-1820, 2. 444 doi:10.1016/S0140-6736(05)67728-8. 445
- Morales, A.; Strauss, F.J.; Hämmerle, C.H.F.; Romandini, M.; Cavalla, F.; Baeza, M.; Sanz, M.; Gamonal, J. 3. 446 Performance of the 2017 AAP/EFP Case Definition Compared with the CDC/AAP Definition in Population-Based 447Studies. Journal of Periodontology 2022, 93, 1003-1013, doi:10.1002/JPER.21-0276. 448
- Loos, B.G.; Needleman, I. Endpoints of Active Periodontal Therapy. Journal of Clinical Periodontology 2020, 47, 61-4. 449 71, doi:10.1111/jcpe.13253. 450
- 5. Matuliene, G.; Pjetursson, B.E.; Salvi, G.E.; Schmidlin, K.; Brägger, U.; Zwahlen, M.; Lang, N.P. Influence of 451 Residual Pockets on Progression of Periodontitis and Tooth Loss: Results after 11 Years of Maintenance. J Clin 452 Periodontol 2008, 35, 685-695, doi:10.1111/j.1600-051X.2008.01245.x. 453
- Marruganti, C.; Romandini, M.; Gaeta, C.; Cagidiaco, E.F.; Discepoli, N.; Parrini, S.; Graziani, F.; Grandini, S. 6. 454 Healthy Lifestyles Are Associated with a Better Response to Periodontal Therapy: A Prospective Cohort Study. J 455 Clin Periodontol 2023, doi:10.1111/jcpe.13813. 456
- Ferrarotti, F.; Baima, G.; Rendinelli, M.; Citterio, F.; Mariani, G.M.; Mussano, F.; Romano, F.; Romandini, M.; 7. 457 Aimetti, M. Pocket Closure after Repeated Subgingival Instrumentation: A Stress Test to the EFP Guideline for 458 Stage III-IV Periodontitis. Clin Oral Investig 2023, doi:10.1007/s00784-023-05279-6. 459
- Suvan, J.; Leira, Y.; Moreno Sancho, F.M.; Graziani, F.; Derks, J.; Tomasi, C. Subgingival Instrumentation for 8. 460 Treatment of Periodontitis. A Systematic Review. J Clinic Periodontology 2020, 47, 155–175, doi:10.1111/jcpe.13245. 461
- 9. Sanz, M.; Herrera, D.; Kebschull, M.; Chapple, I.; Jepsen, S.; Berglundh, T.; Sculean, A.; Tonetti, M.S. Treatment of 462 Stage I-III Periodontitis-The EFP S3 Level Clinical Practice Guideline. Journal of Clinical Periodontology 2020, 47, 463 4-60, doi:https://doi.org/10.1111/jcpe.13290. 464
- Nibali, L.; Koidou, V.P.; Nieri, M.; Barbato, L.; Pagliaro, U.; Cairo, F. Regenerative Surgery versus Access Flap for 10. 465 the Treatment of Intra-Bony Periodontal Defects: A Systematic Review and Meta-Analysis. Journal of Clinical 466 Periodontology 2020, 47, 320-351, doi:10.1111/jcpe.13237. 467
- Aimetti, M.; Baima, G.; Aliyeva, N.; Lorenzetti, V.; Citterio, F.; Franco, F.; Di Scipio, F.; Berta, G.N.; Romano, F. 11. 468 Influence of Locally Delivered Doxycycline on the Clinical and Molecular Inflammatory Status of Intrabony 469 Defects Prior to Periodontal Regeneration: A Double-Blind Randomized Controlled Trial. Journal of Periodontal 470 Research 2023, 58, 1096–1104, doi:10.1111/jre.13174. 471
- Sculean, A.; Nikolidakis, D.; Nikou, G.; Ivanovic, A.; Chapple, I.L.C.; Stavropoulos, A. Biomaterials for Promoting 12. 472 Periodontal Regeneration in Human Intrabony Defects: A Systematic Review. Periodontal 2000 2015, 68, 182-216, 473 doi:10.1111/prd.12086. 474
- 13. Aimetti, M.; Fratini, A.; Manavella, V.; Giraudi, M.; Citterio, F.; Ferrarotti, F.; Mariani, G.M.; Cairo, F.; Baima, G.; 475 Romano, F. Pocket Resolution in Regenerative Treatment of Intrabony Defects with Papilla Preservation 476 Techniques: A Systematic Review and Meta-Analysis of Randomized Clinical Trials. J Clin Periodontol 2021, 48, 477 843-858, doi:10.1111/jcpe.13428. 478

