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**The *cardiac caval index*. Improving non-invasive assessment of
cardiac preload**

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Running title

The Cardiac Caval Index

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25 **Abstract**

26 **Objectives** - Inferior Vena Cava (IVC) pulsatility quantified by the Caval Index (CI) is
27 characterized by poor reliability, also due to the irregular magnitude of spontaneous respiratory
28 activity generating the major pulsatile component. The aim of this study was to test whether
29 the IVC cardiac oscillatory component could provide a more stable index (Cardiac CI - CCI)
30 compared to CI or respiratory CI (RCI).

31 **Methods** - Nine healthy volunteers underwent long-term monitoring in supine position of IVC,
32 followed by 3 min Passive Leg Raising (PLR). CI, RCI and CCI were extracted from video
33 recordings by automated edge-tracking and CCI was averaged over each respiratory cycle
34 (aCCI). Cardiac Output (CO), Mean Arterial Pressure (MAP) and Heart Rate (HR) were also
35 recorded during baseline (1 min prior to PLR) and PLR (first minute).

36 **Results** - In response to PLR, all IVC indices decreased ($p < 0.01$), CO increased by $4 \pm 4\%$
37 ($p = 0.055$) while HR and MAP did not vary. The Coefficient of Variation (CoV) of aCCI
38 ($13 \pm 5\%$) was lower than that of CI ($17 \pm 5\%$, $p < 0.01$), RCI ($26 \pm 7\%$, $p < 0.001$) and CCI ($25 \pm 7\%$,
39 $p < 0.001$). The mutual correlations in time of the indices were 0.81 (CI-RCI), 0.49 (CI-aCCI)
40 and 0.2 (RCI-aCCI).

41 **Conclusions** - Long-term IVC monitoring by automated edge-tracking allowed us to evidence
42 that 1) respiratory and average cardiac pulsatility components are uncorrelated and thus carry
43 different information and 2) the new index aCCI, exhibiting the lowest CoV while maintaining
44 good sensitivity to blood volume changes, may overcome the poor reliability of CI and RCI.

45

46 **Key Words:** inferior vena cava; passive leg raising; volume status; fluid responsiveness;
47 automatic edge tracking.

48

49 **Introduction**

50 In the clinical setting, deciding whether and what amount of fluid to administer
51 intravenously to a patient, i.e., the prediction of fluid responsiveness, is a long-standing open
52 issue, whose relevance is paramount. Indeed, it has been shown that only half of the
53 haemodynamic unstable patients exhibits a positive outcome after a fluid challenge ¹, while the
54 remaining ones are exposed to the risk of fluid overload ²⁻⁴. Since no satisfactory solution to
55 this problem has been found yet, improvements in the existing techniques as well as new
56 methodological approaches are constantly investigated ⁵⁻⁹.

57 Due to its fast and non-invasive approach, the echographic assessment of the Inferior Vena
58 Cava (IVC) pulsatility is a widely adopted monitoring technique ⁷. From the analysis of
59 pulsatility, it possible to infer about the mechanical characteristics of blood vessels, such as
60 stiffness and compliance, and about their determinants, such as blood pressure, blood volume,
61 vessel tone, etc. (Mesin et al., *submitted*). IVC pulsatility is often quantified by means of the
62 Caval Index (CI), which conveniently normalizes the respirophasic diameter variation (d_{\max} -
63 d_{\min}) to d_{\max} , thus accounting for individual differences in IVC size (d_{\max} and d_{\min} being the
64 maximum and minimum diameters, as measured at the end of the expiratory and inspiratory
65 phases, respectively). However, this index suffers of a large variability and, consequently, of
66 poor reliability ¹⁰⁻¹². Based on the development of new image processing algorithms, several
67 sources of variability were recently investigated and compensated for, e.g., by tracking the
68 displacement of the vessel with respect to the ultrasound (US) probe ^{13,14} and by averaging the
69 measurements over several IVC diameters in either short ¹⁵ or long axis ^{14,16}, which contributed
70 to improve the repeatability of the measurements ^{15,16}. However, a major source of variability
71 in the respirophasic oscillation of IVC size is the intrinsic variability of spontaneous respiratory
72 activity, in terms of magnitude, frequency and relative extent of thoracic/diaphragmatic

73 respiration^{17,18}, all these aspects providing consistent effects on IVC pulsatility¹⁸⁻²¹. A possible
74 solution to this problem was originally suggested by Nakamura²² who proposed to consider
75 the cardiac component of IVC pulsatility rather than the respiratory. The issue was followed-
76 up in few subsequent studies^{15,16,19,23}. In these studies, the automated analysis of US video
77 clips yielded the continuous description of IVC size changes with high time resolution (equal
78 to the frame rate of the US video recording), so that the cardiac and respiratory components of
79 IVC pulsatility could be easily separated, based on their different frequency contents, and
80 independently analysed^{15,16,23}. On this basis, the respiratory CI (RCI) and the cardiac CI (CCI),
81 specifically quantifying the respiratory and cardiac component of IVC pulsatility, were
82 introduced and compared. The results showed that also the CCI could be used as an index of
83 vascular filling^{22,23} and that it was characterized by a lower variability (as quantified by the
84 coefficient of variation, CoV) than the RCI, although, contrary to the expectation, not lower
85 than the variability of the CI¹⁶. Two reasons may possibly explain this observation: 1) the
86 above mentioned results refer to index variability over different subjects and measurement
87 sessions, while the actual CCI variability over time has never been assessed; 2) the cardiac
88 pulsatility may still be affected by a respiratory modulation, as apparent from published
89 recordings^{16,24} and confirmed in preliminary observations. However, to our knowledge, all
90 studies generally considered only short time intervals lasting 10-15 s, and the time correlation
91 between pulsatile components has never been investigated.

92 We hypothesized that 1) by further improving the signal processing we could effectively
93 reduce the respiratory modulation of CCI and obtain a more stable hemodynamic index of
94 vascular filling; 2) due to its different nature, the CCI could differ from RCI and CI in terms of
95 time course and responsiveness to fluid challenges.

96 To this purpose, continuous and long-duration recordings of B-mode IVC imaging were
97 performed with the help of a dedicated probe holder, in resting conditions and during a
98 simulated fluid challenge, as produced by passive leg raising (PLR).

99 **Materials and Methods**

100 *Subjects*

101 Nine healthy volunteers (7 M, 2 F, age 34 ± 9) were included in the study, with the only
102 exclusion criteria being a poor quality of the echographic imaging. The study was approved by
103 the Ethics Committee of the University of Torino (March 23, 2015) and all participants gave
104 their informed consent according to the principles of the Helsinki Declaration.

105 *Experimental set-up and protocol*

106 Participants remained supine on a clinical bed for at least 30 min before starting the
107 experiment, in order to stabilize the equilibrium between fluid compartments^{19,25}. A two-
108 dimensional B-mode longitudinal view of the IVC was recorded by means of a MyLab 25 Gold
109 system (ESAOTE, Genova, Italy) equipped with a convex 2-5 MHz US probe, according to a
110 *subxiphoid* approach²⁶. To achieve long-lasting ultrasound (US) monitoring, we made use of
111 a probe holder, as successfully implemented in previous studies for stable echo-Doppler
112 monitoring of arteries and veins of upper and lower limbs^{9,27-30}. In the present case, the probe
113 holder was stemming from one side of the bed and its 40-cm long horizontal arm was allowed
114 to freely rotate about a joint at one of its ends. The US probe, located at the other end, due to
115 its own weight, could then exert a light pressure on the abdomen and maintain adequate
116 acoustic contact, accommodating with virtually vertical displacements the small abdominal

117 movements during respiration. This arrangement allowed us to continuously monitor the IVC
118 for the whole duration of the protocol (4 min).

119 The experimental protocol consisted of 1 min of rest in supine position (baseline), followed
120 by 3 min during which the legs were passively raised and maintained at about 45 deg (passive
121 leg raising, PLR) and 1 min of rest, again in supine position. During the entire protocol, the US
122 video of the IVC longitudinal section (in the sagittal plane) was recorded for the subsequent
123 processing and analysis. In addition, Heart Rate (HR), Mean Arterial Pressure (MAP) and
124 Cardiac Output (CO) were non-invasively monitored by photoplethysmography (CNAP®,
125 CNSystems Medizintechnik, Graz, Austria) while breathing was monitored by means of a
126 custom-made strain gauge band placed around the chest (the recorded signal is referred to as
127 Breath in the following). All these signals were digitally and synchronously recorded by a
128 general-purpose acquisition board (Micro 1401 IImk, CED, Cambridge, UK, with Spike2
129 software): IVC videos were acquired at about 30 fps while HR, MAP, CO and breathing were
130 sampled at 10 Hz.

131 *IVC segmentation*

132 US videos were processed by a custom-made software (implemented in MATLAB 2020a,
133 The MathWorks, Natick, MA) for IVC edge-tracking. The routines were based on a previously
134 developed algorithm¹⁴. The tracking algorithm was improved to attenuate the effect of small
135 drifts, which would produce detrimental effects with videos of long duration considered here
136 (manuscript in preparation). The edges of the IVC were estimated as previously described¹⁴,
137 by sampling along 21 directions crossing the blood vessel, considering a portion selected by an
138 operator (PPO), who was blinded to the results. Along each direction, the software estimated
139 the US pixel intensity by interpolation. Then, abrupt variations of this estimated US intensity
140 were identified as the locations of the two IVC edges along the considered direction (see Mesin

141 et al. ¹⁴ for additional details). The length of the segment between each couple of points placed
142 on the upper and lower vein edges was the IVC diameter along that direction.

143 The median axis of the vein was estimated (as the mean of the two sampled edges),
144 interpolated by a second order polynomial and used to rotate the 21 diameters mentioned above
145 to be orthogonal to it. By considering all frames of the US video, each diameter was a time
146 series. High frequency contributions in these time series of diameters (mostly related to
147 superimposed noise) were removed. For the identification of the cut-off frequency, the power
148 spectrum density (PSD) of the diameters was first computed (Burg method, with order 40 ³¹),
149 from which the highest frequency of our interest was identified as follows. First, we have
150 searched for a peak in the PSD between 40bpm and 120bpm, which reflected the cardiac
151 component. Then, the median (across diameters) of peak frequencies (mf) was computed (this
152 parameter was used later to define the cut-off frequency of the filter). Then, a portion of 15 mm
153 around the position of the diameter showing the highest peak of the cardiac component was
154 selected (assuming that such a diameter provided reliable information on the cardiac
155 contribution and that it was less affected by noise than the other diameters). Upper and lower
156 border points of this portion of the vein were then interpolated with two straight lines. Finally,
157 the *mean IVC diameter*, for each frame, was calculated as the area of the IVC section
158 considered above, divided by its length (i.e., 15 mm).

159 Such a mean diameter was low pass filtered, with cut-off frequency equal to $mf + 0.5Hz$
160 (Chebyshev of type I, stop band starting at $mf + 1.5Hz$, minimum attenuation of 30 dB,
161 passband from 0 to $mf + 0.5Hz$ with ripple of 0.5 dB) and indicated with *dIVC*.

162 The respiratory and cardiac components of IVC pulsatility were estimated from the mean
163 diameter just obtained. The respiratory diameter, indicated as *R-dIVC*, was estimated by the
164 first step of the Empirical Mode Decomposition applied to the mean diameter. Specifically,
165 two curves were first obtained by interpolating the local maxima and the local minima of *dIVC*.