438

- Calzavara, D.; Morante, S.; Sanz, J.; Noguerol, F.; Gonzalez, J.; Romandini, M.; Sanz, M. The Apically Incised
 Coronally Advanced Surgical Technique (AICAST) for Periodontal Regeneration in Isolated Defects: A Case Series.
 Quintessence Int 2021, 53, 24–34, doi:10.3290/j.qi.b1763645.
- Cortellini, P.; Tonetti, M.S. Clinical and Radiographic Outcomes of the Modified Minimally Invasive Surgical
 Technique with and without Regenerative Materials: A Randomized-Controlled Trial in Intra-Bony Defects. *J Clin Periodontol* 2011, 38, 365–373, doi:10.1111/j.1600-051X.2011.01705.x.
- Cortellini, P.; Tonetti, M.S. Improved Wound Stability with a Modified Minimally Invasive Surgical Technique in the Regenerative Treatment of Isolated Interdental Intrabony Defects. J Clin Periodontol 2009, 36, 157–163, 486 doi:10.1111/j.1600-051X.2008.01352.x.
- Nibali, L.; Sultan, D.; Arena, C.; Pelekos, G.; Lin, G.-H.; Tonetti, M. Periodontal Infrabony Defects: Systematic
 Review of Healing by Defect Morphology Following Regenerative Surgery. *Journal of Clinical Periodontology* 2021,
 489
 48, 101–114, doi:10.1111/jcpe.13381.
- Nibali, L.; Mehta, J.; Al-Shemeri, D.; Anoixiadou, S.; Parashis, A.; Vouros, I. Association between Defect 491 Morphology and Healing of Intrabony Defects Treated with Minimally Invasive Non-Surgical Therapy: A Pilot 492 Exploratory Analysis of Two Cohorts. *J Periodontal Res* 2023, *58*, 708–714, doi:10.1111/jre.13119.
- Chen, F.-M.; Gao, L.-N.; Tian, B.-M.; Zhang, X.-Y.; Zhang, Y.-J.; Dong, G.-Y.; Lu, H.; Chu, Q.; Xu, J.; Yu, Y.; et al.
 Treatment of Periodontal Intrabony Defects Using Autologous Periodontal Ligament Stem Cells: A Randomized
 Clinical Trial. *Stem Cell Res Ther* 2016, 7, 33, doi:10.1186/s13287-016-0288-1.
- 20. Kanemoto, Y.; Miyaji, H.; Nishida, E.; Miyata, S.; Mayumi, K.; Yoshino, Y.; Kato, A.; Sugaya, T.; Akasaka, T.;
 497 Nathanael, A.J.; et al. Periodontal Tissue Engineering Using an Apatite/Collagen Scaffold Obtained by a Plasma498 and Precursor-Assisted Biomimetic Process. *J Periodontal Res* 2022, *57*, 205–218, doi:10.1111/jre.12954.
 499
- Bianchi, S.; Bernardi, S.; Simeone, D.; Torge, D.; Macchiarelli, G.; Marchetti, E. Proliferation and Morphological 500 Assessment of Human Periodontal Ligament Fibroblast towards Bovine Pericardium Membranes: An In Vitro 501 Study. *Materials* 2022, 15, 8284, doi:10.3390/ma15238284. 502
- Shi, Y.; Cao, J.; Wang, Y. Rethinking Regeneration: Empowerment of Stem Cells by Inflammation. *Cell Death Differ* 503
 2015, 22, 1891–1892, doi:10.1038/cdd.2015.127.
- Gao, P.; Kajiya, M.; Motoike, S.; Ikeya, M.; Yang, J. Application of Mesenchymal Stem/Stromal Cells in Periodontal 505 Regeneration: Opportunities and Challenges. *Jpn Dent Sci Rev* 2024, 60, 95–108, doi:10.1016/j.jdsr.2024.01.001. 506
- Fernandes, S.; Chong, J.J.H.; Paige, S.L.; Iwata, M.; Torok-Storb, B.; Keller, G.; Reinecke, H.; Murry, C.E. 507
 Comparison of Human Embryonic Stem Cell-Derived Cardiomyocytes, Cardiovascular Progenitors, and Bone 508
 Marrow Mononuclear Cells for Cardiac Repair. *Stem Cell Reports* 2015, *5*, 753–762, doi:10.1016/j.stemcr.2015.09.011. 509
- Nakamura, K.; Murry, C.E. Function Follows Form A Review of Cardiac Cell Therapy. *Circ J* 2019, *83*, 2399– 510
 2412, doi:10.1253/circj.CJ-19-0567. 511
- Kaigler, D.; Pagni, G.; Park, C.H.; Braun, T.M.; Holman, L.A.; Yi, E.; Tarle, S.A.; Bartel, R.L.; Giannobile, W.V. Stem
 Cell Therapy for Craniofacial Bone Regeneration: A Randomized, Controlled Feasibility Trial. *Cell Transplant* 2013, 22, 767–777, doi:10.3727/096368912X652968.
- Hoang, D.M.; Pham, P.T.; Bach, T.Q.; Ngo, A.T.L.; Nguyen, Q.T.; Phan, T.T.K.; Nguyen, G.H.; Le, P.T.T.; Hoang, 515
 V.T.; Forsyth, N.R.; et al. Stem Cell-Based Therapy for Human Diseases. *Sig Transduct Target Ther* 2022, 7, 1–41, 516
 doi:10.1038/s41392-022-01134-4. 517
- Han, P.; Johnson, N.; Abdal-Hay, A.; Moran, C.S.; Salomon, C.; Ivanovski, S. Effects of Periodontal Cells-Derived 518 Extracellular Vesicles on Mesenchymal Stromal Cell Function. J Periodontal Res 2023, 58, 1188–1200, 519 doi:10.1111/jre.13171. 520