166 The curve *R-IVC* was defined as the mean of these two curves. Notice that this technique allows
167 to estimate each respiration cycle. On the other hand, a filter with fixed passband was used in
168 previous works ¹⁶: such a filter had lower performances than the one used here, especially with
169 our long recordings, in which respiration cycles could have very different durations (thus, being
170 attenuated differently by a fixed filter). The cardiac diameter, called *C-dIVC*, was computed as
171 $C-dIVC = dIVC - R-dIVC + s-dIVC$ and is equivalent to the mean diameter deprived of the
172 respiratory oscillations. The term *s-dIVC* indicates the lowpass filtered mean diameter with cut-
173 off 0.05 Hz (Chebyshev filter of type I, stop band starting at 0.5 Hz, minimum attenuation of
174 30 dB, passband ripple of 0.5 dB), where only the *slow* sub respiratory frequencies are left.
175 This low pass filter was chosen in order to remove any oscillation and keep only the low
176 frequency trend reflecting slow IVC size variations induced by the PLR.

177 At this point of the analysis, the three diameters, *dIVC*, *R-dIVC* and *C-dIVC* were available
178 as time series (see Fig. 1) and were used to estimate the collapsibility indicators Caval Index
179 (CI), Respiratory Caval Index (RCI) and Cardiac Caval Index (CCI) respectively, according to
180 the usual formula: $(d_{\max} - d_{\min})/d_{\max}$ (Fig. 1, bottom). Note that, while for CI and RCI one
181 estimate per respiratory cycle is obtained, the CCI yields one estimate per cardiac cycle.

182 In addition, an *averaged* version of the CCI, aCCI, was computed by averaging the CCI
183 over distinct respiratory cycles. The aCCI estimates could then be considered synchronous with
184 CI and RCI (one estimate per respiratory cycle).

185 *Data analysis*

186 HR, MAP and CO were exported from Spike2 software to MATLAB® (version 2020b) for
187 off-line analysis. As a first step, they were aligned in time with the time-series of the IVC
188 collapsibility indexes, computed separately as explained above, that presented a non-uniform
189 sampling rate due to their nature. Indeed, CI, RCI and aCCI had one sample per respiratory

190 cycle while CCI one per heartbeat: the sample location in time, within the respiratory cycle,
191 was arbitrary and we chose to be at the minimum of the IVC diameter component for all the
192 three indexes.

193 The intra-subject variability in time of each IVC collapsibility index was quantified during
194 baseline by the coefficient of variation ($\text{CoV} = (\text{STD} / \text{MEAN}) \times 100$) and averaged across all
195 subjects. The correlation of time course in baseline was tested among CI, RCI and aCCI, for
196 each subject, using the Pearson correlation coefficient (ρ); then, the mean ρ across subjects was
197 computed by averaging the individual ρ values after a Fisher Z-transformation and
198 subsequently applying an inverse transformation to the result.

199 In order to perform the correlation of IVC indices with other signals, they were resampled
200 at 10 Hz, after a shape-preserving piecewise cubic interpolation. This was necessary to test the
201 correlation of CCI and aCCI with the respiratory pattern. However, since the delay between
202 the respiratory effort and the resulting changes in size of the IVC cannot be assumed constant
203 neither across subject nor over time, the normalized cross-correlation function on the
204 appropriately standardized signals, instead of Pearson correlation coefficient, was employed
205 and its maximum value, irrespective of the delay, was chosen as the correlation coefficient (ρ).
206 Then, the ρ values obtained were averaged across subjects using the Fisher transformation as
207 explained above.

208 The response to the PLR is known to take place within the first minute after raising the legs
209 ³² and, for each of the variables, it was assessed as the difference between the mean value
210 calculated during the first minute of PLR and during the whole baseline (1 min), as $\text{DELTA} =$
211 $\text{PLR} - \text{baseline}$, and considered in both absolute and relative (percentage) terms. For both basal
212 and DELTA values, mutual correlations among CI, RCI and aCCI were quantified by the
213 Pearson correlation coefficient, presented along with the 95% confidence interval in between
214 brackets. The effect of PLR on each variable was assessed considering the distribution of

215 DELTA values and testing if the mean differed from zero with a level of significance set at
216 0.05, by means of the Wilcoxon signed rank test. The same test was used to compare the
217 variability in time (as expressed by the CoV) of aCCI with CI and RCI. The IVC indices
218 accuracy in predicting the subject response to the simulated fluid challenge, as induced by PLR,
219 was analysed by means of Receiver Operating Characteristic (ROC) curves built using a 10%
220 increase in CO as marker of fluid responsiveness^{6,33}.

221 Finally, it is worth to mention that, given the non-uniform sampling rate for the IVC
222 collapsibility indexes, their average time course, across subjects, was obtained by averaging
223 the interpolated curves.

224 **Results**

225 *Basal conditions*

226 An example of the original tracings from a representative subject is shown in Figure 1
227 which includes the continuous recordings of some systemic variables like HR, MAP, and
228 respiratory activity as well as variables extracted from the US monitoring of the IVC, i.e., the
229 IVC diameter (average diameter of the considered IVC segment), the respiratory diameter (high
230 frequency components are filtered out) and the cardiac diameter (the respiratory component is
231 filtered out). At the bottom of the figure are the different indices, automatically calculated. Two
232 aspects need to be observed.

233 1) A strong correspondence exists between the magnitude of respiratory acts and the
234 respiratory changes in IVC diameter. Accordingly, CI and RCI (bottom) are also
235 modulated by the depth of respiration, in particular, it is worth to notice how CI and
236 RCI drop (the variation is in the order of 10%) around the fourth second of the

237 recording, concomitantly with a reduced inspiratory depth, as revealed by the Breath
238 signal.

239 2) Even the cardiac pulsatility is modulated by the respiratory activity. Accordingly, such
240 modulation is preserved in CCI and affects its variability in time.

241 We first tested whether the averaging over single respiratory cycles, as implemented for
242 the calculation of aCCI, was effective in reducing the respiratory modulation affecting CCI. In
243 Figure 2, the distributions of the maxima of the cross-correlation in time between Breath and
244 the two indexes, CCI and aCCI, are shown. Note that the averaging introduced in aCCI has
245 drastically reduced the correlation with respiration from 0.4 (for CCI) to 0.02 (for aCCI).

246 We then tested whether this new feature was effective in reducing the overall variability in
247 time, as assessed by the CoV. In Figure 3 it is possible to observe the distribution of the
248 individual CoVs, depicted by means of box and whiskers plots, for each IVC collapsibility
249 index, including the original CCI. The MEAN \pm STD CoV values of CI, RCI, CCI and aCCI
250 are, respectively, $17 \pm 5 \%$, $26 \pm 7 \%$, $25 \pm 7 \%$, $13 \pm 5 \%$. As expected, aCCI exhibited a lower
251 variability than CCI. The improved stability over time can also be observed by comparing the
252 corresponding tracings in Figure 1. Moreover, aCCI also achieved a lower CoV than CI
253 ($p < 0.01$).

254 *Response to PLR*

255 On a different timescale, the full representation of the response to PLR is shown in Figure
256 4. It can be observed that during PLR, starting at time 0 s, both cardiac and respiratory pulsatile
257 components are reduced, and that the IVC diameter is increased. This results in a reduction of
258 all indices during PLR, as displayed at the bottom.

259 In Figure 5, the averaged (across subjects) time course of HR, MAP and CO are presented
260 in terms of percentage changes with respect to the mean baseline value (i.e., DELTA in

261 percentage terms). It can be observed that at the beginning of PLR (time 0 s), HR and MAP
262 exhibit only small fluctuations while CO immediately begins to rise reaching a peak at around
263 30 s and returning to the basal value at around 60 s, before the end of PLR (180 s).

264 Figure 6 shows, on the same timescale, the time course of IVC collapsibility indexes,
265 namely CI, RCI and aCCI (with CCI superimposed as dashed line). Here the variations with
266 respect to the mean baseline values are not translated in percentage terms, since the indexes are
267 already expressed as percentages, so that absolute variations (of a percentage) are considered
268 (i.e., DELTA in absolute terms). It can be observed that all indices exhibit a consistent decrease,
269 which is maintained throughout PLR, and that aCCI exhibited a sharper decrease at the
270 beginning of PLR, compared to CI and RCI.

271 In Table 1, the values averaged across subjects for baseline, PLR, and DELTA (in
272 percentage terms) for all the physiological variables of interest are listed, as well as the
273 statistical significance against the null hypothesis of no effects induced by PLR. As it can be
274 noticed, HR and MAP did not change following PLR while CO and all the IVC collapsibility
275 indexes did.

276 Finally, regarding the prediction of fluid responsiveness, both CI and aCCI performed as
277 perfect classifiers (i.e., AUCROC 1) with threshold of 21% and 9%, respectively, while RCI
278 reached a poorer performance (AUCROC 0.78).

279 *Correlations among indices*

280 In Figure 7, the distributions of the individual mutual correlations in time among the IVC
281 collapsibility indexes are shown (the original CCI is no longer considered): the biggest
282 correlation is between CI and RCI (mean value 0.81), while the smallest one is between RCI
283 and aCCI (mean value 0.2). It is worth to notice how the small interquartile range of CI-RCI
284 distribution, depicted in Figure 7, highlights the robustness of the link between the two indexes,

285 while the same cannot be asserted for the CI-aCCI correlation in time, despite the relatively
286 high mean value (0.49). Finally, the Pearson correlation coefficient among the averaged
287 baseline values of CI-RCI, CI-aCCI and RCI-aCCI was 0.92 (0.66, 0.98), 0.76 (0.19, 0.95) and
288 0.50 (-0.25, 0.87), respectively. The same coefficients for the DELTA values, following the
289 same order, were 0.74 (0.16, 0.94), 0.73 (0.13, 0.94), 0.2 (-0.53, 0.76).

290 **Discussion**

291 The present study allowed to confirm preliminary observations and to achieve new relevant
292 results, which can be synthetized as follows:

- 293 1) Although the CI is generally considered as an index of the respiratory-induced
294 pulsatility of the IVC, it is heavily affected (or disturbed) by a pulsatility of cardiac
295 nature.
- 296 2) The magnitude of the cardiac pulsatility of IVC is still modulated by the respiratory
297 activity, which negatively impacts on the reliability of the CCI.
- 298 3) Averaging the CCI over single respiratory cycles effectively eliminates the respiratory
299 modulation and improves its stability in time.
- 300 4) The averaged cardiac collapsibility index (aCCI) responsiveness to PLR is uncorrelated
301 to the respiratory collapsibility index (RCI), suggesting that the two indices may carry
302 different information.

303 To our knowledge, this is the first study reporting a long-term monitoring and analysis of
304 IVC pulsatility, which was achieved thanks to a newly devised experimental set-up and
305 consolidated image processing algorithms^{13,14,16,24,34}. With this approach, a continuous time
306 series of the average IVC diameter, with high time resolution, could be analysed along with
307 other physiological variables: such an analysis included the identification of the oscillatory

308 components of the IVC diameter of respiratory and cardiac origin and the automated
309 calculation of the corresponding collapsibility indices RCI and CCI (Fig. 1) ¹⁶.

310 *The pivotal role of heart in Inferior Vena Cava respirophasic oscillations*

311 The aforementioned framework gave us the possibility to carefully observe the interplay
312 between respiration and heartbeat in generating the IVC pulsatility. Indeed, although IVC
313 pulsatility has been already the object of hundreds of studies ^{11,12} and its use in the clinical
314 settings, as predictor of fluid responsiveness ^{6,7} or as surrogate measure of central venous
315 pressure ³⁴, has been extensively investigated, only recently the cardiac component of the IVC
316 pulsatility has been described ²². This component was probably too weak or too fast to be
317 detected and disentangled from the slower respiratory component by means of just the visual
318 assessment and the standard tools available on US machines. On the one hand, these limitations
319 delayed the recognition and the investigation of the characteristics and meaning of the cardiac
320 component, on the other hand, the unrecognized cardiac oscillation, merging with the primary
321 respiratory oscillation, decreased the “signal-to-noise ratio” and increased the variability of the
322 oscillatory pattern. We speculate that this overlooked “disturbance” on the assessment of IVC
323 diameter may at least partly explain the poor reliability and clinical applicability of the CI ^{11,12}.
324 Notably, in the present study, removal of the cardiac pulsatility reduced the IVC collapsibility
325 index by about 50% (i.e., RCI is about 50% of CI) and, accordingly, the aCCI approximately
326 accounts for the other 50% (see Table 1, baseline). These results challenge the concept that the
327 classical IVC CI quantifies the “respirophasic” changes in IVC diameter.