- Shimizu, Y.; Takeda-Kawaguchi, T.; Kuroda, I.; Hotta, Y.; Kawasaki, H.; Hariyama, T.; Shibata, T.; Akao, Y.;
 Kunisada, T.; Tatsumi, J.; et al. Exosomes from Dental Pulp Cells Attenuate Bone Loss in Mouse Experimental
 Periodontitis. J Periodontal Res 2022, 57, 162–172, doi:10.1111/jre.12949.
- Dominici, M.; Le Blanc, K.; Mueller, I.; Slaper-Cortenbach, I.; Marini, F.; Krause, D.; Deans, R.; Keating, A.; Prockop,
 D.; Horwitz, E. Minimal Criteria for Defining Multipotent Mesenchymal Stromal Cells. The International Society
 for Cellular Therapy Position Statement. *Cytotherapy* 2006, *8*, 315–317, doi:10.1080/14653240600855905.
- Zeng, Y.; Deng, J.-J.; Jiang, Q.-L.; Wang, C.-L.; Zhang, L.; Li, T.; Jiang, J. Thyrotropin Inhibits Osteogenic 527 Differentiation of Human Periodontal Ligament Stem Cells. J Periodontal Res 2023, 58, 668–678, 528 doi:10.1111/jre.13109. 529
- Yu, B.; Li, Q.; Sun, X.; Yin, Y. O-GlcNAcylation of TLR4 Inhibits Osteogenic Differentiation of Periodontal 530
 Ligament Stem Cells. J Periodontal Res 2024, 59, 119–127, doi:10.1111/jre.13193.
- Gronthos, S.; Mankani, M.; Brahim, J.; Robey, P.G.; Shi, S. Postnatal Human Dental Pulp Stem Cells (DPSCs) in
 Vitro and in Vivo. *Proc Natl Acad Sci U S A* 2000, *97*, 13625–13630, doi:10.1073/pnas.240309797.
- Miura, M.; Gronthos, S.; Zhao, M.; Lu, B.; Fisher, L.W.; Robey, P.G.; Shi, S. SHED: Stem Cells from Human 534 Exfoliated Deciduous Teeth. *Proc Natl Acad Sci U S A* 2003, 100, 5807–5812, doi:10.1073/pnas.0937635100.
- Seo, B.-M.; Miura, M.; Gronthos, S.; Bartold, P.M.; Batouli, S.; Brahim, J.; Young, M.; Robey, P.G.; Wang, C.-Y.; Shi,
 S. Investigation of Multipotent Postnatal Stem Cells from Human Periodontal Ligament. *Lancet* 2004, 364, 149–155,
 doi:10.1016/S0140-6736(04)16627-0.
- Roato, I.; Baima, G.; Orrico, C.; Mosca Balma, A.; Alotto, D.; Romano, F.; Ferracini, R.; Aimetti, M.; Mussano, F.
 Senescent Markers Expressed by Periodontal Ligament-Derived Stem Cells (PDLSCs) Harvested from Patients