328 As shown in the representative recordings of Figure 1, as well as in other figures previously
329 published ^{15,16}, the cardiac pulsatility is modulated by respiration: the magnitude of the
330 oscillation increases at low IVC diameter, which occurs approximately at the end of the
331 inspiratory phase (maximum lung volume). This modulatory pattern fits with the idea that the

332 pulse pressure, mainly provided by the atrial contraction, results in a lower volume increase
333 (reflected by a lower IVC diameter increase) when the IVC compliance is lower, which occurs
334 at larger IVC size (Mesin et al, *submitted*). Surprisingly an opposite pattern is shown in Figure
335 4 of the study from Sonoo et al ²³, i.e., wider cardiac pulsatility during expiration compared to
336 inspiration. However, their average findings (collected from 142 patients enrolled in an
337 emergency department) confirms a higher CCI during inspiration (13.8%) compared to
338 expiration (11.0%). This modulatory action is responsible for the high CoV of the CCI in time,
339 similar to the CoV of CI (Fig. 3) ¹⁶ and negatively impacts on its potential clinical usefulness.
340 By simply averaging over single respiratory cycles (aCCI), this problem was effectively
341 addressed and the CoV in time considerably reduced.

342 As discussed above, cardiac pulsatility is larger when the vessel size is smaller and vessel
343 compliance is larger. As such, aCCI candidates as a possible indicator of IVC compliance and
344 of poor vascular filling. In this respect, it is interesting to observe that it was shown to correlate
345 to CI, both in time (intra-subject) and across different subjects (in basal conditions). Moreover,
346 it was significantly affected by PLR (-28%, $p < 0.01$).

347 On the other hand, while a similar performance was reported by RCI (good correlation with
348 CI and significant decrease during PLR), aCCI and RCI were very poorly correlated: their
349 spontaneous oscillations in resting conditions are uncorrelated, their absolute values assessed
350 in resting conditions are uncorrelated, their responses to a (simulated) fluid challenge are
351 uncorrelated. These results strongly suggest that RCI and CCI are carrier of different
352 information (although both sensitive to fluid challenges). Their different time course in the
353 response to PLR (Fig. 6, aCCI exhibiting a faster and sharper response than RCI) further
354 supports this proposition.

355 *Clinical implications*

356 While further studies are necessary to understand the distinct physio-pathological meaning
357 of the two indices and their possible integration for clinical purposes, the possibility to get
358 increased and more reliable information from the same fast and non-invasive US examination
359 is intriguing. To date, only few studies have included a cardiac IVC collapsibility index in their
360 outcomes. In particular, the presence of CCI enhanced the capacity to predict the volume status
361 ²⁴ and right atrial pressure in patients ³⁴. However, no one has yet investigated the potential of
362 CCI in predicting a fluid challenge. For this reason, although we were curious to perform such
363 an investigation, we are aware that, given our small dataset (N=9) and the limitations of the
364 photoplethysmographic finger-cuff pulse contour analysis techniques in reliably monitoring
365 CO ³⁵, the extrapolated ROC analysis presented in this work are not relevant for a valid fluid
366 responsiveness study. Beyond that, we believe that the present findings, although obtained on
367 healthy volunteers, can add new useful information to the widespread use of the IVC
368 collapsibility indexes in predicting the fluid responsiveness in patients. Finally, the possibility
369 that CCI could be influenced by additional factors related to pathological alterations of cardiac
370 function, e.g., contractility, stiffness, valvular efficiency, etc., as well as changes in intra-
371 abdominal pressure, remains to be explored and deserves further studies on specific patient
372 populations.

373 *Physiologic response to Passive Leg Raising*

374 A final comment concerns the general response to PLR in this group of healthy subjects,
375 which provides a nice description of the physiological adaptation of the body to the new
376 situation (Figs. 5 and 6): no apparent effect on MAP, only a minor (pre-) activation of HR,
377 probably an increase in alertness due to the passive leg movement, along with a small but
378 visible transient increase in CO ³². In comparison, all the IVC indices detect a net variation

379 during PLR. Interestingly the exhibited changes are not transient, but last throughout the 3-min
380 duration of the test, which likely indicates that this time is too short for adjustments in blood
381 volume. Moreover, they show that the duration of the transients is shorter at the onset (< 15 s)
382 than at the termination of PLR (about 1 min).

383 *Limitations*

384 One limitation of the study is related to the way IVC videos were acquired, i.e., with the
385 probe held in place by a probe holder rather than by the hand of the echographer. While this
386 was a necessary implementation to achieve stable recordings lasting several minutes, it is not
387 without drawbacks, as involuntary spontaneous movements of the subject as well as
388 movements resulting from the PLR manoeuvre could occasionally interrupt the correct IVC
389 tracking. However, thanks to the prompt intervention of the operators, the proper probe
390 orientation was generally restored within seconds with no impact on the subsequent analysis.
391 Prospectively, with the increasing adoption of 4D US machines, the edge-tracking will be likely
392 extended to 3D images, which will minimize misalignment problems related to latero-lateral
393 displacement of IVC with respect to both hand-held and fixed US probe.