 with Periodontitis Can Be Rejuvenated by RG108. *Biomedicines* 2023, 11, 2535, doi:10.3390/biomedicines11092535.
- Melcher, A.H. On the Repair Potential of Periodontal Tissues. J Periodontol 1976, 47, 256–260, 542 doi:10.1902/jop.1976.47.5.256.
 543
- 38. Karring, T.; Nyman, S.; Lindhe, J. Healing Following Implantation of Periodontitis Affected Roots into Bone Tissue.
 544 *J Clin Periodontol* 1980, 7, 96–105, doi:10.1111/j.1600-051x.1980.tb01952.x.
 545
- Huang, G.T.-J.; Gronthos, S.; Shi, S. Mesenchymal Stem Cells Derived from Dental Tissues vs. Those from Other 546 Sources: Their Biology and Role in Regenerative Medicine. J Dent Res 2009, 88, 792–806, 547 doi:10.1177/0022034509340867. 548
- Li, Z.; Wu, Z.; Xi, X.; Zhao, F.; Liu, H.; Liu, D. Cellular Communication Network Factor 1 Interlinks Autophagy and ERK Signaling to Promote Osteogenesis of Periodontal Ligament Stem Cells. J Periodontal Res 2022, 57, 1169– 1182, doi:10.1111/jre.13054.
- Iwata, T.; Kaneda-Ikeda, E.; Takahashi, K.; Takeda, K.; Nagahara, T.; Kajiya, M.; Sasaki, S.; Ishida, S.; Yoshioka,
 M.; Matsuda, S.; et al. Regulation of Osteogenesis in Bone Marrow-Derived Mesenchymal Stem Cells via Histone
 Deacetylase 1 and 2 Co-Cultured with Human Gingival Fibroblasts and Periodontal Ligament Cells. *J Periodontal Res* 2023, *58*, 83–96, doi:10.1111/jre.13070.
- Chauca-Bajaña, L.; Velasquez-Ron, B.; Tomás-Carmona, I.; Camacho-Alonso, F.; Pérez-Jardón, A.; Pérez-Sayáns, 556
 M. Regeneration of Periodontal Bone Defects with Mesenchymal Stem Cells in Animal Models. Systematic Review 557
 and Meta-Analysis. Odontology 2023, 111, 105–122, doi:10.1007/s10266-022-00725-5. 558
- 43. Nagahara, T.; Yoshimatsu, S.; Shiba, H.; Kawaguchi, H.; Takeda, K.; Iwata, T.; Mizuno, N.; Fujita, T.; Kurihara, H.
 559 Introduction of a Mixture of β-Tricalcium Phosphate into a Complex of Bone Marrow Mesenchymal Stem Cells
 560 and Type I Collagen Can Augment the Volume of Alveolar Bone without Impairing Cementum Regeneration. *J*561 *Periodontol* 2015, *86*, 456–464, doi:10.1902/jop.2014.140384.
 562