394 Secondly, the experiment was performed only on healthy volunteers posing some
395 limitations to the extrapolation of the results to the clinical setting. Moreover, we had to exclude
396 subjects that could not present good quality imaging of the IVC, as required by the image
397 processing routines. This criterion slightly biased the recruited sample towards a prevalence of
398 males, possibly due to their lower thickness of abdominal adipose tissue layer. However, we
399 are not aware of sex-related differences in IVC indices that could have affected the general
400 validity of the present results. Unfortunately, this is a known limitation of US studies that
401 require high quality imaging.

402 *Conclusions*

403 With this methodological study on healthy subjects, we evidenced that through echographic
404 long-term monitoring of the IVC longitudinal section, in association with an automated edge
405 tracking software, it is possible to record the IVC diameter and distinct respiratory and cardiac
406 collapsibility indexes as continuous time-series. A newly defined *averaged cardiac*
407 *collapsibility index*, aCCI, exhibited 1) the lowest variability in time, 2) good sensitivity to
408 simulated blood volume changes, as induced by PLR and 3) poor correlation with the RCI in
409 time, among subjects, and in their response to PLR, supporting the hypothesis that they carry
410 different information. Therefore, we believe that aCCI has the potential to overcome the poor
411 reliability of the classical CI in the fluid responsiveness prediction. Further studies in patients
412 are needed to understand its specificity and explore its applicability in the clinical practice.

413

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416 and by Proof of Concept “Vein Image Processing for Edge Rendering – VIPER”, supported by
417 the Italian Ministry of Economic Development, CUP C16I20000080006.

418 **Conflict of Interest**

419 An instrument implementing the algorithm used in this report to automatically track IVC
420 edges and to extract the mean IVC diameter was patented by Politecnico of Torino and
421 University of Torino (WO 2018/134726).

422

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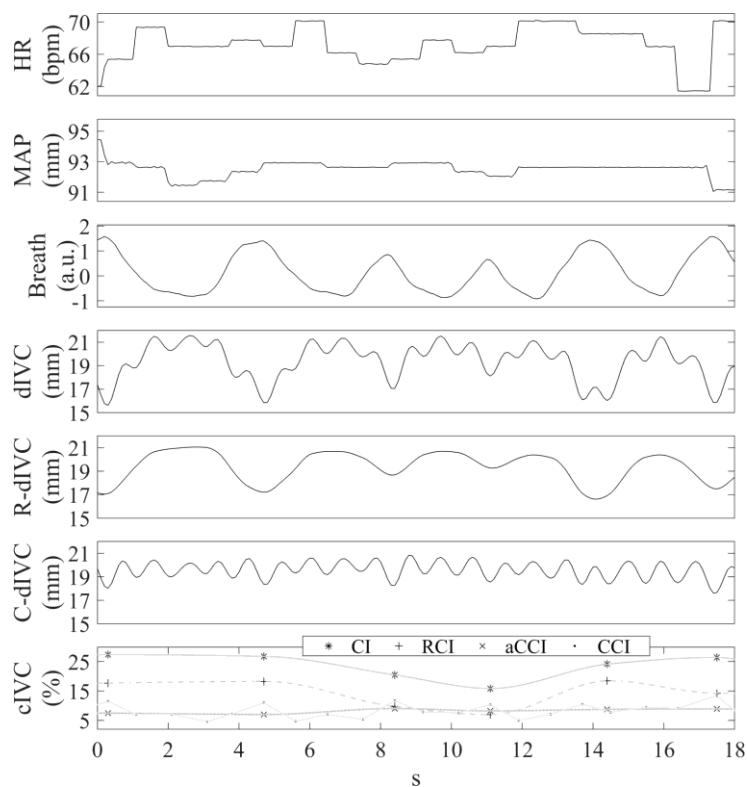
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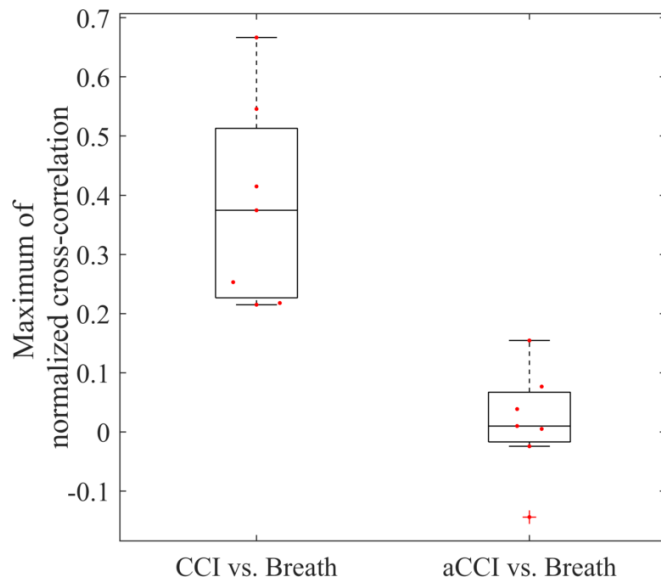
535 **Figures with captions**



536

537 **Fig. 1.** Tracings from a representative subject, in resting condition. The time course of each
538 one of the following variables is shown: Heart Rate (HR), Mean Arterial Pressure (MAP),
539 Respiration (Breath), Inferior Vena Cava (IVC) respiratory (R-dIVC) and cardiac (C-dIVC)
540 components of the native diameter trace (dIVC) and their respective indexes, namely Caval
541 Index (CI), Respiratory Caval Index (RCI) and averaged Cardiac Caval Index (aCCI). In the
542 latter graph, the markers indicate the exact sample of each IVC collapsibility indexes, as
543 described in the legend, while the continuous grey lines are the respective cubic interpolation
544 that were superimposed for a better visualization.

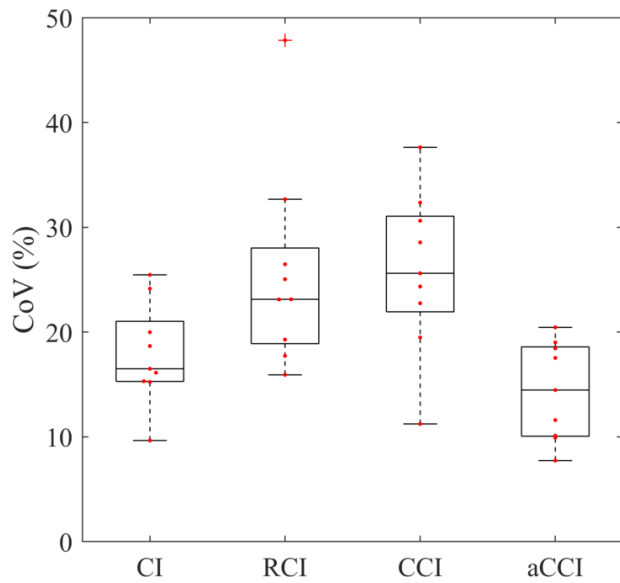
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546

547 **Fig. 2.** Correlation between respiration and inferior vena cava (IVC) cardiac collapsibility
 548 indexes. Distribution of the individual maximum value of cross-correlation among breathing
 549 signal and the two versions of the cardiac IVC collapsibility index (native and averaged, CCI
 550 and aCCI, respectively), during baseline.

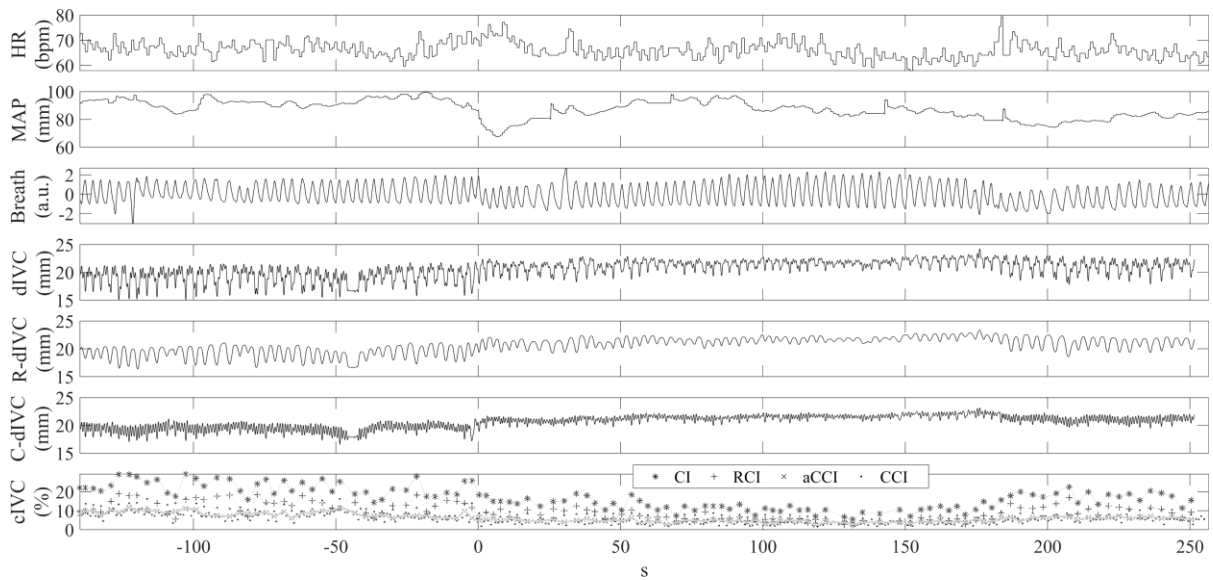
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553 **Fig. 3.** Coefficient of Variation (CoV) of collapsibility indexes of the inferior vena cava. The
 554 CoV distributions across subjects computed during baseline are shown for Caval Index (CI),
 555 Respiratory Caval Index (RCI), Cardiac Caval Index (CCI) and averaged Cardiac Caval Index
 556 (aCCI). The red dots indicate the individual data.

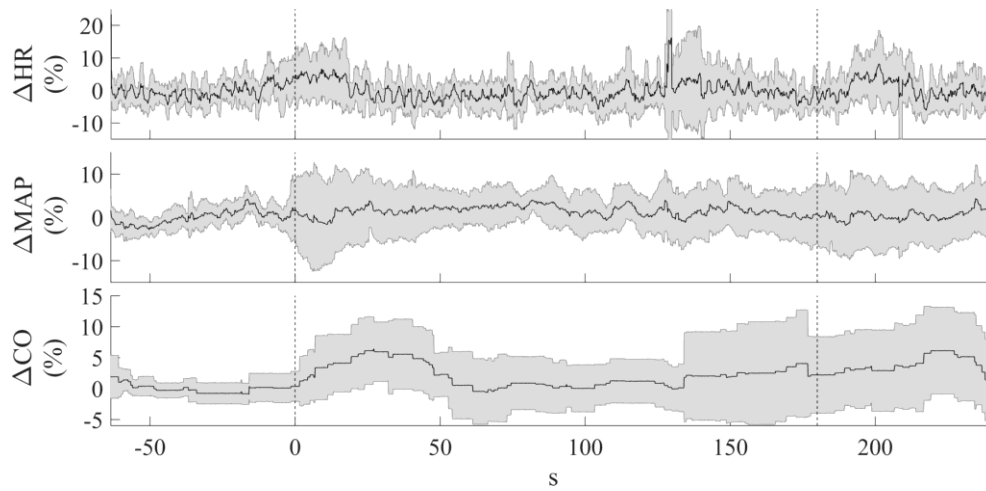
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559 **Fig. 4.** Example of a complete individual recording. The time course of each one of the
 560 following variables is shown: Heart Rate (HR), Mean Arterial Pressure (MAP), Respiration
 561 (Breath), Inferior Vena Cava (IVC) respiratory (R-dIVC) and cardiac (C-dIVC) components
 562 of the native diameter trace (dIVC) and their respective indexes, namely Caval Index (CI),
 563 Respiratory Caval Index (RCI) and averaged Cardiac Caval Index (aCCI). In the latter graph,
 564 the markers indicate the exact sample of each IVC collapsibility indexes, as described in the
 565 legend, while the continuous grey lines are the respective cubic interpolation that were
 566 superimposed for a better visualization.