- Wang, M.; He, M.; Xu, X.; Wu, Z.; Tao, J.; Yin, F.; Luo, K.; Jiang, J. Cementum Protein 1 Gene-Modified Adipose-Derived Mesenchymal Stem Cell Sheets Enhance Periodontal Regeneration in Osteoporosis Rat. *J Periodontal Res* 564 2023, 58, 755–768, doi:10.1111/jre.13133.
- Wang, Q.; Yang, X.; Wang, X.; Wang, X.; Zhang, J.; Gao, Y.; Pan, J.; Wang, S. Identifying Genes for Regulating 566
 Osteogenic Differentiation of Human Periodontal Ligament Stem Cells in Inflammatory Environments by 567
 Bioinformatics Analysis. J Periodontal Res 2023, doi:10.1111/jre.13215. 568
- Sun, L.; Du, X.; Kuang, H.; Sun, H.; Luo, W.; Yang, C. Stem Cell-Based Therapy in Periodontal Regeneration: A 569 Systematic Review and Meta-Analysis of Clinical Studies. *BMC Oral Health* 2023, 23, 492, doi:10.1186/s12903-023- 570 03186-6. 571
- Zhang, Y.; Zhao, W.; Jia, L.; Xu, N.; Xiao, Y.; Li, Q. THE APPLICATION OF STEM CELLS IN TISSUE
 ENGINEERING FOR THE REGENERATION OF PERIODONTAL DEFECTS IN RANDOMIZED CONTROLLED
 TRIAL: A SYSTEMATIC REVIEW AND META-ANALYSIS. J Evid Based Dent Pract 2022, 22, 101713, 574
 doi:10.1016/j.jebdp.2022.101713.
- Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G.; PRISMA Group Preferred Reporting Items for Systematic 576 Reviews and Meta-Analyses: The PRISMA Statement. J Clin Epidemiol 2009, 62, 1006–1012, 577 doi:10.1016/j.jclinepi.2009.06.005. 578
- 49. Higgins, J.P.T.; Altman, D.G.; Gøtzsche, P.C.; Jüni, P.; Moher, D.; Oxman, A.D.; Savovic, J.; Schulz, K.F.; Weeks, L.; 579
 Sterne, J.A.C.; et al. The Cochrane Collaboration's Tool for Assessing Risk of Bias in Randomised Trials. *BMJ* 2011, 580
 343, d5928, doi:10.1136/bmj.d5928. 581
- 50. Baima, G.; Citterio, F.; Romandini, M.; Romano, F.; Mariani, G.M.; Buduneli, N.; Aimetti, M. Surface
 582 Decontamination Protocols for Surgical Treatment of Peri-Implantitis: A Systematic Review with Meta-Analysis.
 583 Clin Oral Implants Res 2022, 33, 1069–1086, doi:10.1111/clr.13992.
- Romano, F.; Perotto, S.; Baima, G.; Macrì, G.; Picollo, F.; Romandini, M.; Mariani, G.M.; Aimetti, M. Estimates and Multivariable Risk Assessment of Mid-Buccal Gingival Recessions in an Italian Adult Population According to the 2018 World Workshop Classification System. *Clin Oral Investig* 2022, *26*, 4769–4780, doi:10.1007/s00784-022-04441-w.
- Hernández-Monjaraz, B.; Santiago-Osorio, E.; Ledesma-Martínez, E.; Aguiñiga-Sánchez, I.; Sosa-Hernández, N.A.;
 Mendoza-Núñez, V.M. Dental Pulp Mesenchymal Stem Cells as a Treatment for Periodontal Disease in Older
 Adults. Stem Cells Int 2020, 2020, 8890873, doi:10.1155/2020/8890873.
- 53. Shalini, H.S.; Vandana, K.L. Direct Application of Autologous Periodontal Ligament Stem Cell Niche in Treatment
 592 of Periodontal Osseous Defects: A Randomized Controlled Trial. J Indian Soc Periodontol 2018, 22, 503–512,
 593 doi:10.4103/jisp.jisp_92_18.
- Abdal-Wahab, M.; Abdel Ghaffar, K.A.; Ezzatt, O.M.; Hassan, A.A.A.; El Ansary, M.M.S.; Gamal, A.Y.
 Regenerative Potential of Cultured Gingival Fibroblasts in Treatment of Periodontal Intrabony Defects
 (Randomized Clinical and Biochemical Trial). J Periodontal Res 2020, 55, 441–452, doi:10.1111/jre.12728.
- Apatzidou, D.A.; Bakopoulou, A.A.; Kouzi-Koliakou, K.; Karagiannis, V.; Konstantinidis, A. A Tissue-Engineered
 Biocomplex for Periodontal Reconstruction. A Proof-of-Principle Randomized Clinical Study. *J Clin Periodontol* 2021, 48, 1111–1125, doi:10.1111/jcpe.13474.
- 56. Ferrarotti, F.; Romano, F.; Gamba, M.N.; Quirico, A.; Giraudi, M.; Audagna, M.; Aimetti, M. Human Intrabony
 601
 Defect Regeneration with Micrografts Containing Dental Pulp Stem Cells: A Randomized Controlled Clinical Trial.
 602
 J Clin Periodontol 2018, 45, 841–850, doi:10.1111/jcpe.12931.
 603