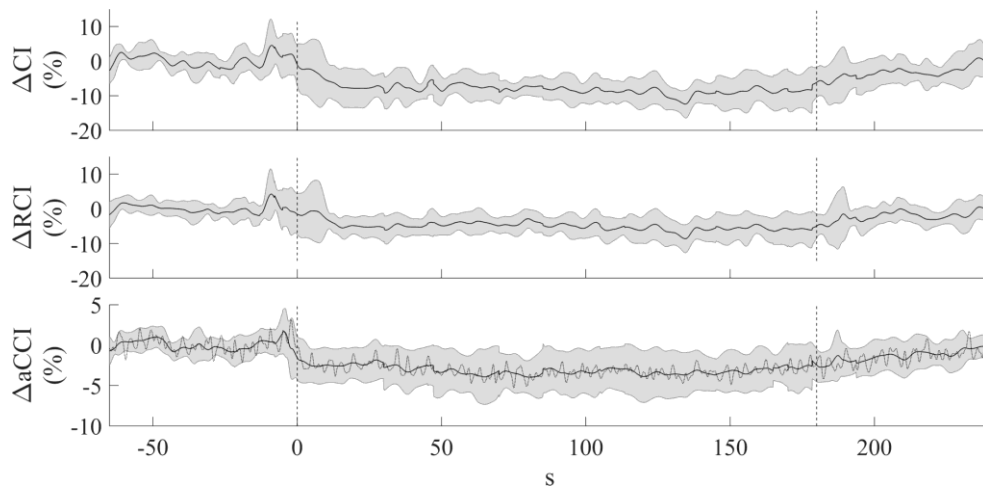
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568

569 **Fig. 5.** Haemodynamic variables averaged (across subjects) time course. Percentage changes
 570 with respect to the mean baseline value of Heart Rate (Δ HR), Mean Arterial Pressure (Δ MAP)
 571 and Cardiac Output (Δ CO): the black solid line represents the mean while the shaded grey error
 572 bar represent mean \pm std. The vertical dashed lines mark the beginning (left one) and the end
 573 (right one) of the PLR.

574



575

576 **Fig. 6.** Inferior Vena Cava collapsibility indexes averaged (across subjects) time course.

577 Absolute changes with respect to the mean baseline value of Caval Index (Δ CI), Respiratory

578 Caval Index (Δ RCI) and averaged Cardiac Caval Index (Δ aCCI): the black solid line represents

579 the mean while the shaded grey error bar represents mean \pm std. The latter graph presents also

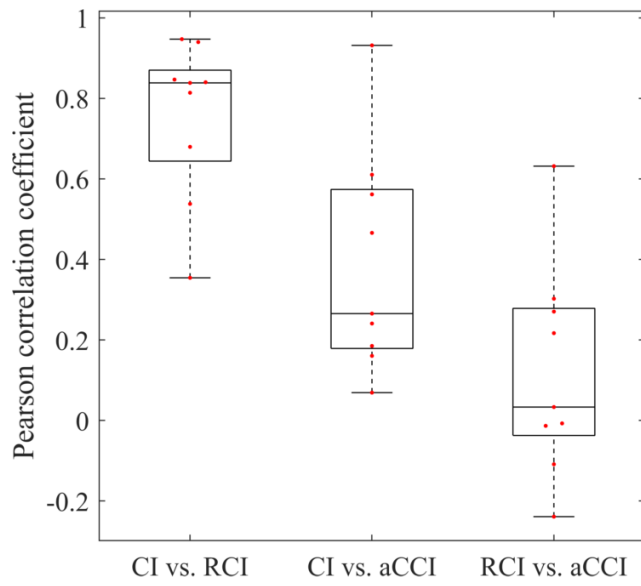
580 a superimposed dashed line trace that is the native Cardiac Caval Index: note the oscillations

581 due to the respiratory modulation of the cardiac induced pulsatility which are removed in the

582 aCCI trace (black solid line). The vertical dashed lines mark the beginning (left one) and the

583 end (right one) of the PLR.

584



585

586 **Fig. 7.** Inferior vena cava collapsibility indexes correlations. Distributions of individual mutual

587 correlations among inferior vena cava collapsibility indexes, computed during baseline.

588

589 **Tables and Appendices**

| Variable | Baseline | PLR | DELTA % | p-value |
|----------------------------------|-----------|-----------|----------|---------|
| Heart Rate (bpm) | 59 ± 9 | 59 ± 10 | 0 ± 3 | 0.82 |
| Mean Arterial Pressure (mmHg) | 89 ± 11 | 90 ± 9 | 1 ± 6 | 0.43 |
| Cardiac Output (L/min) | 5.0 ± 0.8 | 5.1 ± 0.7 | 4 ± 4 | 0.03 |
| Caval Index (%) | 27 ± 6 | 19 ± 7 | -31 ± 17 | 0.004 |
| Respiratory Caval Index (%) | 14 ± 4 | 9 ± 4 | -35 ± 17 | 0.004 |
| averaged Cardiac Caval Index (%) | 13 ± 4 | 9 ± 4 | -28 ± 21 | 0.008 |

590

591 **Table 1.** Averaged values of Heart Rate, Mean Arterial Pressure, Cardiac Output, Caval Index,
 592 Respiratory Caval Index, and averaged Cardiac Caval Index in absolute values, during baseline
 593 and PLR, and in terms of percentage variation during PLR w.r.t the mean baseline value
 594 (DELTA). Values are expressed as MEAN ± STD. Last column reports the p-value of the
 595 paired statistical comparison, by means of a paired Wilcoxon signed rank test, among the two
 596 distributions of PLR and baseline individually averaged values.