.. ..

. .

.....

. . .

people or property resulting from any ideas, methods, instructions or products referred to in the content.

o / 1

57.	Sanchez, N.; Fierravanti, L.; Nunez, J.; Vignoletti, F.; Gonzalez-Zamora, M.; Santamaria, S.; Suarez-Sancho, S.;	604
	Fernández-Santos, M.E.; Figuero, E.; Herrera, D.; et al. Periodontal Regeneration Using a Xenogeneic Bone	605
	Substitute Seeded with Autologous Periodontal Ligament-Derived Mesenchymal Stem Cells: A 12-Month Quasi-	606
	Randomized Controlled Pilot Clinical Trial. J Clin Periodontol 2020, 47, 1391–1402, doi:10.1111/jcpe.13368.	607
58.	Mancini, L.; Romandini, M.; Fratini, A.; Americo, L.M.; Panda, S.; Marchetti, E. Biomaterials for Periodontal and	608
	Peri-Implant Regeneration. Materials 2021, 14, 3319, doi:10.3390/ma14123319.	609
59.	Graziano, A.; d'Aquino, R.; Laino, G.; Papaccio, G. Dental Pulp Stem Cells: A Promising Tool for Bone	610
	Regeneration. Stem Cell Rev 2008, 4, 21–26, doi:10.1007/s12015-008-9013-5.	611
60.	Monti, M.; Graziano, A.; Rizzo, S.; Perotti, C.; Del Fante, C.; d'Aquino, R.; Redi, C.A.; Rodriguez y Baena, R. In	612
	Vitro and In Vivo Differentiation of Progenitor Stem Cells Obtained After Mechanical Digestion of Human Dental	613
	Pulp. Journal of Cellular Physiology 2017, 232, 548–555, doi:10.1002/jcp.25452.	614
61.	Zhu, W.; Liang, M. Periodontal Ligament Stem Cells: Current Status, Concerns, and Future Prospects. Stem Cells	615
	International 2015 , 2015, e972313, doi:10.1155/2015/972313.	616
62.	Roguljic, H.; Matthews, B.G.; Yang, W.; Cvija, H.; Mina, M.; Kalajzic, I. In Vivo Identification of Periodontal	617
	Progenitor Cells. J Dent Res 2013, 92, 709-715, doi:10.1177/0022034513493434.	618
63.	Zakrzewski, W.; Dobrzyński, M.; Szymonowicz, M.; Rybak, Z. Stem Cells: Past, Present, and Future. Stem Cell Res	619
	<i>Ther</i> 2019 , <i>10</i> , 68, doi:10.1186/s13287-019-1165-5.	620
64.	Mason, S.; Tarle, S.A.; Osibin, W.; Kinfu, Y.; Kaigler, D. Standardization and Safety of Alveolar Bone-Derived	621
	Stem Cell Isolation. J Dent Res 2014, 93, 55–61, doi:10.1177/0022034513510530.	622
65.	Cao, C.; Tarlé, S.; Kaigler, D. Characterization of the Immunomodulatory Properties of Alveolar Bone-Derived	623
	Mesenchymal Stem Cells. Stem Cell Research & Therapy 2020, 11, 102, doi:10.1186/s13287-020-01605-x.	624
66.	Lemaitre, M.; Monsarrat, P.; Blasco-Baque, V.; Loubières, P.; Burcelin, R.; Casteilla, L.; Planat-Bénard, V.; Kémoun,	625
	P. Periodontal Tissue Regeneration Using Syngeneic Adipose-Derived Stromal Cells in a Mouse Model. Stem Cells	626
	<i>Transl Med</i> 2017 , <i>6</i> , 656–665, doi:10.5966/sctm.2016-0028.	627
67.	Nguyen-Thi, TD.; Nguyen-Huynh, BH.; Vo-Hoang, TT.; Nguyen-Thanh, T. Stem Cell Therapies for	628
	Periodontal Tissue Regeneration: A Meta-Analysis of Clinical Trials. Journal of Oral Biology and Craniofacial Research	629
	2023 , 13, 589–597, doi:10.1016/j.jobcr.2023.07.001.	630
68.	Chen, M.X.; Zhong, Y.J.; Dong, Q.Q.; Wong, H.M.; Wen, Y.F. Global, Regional, and National Burden of Severe	631
	Periodontitis, 1990-2019: An Analysis of the Global Burden of Disease Study 2019. J Clin Periodontol 2021, 48, 1165-	632
	1188, doi:10.1111/jcpe.13506.	633
		634
	Claimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual au-	635
thor((s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to	636

_

1.1